

# 1    **Response of Vegetable Crops to Heavy Metal Exposure in Contaminated**

## 2    **Irrigation Water and Its Implications for Food Safety**

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## 10    **Abstract**

11    Heavy metal contamination in irrigation water poses serious risks to crop productivity and food safety,  
12    particularly for leafy vegetables that readily accumulate toxic elements. This study evaluated the growth  
13    response, bioaccumulation behavior, and health risks of three commonly consumed leafy vegetables,  
14    velvetleaf (*Limnocharis flava*), kailan (*Brassica oleracea* var. *alboglabra*), and kangkong (*Ipomoea*  
15    *aquatica*), exposed to Pb at 50 mg L<sup>-1</sup>, Cd at 1 mg L<sup>-1</sup>, and Cu at 2 mg L<sup>-1</sup> under controlled irrigation  
16    conditions. Plant length, leaf number, and leaf area were monitored over an eighteen-day exposure  
17    period. Heavy metal concentrations in roots and leaves were quantified using ICP-OES with high  
18    analytical accuracy. Data were analyzed using analysis of variance, and means compared using the  
19    Duncan Multiple Range Test (DMRT). The results showed specific growth responses for each cultivar.  
20    Kailan exhibited the strongest stress symptoms, including a twenty-four percent reduction in plant  
21    length and a thirty-three percent decrease in leaf area under Cu exposure, while velvetleaf and kangkong  
22    showed comparatively higher tolerance. Bioconcentration factors exceeded one for all metals,  
23    indicating strong accumulation potential, particularly in kailan with Pb at 2873.21, Cd at 99.87, and Cu  
24    at 414.13. Translocation factors were consistently below one, indicating restricted metal movement  
25    from roots to leaves. Despite low translocation, Pb and Cd concentrations in edible tissues exceeded  
26    FAO and WHO safety limits, resulting in very high Hazard Index values, especially for kailan at 3097.6.  
27    These findings demonstrate that plant tolerance alone does not guarantee food safety. The integration  
28    of accurate analytical measurement, mathematical performance evaluation, and health risk assessment  
29    provides a robust framework for identifying safer crop choices and guiding irrigation management in  
30    heavy metal-contaminated environments.

31    **Keywords:** heavy metal stress, plant tolerance, bioaccumulation, food safety, polluted irrigation water.

## 32    **1. INTRODUCTION**

33    Heavy metal contamination in agricultural ecosystems is an increasingly significant  
34    global concern, particularly in peri-urban areas and wetlands exposed to industrial activities

35 and intensive agricultural practices with inadequate waste management (Kim et al., 2020). The  
36 scarcity of clean water resources has resulted in the use of low-quality water as an alternative  
37 irrigation source in agriculture (Dotaniya et al., 2023). Heavy metals in irrigation water can  
38 accumulate in crops, significantly increasing the risk of human exposure through dietary  
39 consumption (Bounar et al., 2020; Khan et al., 2023). These metals are toxic to plants, causing  
40 substantial physiological and morphological changes that adversely affect plant growth and the  
41 nutritional quality of vegetables (Noor et al., 2022; Rahman et al., 2022). Previous studies have  
42 shown that heavy metal contamination in irrigation water leads to reduced biomass, altered  
43 growth patterns, and disturbed plant development (Tekle et al., 2023; Waheed et al., 2022).  
44 The toxic effects of these metals are further exacerbated by the fact that many vegetables absorb  
45 and accumulate heavy metals in their edible parts, such as leaves and fruits, which increases  
46 the potential risks to human health.

47 Exposure to heavy metals such as chromium (Cr), manganese (Mn), iron (Fe), copper  
48 (Cu), nickel (Ni), zinc (Zn), cadmium (Cd), lead (Pb), and mercury (Hg) has been found in  
49 high concentrations in various vegetables sold in the market (Manwani, Vanisree, et al., 2022).  
50 Pb, Cd, and Cu are of particular concern which are commonly found in high concentrations in  
51 various vegetables, including Japanese spinach (*Spinacia oleracea*), tomatoes (*Solanum*  
52 *lycopersicum*), mustard and cabbage (*Brassica parachinensis L.*, *Brassica kompestris L.*,  
53 *Brassica oleracea L.*), lettuce (*Lactuca sativa*) (Abbas et al., 2023; Zhou et al., 2016), and  
54 spinach (*Amaranthus viridis*, *Amaranthus tricolour*, *Amaranthus paniculatus L.*) (Islam &  
55 Hoque, 2014). Pb, Cd, and Cu contamination in irrigation water affects different growth  
56 responses among plant species. Previous study indicates that plant species exhibit varying  
57 levels of tolerance to heavy metals (Singh et al., 2024a), with some plants demonstrating better  
58 adaptation and resilience than others. As the global demand for safe and nutritious vegetables  
59 increases, understanding the morphological responses of plants to heavy metals is crucial for  
60 developing sustainable agricultural practices.

61 Various scientific studies have investigated the morphological responses of vegetables  
62 to heavy metal contamination. Studies on spinach have shown that this plant is capable of  
63 accumulating high levels of Pb and Cd in its roots and leaves when irrigated with contaminated  
64 water (Bonanno & Cirelli, 2017; Ogunkunle et al., 2015). A study on cucumbers has shown  
65 significant changes in growth patterns, including reduced root and shoot biomass, indicating  
66 toxicity due to heavy metal exposure (Othman et al., 2021; Somda et al., 2019). In addition to  
67 growth inhibition, heavy metals often cause chlorosis in leaves, further exposing the noticeable  
68 morphological impact of contamination on irrigation water (Aksouh et al., 2024). Furthermore,

69 research indicates that certain vegetables, such as spinach, lettuce, and cabbage, show increased  
70 heavy metal concentrations when exposed to contaminated water and soil (Abbas et al., 2023).  
71 Understanding plant responses to heavy metal stress and their ability to accumulate and tolerate  
72 metals can enhance vegetable safety and mitigate risks associated with heavy metal  
73 contamination in vegetable cultivation.

74 Although previous studies have reported heavy-metal accumulation in various vegetables, the  
75 current literature still lacks comparative evidence on how different leafy species respond  
76 physiologically to specific metal contaminants supplied through irrigation water under  
77 controlled conditions. Most studies use soil-based exposure, focus on a single crop, or assess  
78 only one metal, which limits the understanding of species-specific tolerance, accumulation,  
79 and translocation pathways relevant to real irrigation risks. Moreover, few studies integrate  
80 morphological responses with quantitative bioaccumulation metrics and health risk assessment  
81 to determine the safe-use potential of vegetables grown under contaminated water.

82 The novelty of this study lies in its combined experimental and risk-based approach. First, the  
83 work provides a controlled, comparative assessment of three widely consumed leafy vegetables  
84 exposed to Pb, Cd, and Cu at environmentally relevant concentrations delivered through  
85 irrigation water. Second, the study quantifies both bioconcentration and translocation factors  
86 to identify species-specific metal movement from roots to edible leaves, which directly  
87 determines food safety. Third, the analysis links plant responses to a health risk framework  
88 based on Estimated Daily Intake, Hazard Quotient, and Hazard Index, allowing a direct  
89 interpretation of the implications for consumers. This integrated approach enables the  
90 identification of species with lower translocation potential and supports crop selection  
91 strategies for cultivation in areas where irrigation water may contain heavy metals.

## 92 **2. LITERATURE REVIEW**

93 Heavy metal contamination of irrigation water and soil has emerged as a critical environmental  
94 and food safety issue worldwide, particularly in regions where untreated or partially treated  
95 wastewater is reused for crop production. Such practices are common in peri-urban and water-  
96 scarce agricultural systems due to limited freshwater availability. Heavy metals such as Pb, Cd,  
97 and Cu are persistent, non-biodegradable contaminants that accumulate in soils and crops,  
98 posing significant risks to ecosystem health and human consumers. Studies have documented  
99 that continuous irrigation with contaminated water increases soil metal concentrations over  
100 time, subsequently enhancing the uptake and accumulation of metals in crops and increasing

101 the likelihood of entry into the food chain. This process remains a major concern for  
102 agricultural sustainability and public health globally (Khaliq et al., 2022).

103 Leafy vegetables are particularly vulnerable to heavy metal contamination due to their  
104 morphological and physiological characteristics, including shallow root systems and high  
105 transpiration rates, which facilitate efficient absorption of metals from soil and irrigation water.  
106 Research indicates that leafy vegetables such as spinach and okra often accumulate higher  
107 metal concentrations compared to other vegetables under similar exposure conditions, making  
108 them important indicators of environmental contamination (Khaliq et al., 2022). Metal uptake  
109 and accumulation in plants are influenced by several factors, including metal speciation, soil  
110 pH, organic matter content, and plant species traits. The extent of metal uptake and internal  
111 distribution can vary widely among species, with some metals like Cd and Pb being readily  
112 absorbed by roots and subsequently transported to aerial parts, while others may be more  
113 strongly retained in root tissue. Copper, although essential at trace levels, becomes toxic at  
114 higher concentrations, leading to physiological stress and reduced crop performance (Ali &  
115 Ahirwar, 2025).

116 To quantify metal accumulation in plants, researchers frequently use mathematical indicators  
117 such as the bioconcentration factor (BCF) and translocation factor (TF). The BCF represents  
118 the ratio of metal concentration in plant tissues to that in the soil or water medium, providing  
119 a measure of the plant's capacity to accumulate metals. The TF describes the efficiency of  
120 metal movement from roots to shoots or edible parts. High BCF values indicate strong  
121 accumulation potential, whereas TF values below one suggest limited internal metal mobility.  
122 Studies have shown that many leafy vegetables exhibit high BCF values but low TFs, indicating  
123 considerable metal retention in roots despite potential accumulation in edible tissues (Kaur et  
124 al., 2025).

125 Assessment of plant tolerance to heavy metal stress often involves morphological and  
126 physiological parameters such as plant height, leaf area, and biomass. Tolerant species are  
127 generally those capable of maintaining growth despite stress by activating detoxification  
128 mechanisms such as metal sequestration, synthesis of metal-binding proteins, and antioxidant  
129 defenses. However, tolerance in terms of growth does not necessarily correlate with safety for  
130 human consumption, as tolerant plants may still accumulate metals at concentrations exceeding  
131 food safety thresholds. This decoupling underscores the need for integrated evaluation  
132 frameworks that consider both plant performance and potential health risks (Ali & Ahirwar,

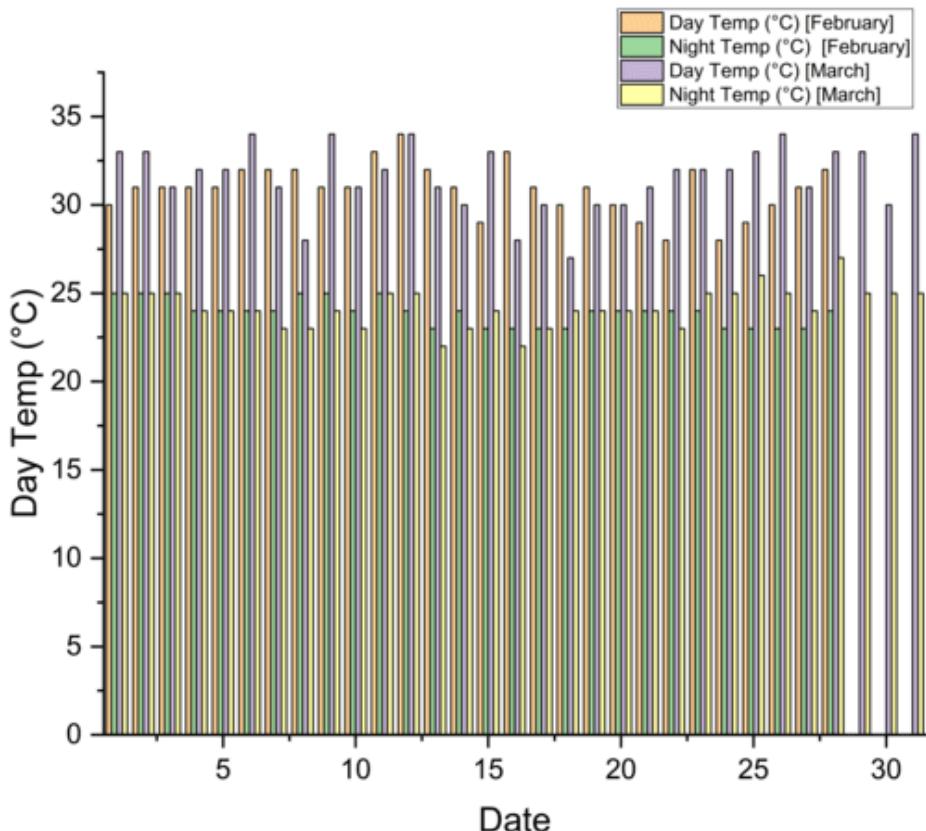
133 2025). From a human health perspective, consumption of vegetables contaminated with heavy  
134 metals represents a primary exposure route. Health risk assessment models, including  
135 Estimated Daily Intake (EDI), Hazard Quotient (HQ), and Hazard Index (HI), are widely used  
136 to evaluate potential non-carcinogenic risks associated with dietary exposure. HQ values  
137 greater than one indicate potential health concerns, and HI integrates multiple metal risks to  
138 reflect combined exposure. Studies consistently report that Pb and Cd contribute  
139 disproportionately to overall health risk due to their high toxicity and low tolerable intake  
140 limits, while children are found to be especially vulnerable due to higher intake relative to body  
141 mass (Ahmed et al., 2022).

142 Despite extensive research on heavy metal uptake and health risk assessment, the literature  
143 reveals a fragmented focus on isolated aspects, such as accumulation metrics or health  
144 implications alone, without an integrated performance evaluation framework. Many studies  
145 lack a unified methodology linking plant growth response, metal accumulation behavior,  
146 translocation patterns, and human health risk, which limits applicability for decision-making  
147 in agricultural management. Recent efforts have emphasized the integration of morphological  
148 measures with mathematical performance indices to provide a more comprehensive and  
149 reproducible evaluation of crop suitability under contaminated irrigation (Mafuyai & Ugbidye,  
150 2021).

### 151 **3. MATERIALS AND METHODS**

#### 152 **3.1 Study site, design, experimental unit, and replication**

153 The experiment was conducted at an open experimental site in Pematang Sulur village,  
154 Telanaipura District, Jambi Province, Indonesia ( $1^{\circ}36'25.6''$  S,  $103^{\circ}33'52.1''$  E). The site was  
155 selected because it is located in a peri-urban agricultural area with potential exposure to low-  
156 quality irrigation water. Daily meteorological data including air temperature and humidity were  
157 obtained from the nearest official station of Badan Meteorologi, Klimatologi, dan Geofisika  
158 (BMKG), located approximately 5 km from the experimental site. Sampling of plant tissues  
159 was conducted at harvest on day 18. Roots and leaves from each treatment pot were collected  
160 as composite samples to reduce variability between individual plants. The details of daily  
161 temperatures during the experimental can be seen in **Figure 1**.

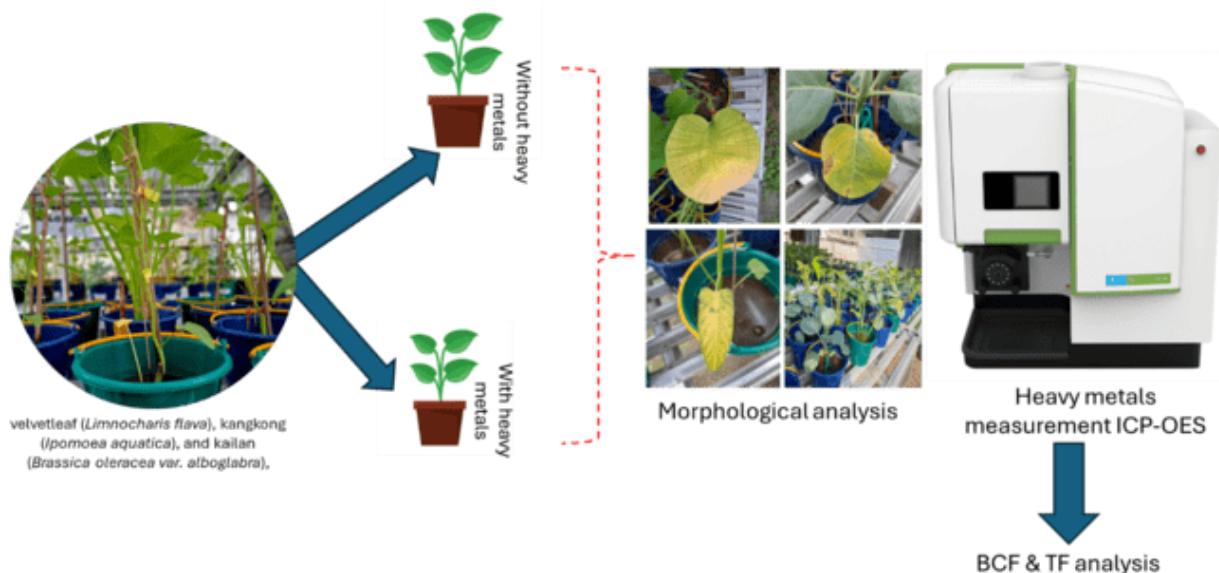


162

163 **Figure 1.** Daily temperature during the experimental period (February–March 2025) recorded  
 164 by the Badan Meteorologi, Klimatologi, dan Geofisika (BMKG) Station of Sultan Thaha  
 165 Airport, Jambi

166 The experimental used a Randomized Block Design (RDB) with three replications,  
 167 consisting of one factor, namely the combination of plant cultivars and type of heavy metal.  
 168 The plants used consist of three vegetables: velvetleaf (*Limnocharis flava*),  
 169 kangkong (*Ipomoea aquatica*), and kailan (*Brassica oleracea* var. *alboglabra*) and the heavy  
 170 metals: non-metals, Plumbum (Pb), Cadmium (Cd), and Copper (Cu), resulting in 12 treatment  
 171 combinations. Each treatment combination was repeated in three replications (three blocks).  
 172 There was a total of 36 treatment combinations, where each treatment unit consist of 4 plants,  
 173 with 2 plants as samples. The total number of plants in this study was 144. Pots (25 cm × 20  
 174 cm) were arranged under a transparent polyethylene shelter to prevent rainfall interference  
 175 while maintaining natural photoperiod and ventilation. Treatments were randomized within  
 176 blocks to minimize positional effects.

177



178

179 **Figure 2.** Experimental workflow of the study

180 **3.2 Experimental Setup and Instrumentation**

181 The experiment was conducted inside a transparent polyethylene shelter ( $6\text{ m} \times 3\text{ m} \times 2.5\text{ m}$ )  
 182 constructed to exclude rainfall while maintaining natural light and ventilation (**Figure 2**). The  
 183 shelter frame was built from galvanized steel and covered with 0.12 mm UV-stabilized  
 184 polyethylene film. Ambient temperature and humidity inside the shelter were monitored  
 185 continuously using a digital thermo-hygrometer (Testo 608-H2, Germany) placed at plant  
 186 canopy height ( $\approx 50\text{ cm}$ ) and recorded every 30 min.

187 The experimental bench was arranged in three blocks, each measuring  $1.8\text{ m} \times 0.9\text{ m}$ , separated  
 188 by 50 cm aisles to minimize edge effects and ensure uniform exposure. Within each block,  
 189 twelve  $25\text{ cm} \times 20\text{ cm}$  plastic pots were positioned on elevated trays (height = 60 cm). Pots  
 190 were randomly assigned to treatment combinations of plant species  $\times$  metal type. Each pot was  
 191 labeled with an alphanumeric code. Irrigation and metal dosing were performed manually using  
 192 graduated glass cylinders (500 mL, Pyrex) to ensure a precise daily application of 400 mL  
 193 solution per pot. Fresh working solutions were prepared every morning from analytical-grade  
 194 salts ( $\text{Pb}(\text{NO}_3)_2$ ,  $\text{Cd}(\text{NO}_3)_2$ , and  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ ) dissolved in distilled water and stored in amber  
 195 glass containers to prevent photodegradation.

196 Concentrations of Pb, Cd, and Cu in the working solutions were verified using Inductively  
 197 Coupled Plasma–Optical Emission Spectrometry (ICP-OES; PlasmaQuant 9100, Analytik  
 198 Jena, Germany). The instrument was calibrated daily using five-point external standards for  
 199 each metal (0.01, 0.1, 1, 5, and 10  $\text{mg L}^{-1}$ ) prepared from certified stock solutions (Merck,

200 Germany). Calibration curves were accepted when the coefficient of determination ( $r^2$ ) was  
201 greater than 0.999. Concentration drift was monitored by analyzing continuing calibration  
202 verification standards every ten samples. If the measured value deviated by more than  $\pm 5$   
203 percent of the expected value, the instrument was recalibrated and previous samples were re-  
204 run. All laboratory glassware was soaked in 10 percent  $\text{HNO}_3$  for 24 h, rinsed three times with  
205 deionized water, and dried in a clean cabinet before use. Method blanks and field blanks were  
206 included in each analytical batch to detect background contamination. Duplicate samples were  
207 analyzed at a frequency of one in every ten samples to evaluate analytical precision.

208 For morphological observations, a fixed-position digital camera (Canon EOS 250D, 24.1 MP)  
209 was mounted on a tripod at 1.0 m distance perpendicular to the plant canopy. This setup ensured  
210 identical framing, distance, and illumination for all photographs. Reference scale markers (2  
211 cm  $\times$  2 cm grid) were placed beside each pot for image calibration. Plant length was measured  
212 with a digital caliper (Mitutoyo 500-196-30, Japan); leaf area at harvest was measured using  
213 Easy Leaf Area software (Easlon & Bloom, 2014) based on individual leave scanned on a flat  
214 A4 surface. Oven-drying of plant samples was performed in a forced-air drying oven  
215 (Memmert UF110, Germany) at 80 °C until constant weight, and dry weight was measured  
216 using an analytical balance (Shimadzu ATX224,  $\pm 0.0001$  g). All laboratory glassware was  
217 acid-washed (10 %  $\text{HNO}_3$ ) and rinsed with deionized water before use. Lastly, all analytical  
218 procedures followed internal quality-management protocols of the Saraswanti Indo Genetech  
219 Laboratory, which are aligned with ISO/IEC 17025 standards.

220 To quantitatively evaluate plant performance under heavy metal exposure, a mathematical  
221 framework integrating growth response, metal accumulation, translocation efficiency, and  
222 human health risk was applied. Morphological performance was expressed as relative growth  
223 change (%) compared to the control treatment. Metal accumulation performance was quantified  
224 using the bioconcentration factor (BCF), while metal mobility within plants was assessed using  
225 the translocation factor (TF). Food safety performance was evaluated using Estimated Daily  
226 Intake (EDI), Hazard Quotient (HQ), and Hazard Index (HI). These indices collectively  
227 represent the ability of a plant to tolerate metal stress while minimizing human exposure risk.  
228 Plants exhibiting high growth tolerance,  $\text{BCF} > 1$ ,  $\text{TF} < 1$ , and  $\text{HI} <$  critical thresholds are  
229 considered to have superior performance under contaminated irrigation conditions.

230 **3.3 Plant material and substrate**  
231 Velvetleaf, kangkong, and kailan seeds were treated differently depending on the plant's  
232 characteristics. Velvetleaf seeds were sown in seedling trays filled with soil until sprouts  
233 appeared in approximately 1 month, then transferred to the nursery. Kangkong was propagated  
234 vegetatively using 10 cm stem cuttings from the base of the stem. The kangkong cuttings were  
235 planted in soil, and once four leaves had developed, the seedlings moved to the nursery.  
236 Meanwhile, kailan seeds were sown in rockwool until sprouts appeared in approximately 3  
237 days and transfer to the nursery. The nursery medium for all three plants is a mixture of soil  
238 and manure in a 1:1 ratio. After two weeks in the nursery, the roots were gently washed free of  
239 adhering soil and each plant was transferred to a plastic pot (25 cm × 20 cm) containing 2 kg  
240 of washed, sieved river sand per pot to provide an inert, low-background substrate. Natural  
241 ventilation and sunlight were maintained; rainfall was excluded by the polyethylene roof.

242 **3.4 Heavy-metal solutions: preparation and treatment on plants**  
243 The exposure phase involved the application of aqueous heavy metal solutions through the root  
244 zone of each plant in 18 days exposure. The metals used were Pb, Cd, and Cu, applied in the  
245 form of  $\text{Pb}(\text{NO}_3)_2$ ,  $\text{Cd}(\text{NO}_3)_2$ , and  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ , respectively. All chemical reagents were  
246 obtained from certified analytical-grade suppliers. Stock solutions were prepared using  
247 distilled water and subsequently diluted to final concentrations of 50 ppm Pb, 1 ppm Cd, and  
248 2 ppm Cu. These concentrations were determined based on previous phytotoxicity studies that  
249 demonstrated observable yet sub-lethal effects on plant growth (Aboelkassem et al., 2022;  
250 Doğan et al., 2022). Fresh working solutions were prepared daily. Concentrations at each  
251 renewal were verified by ICP-OES PlasmaQuant 9100 Series using five-point external  
252 calibration ( $0.01\text{--}10 \text{ mg L}^{-1}$ ;  $r^2 \geq 0.999$ ). Continuing calibration verification, method blanks,  
253 and duplicates were run every 10 samples.

254 Each pot received  $400 \text{ mL day}^{-1}$  of the assigned solution for 18 days to ensure continuous root  
255 contact. Control plants were irrigated with distilled water only. To prevent nutrient deficiency  
256 during exposure, foliar fertilization using Bayfolan D was applied at a concentration of  $2 \text{ mL}$   
257  $\text{L}^{-1}$  every four days throughout the experimental period.

258 **3.5 Morphological observation and harvest**  
259 Morphological observation in this study was conducted to measure and visually document the  
260 growth dynamics of plants under different heavy metal exposure conditions. The observation  
261 included plant length, total leaf number, maximum leaf length, and leaf width, which were

262 measured every two days beginning two days before exposure until harvest (day 18). To  
263 complement quantitative measurements, visual documentation was performed throughout the  
264 experiment. Each plant was photographed at the same distance, angle, and illumination using  
265 a fixed-position digital camera, enabling accurate visual comparison of morphological growth  
266 across treatments (**Figure 3**). These photographs provided qualitative evidence of growth  
267 inhibition and stress symptoms under Pb, Cd, and Cu exposure. At the end of the exposure  
268 period (day 18), all plants were carefully harvested and separated into roots, stems, and leaves.  
269 Each tissue was thoroughly rinsed with deionized water, followed by a brief wash in 0.01 M  
270 EDTA for 30 seconds to remove any surface-adsorbed metals, then rinsed again, blotted dry,  
271 and oven-dried at 80 °C to constant weight. Leaf area at harvest was quantified using scanned-  
272 image analysis with the Easy Leaf Area software (Easlon & Bloom, 2014). During the growth  
273 phase, daily leaf area was estimated using a validated linear regression model based on non-  
274 destructive morphological measurements (Lakitan et al., 2023, 2025). Overall, this combined  
275 quantitative and photographic approach allowed real-time tracking of morphological  
276 responses, providing both visual and analytical evidence of stress adaptation in velvetleaf,  
277 kangkong, and kailan exposed to Pb, Cd, and Cu.



278  
279 **Figure 3.** Morphological observation of velvetleaf (*Limnocharis flava*), kangkong (*Ipomoea*  
280 *aquatica*), and kailan (*Brassica oleracea* var. *alboglabra*) during 18 days of exposure to Pb  
281 (50 ppm), Cd (1 ppm), and Cu (2 ppm).

282 **3.6 Heavy metal measurements, Bioaccumulation and Translocation Factors**

283 Heavy metal analysis was performed on dried plant roots and leaves. Plant samples were taken  
284 as composites by collecting leaves and roots from each treatment. Measurement of heavy metal  
285 concentration using Inductively Coupled Plasma Optical Emission Spectrometry (ICP-OES).

286 Bioconcentration factor (BCF) and translocation factor (TF) were calculated as

287 
$$BCF = \frac{C_p}{C_w} \quad (1)$$

288 where  $C_p$  represents the metal concentration in the plant tissue ( $\text{mg kg}^{-1}$ ) and  $C_w$  denotes the  
289 metal concentration in irrigation water ( $\text{mg L}^{-1}$ ). The TF, which represents the efficiency of  
290 metal transport from roots to leaves, was calculated as:

291 
$$TF = \frac{C_{leaf}}{C_{root}} \quad (2)$$

292 where  $C_{leaf}$  and  $C_{root}$  are the metal concentrations in the leaves and roots, respectively. BCF  
293 values greater than 1 indicate a high accumulation capacity, while TF values greater than 1  
294 suggest efficient translocation from roots to aerial tissues (Wibowo et al., 2023).

295 **3.7 Health Risk Assessment**

296 Health Risk Assessment (HRA) was conducted to evaluate the potential risk of heavy  
297 metal contamination in vegetables grown under polluted irrigation water. The analysis focused  
298 on Estimated Daily Intake (EDI), Hazard Quotient (HQ), and Hazard Index (HI), which are  
299 commonly used to assess the potential health risks posed by heavy metal accumulation through  
300 consumption of contaminated plants. The EDI for each metal (Pb, Cd, Cu) was calculated using  
301 the following equation:

302 
$$EDI = \frac{C_{metal} \times F \times W}{BW \times 1000} \quad (3)$$

303  $C_{metal}$  is the metal concentration in the edible part of the vegetable ( $\text{mg/kg}$ ), F is the average  
304 consumption rate of the vegetable ( $\text{g/day}$ ), W is the weight of the edible part ( $\text{kg}$ ), BW is the  
305 average body weight ( $\text{kg}$ ) of the population (typically 60 kg for adults).

306 The Hazard Quotient (HQ) is used to estimate the potential health risk of individual metals by  
307 comparing the intake with a reference dose (RfD), which is a level of daily exposure to a  
308 contaminant that is considered to be without risk of harmful effects.

309 
$$HQ = \frac{EDI}{RfD} \quad (4)$$

310 RfD is the reference dose (mg/kg/day), which is a standard value determined by health  
311 agencies, the standard for Pb RfD: 0.0035 mg/kg/day; Cd RfD: 0.001 mg/kg/day and Cu  
312 RfD: 0.05 mg/kg/day.

313 The Hazard Index (HI) is the sum of the HQ values for each metal. If the HI is greater than 1,  
314 it indicates a potential health risk due to the combined exposure to multiple metals.

315 
$$HI = \sum_i HQ_i \quad (5)$$

316 Where HQi is the Hazard Quotient for each metal (Pb, Cd, Cu). HQ < 1: No significant health  
317 risk; HQ  $\geq$  1: Potential health risk; HI  $\geq$  1: Cumulative health risk from exposure to multiple  
318 heavy metals.

### 319 **3.8 Statistical Analysis**

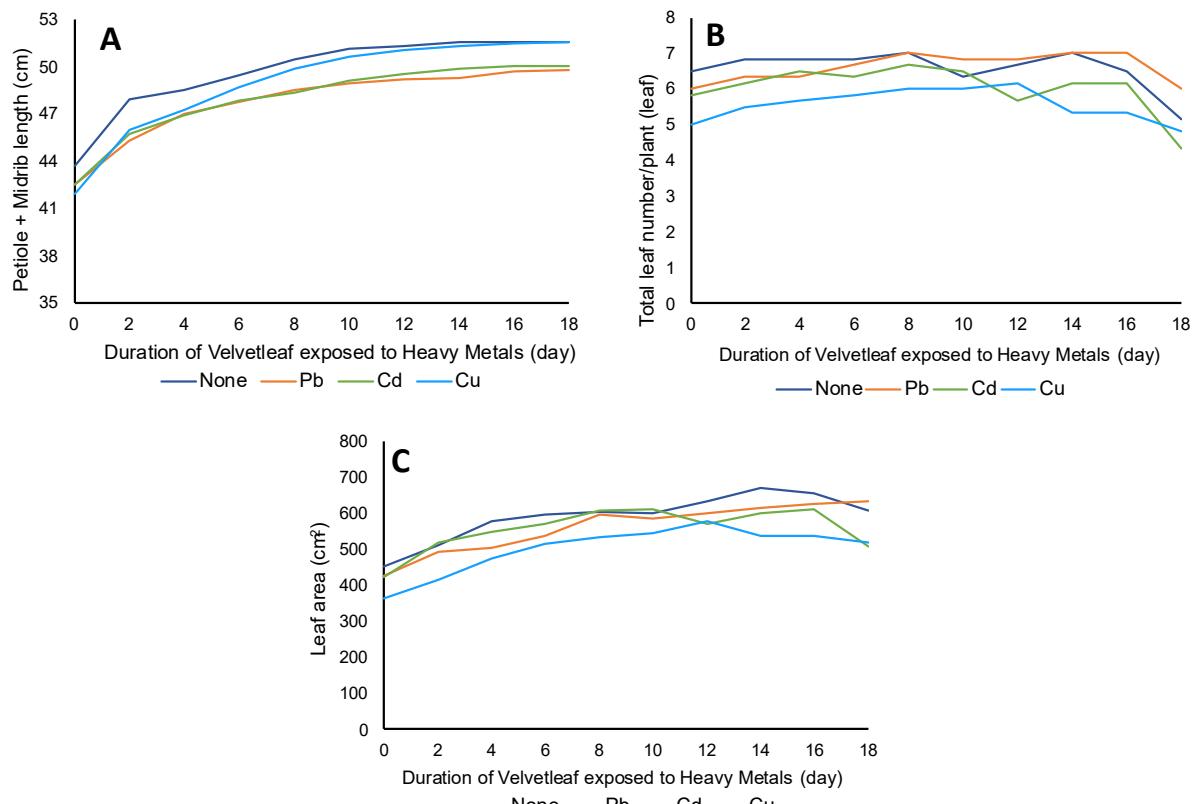
320 All analyses were performed in using IBM SPSS Statistics 25 for Windows (IBM  
321 Corporation, Armonk, USA). Pot-level means were used as observations. Normality and  
322 homoscedasticity were assessed by Shapiro–Wilk and Levene tests; variables were transformed  
323 when needed. Treatment effects within each species were tested by one-way ANOVA followed  
324 by Duncan’s Multiple Range Test at  $\alpha = 0.05$ . Results are reported as mean  $\pm$  SD across pots  
325 ( $n = 3$ ). Last, reliability and reproducibility were strengthened by the use of independent  
326 experimental replications, standardized measurement protocols, and analytical quality control  
327 procedures, ensuring that observed treatment effects were not artifacts of measurement or  
328 sampling bias.

## 329 **4. RESULTS**

### 330 **4.1 Morphological Response of Plants**

331 The results showed that different types of plants exhibited different growth patterns due  
332 to exposure to various kinds of heavy metals. Exposure to Pb and Cd in velvetleaf showed a  
333 decrease in petiole length compared to velvetleaf without metals. Conversely, Cu exposure did  
334 not have a significant effect on velvetleaf, as velvetleaf did not experience any inhibition in  
335 petiole length. A decrease in plant length began to appear on days 10 to 18 in velvetleaf samples  
336 (**Figure 4**). The leaf number of velvetleaf tends to decrease in both exposure to metals and  
337 without metals. **Figure 4A** shows that petiole + midrib length increased gradually in all  
338 treatments during the initial exposure phase (0–8 days), indicating that early growth was not  
339 immediately inhibited by heavy metal stress. However, plants exposed to Pb and Cd exhibited  
340 a slower rate of elongation compared with the control, suggesting mild growth suppression due

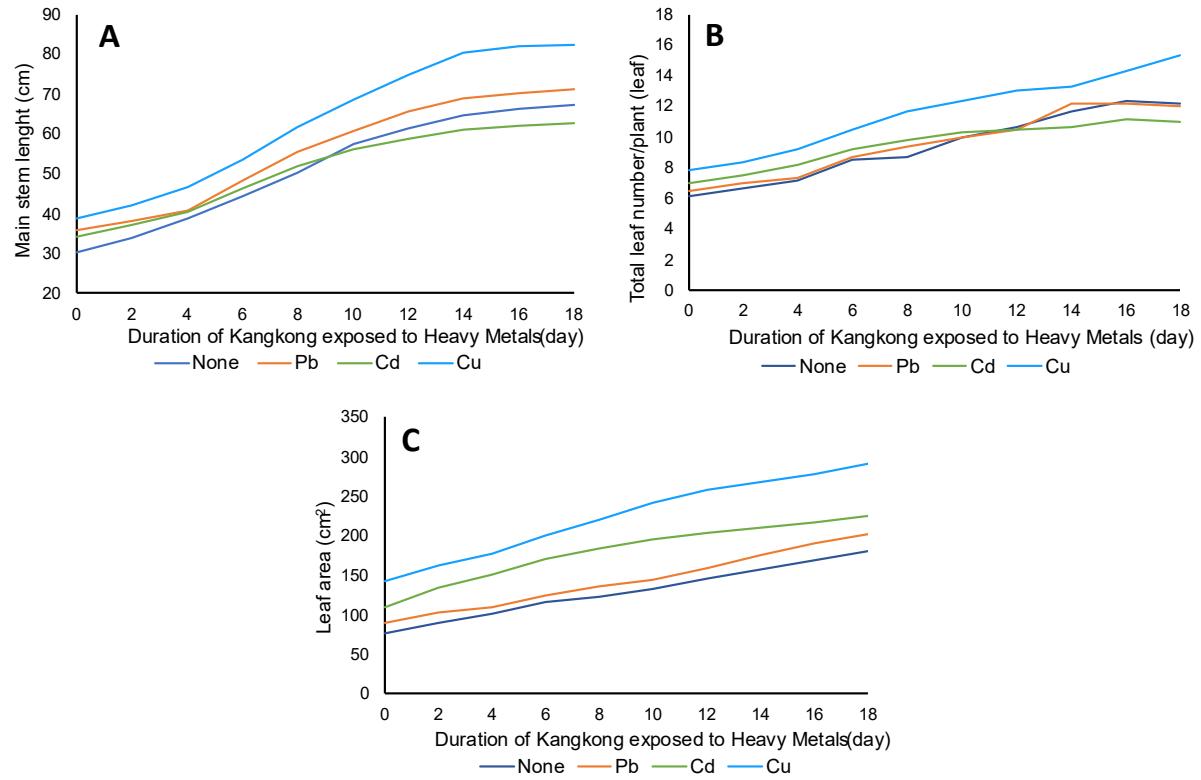
341 to metal toxicity. Among the metals, Cd had the most pronounced inhibitory effect after 12  
 342 days, consistent with its strong affinity for binding to cellular components and disrupting  
 343 physiological processes such as photosynthesis and enzyme activity. **Figure 4B** presents the  
 344 total leaf number per plant, which remained relatively stable throughout the exposure period.  
 345 A slight reduction in leaf number was observed in the Cd and Cu treatments after day 10,  
 346 indicating that prolonged exposure may have interfered with leaf initiation or accelerated  
 347 senescence. Pb exposure produced minimal effect, suggesting that Velvetleaf may exhibit some  
 348 degree of tolerance to Cd stress compared with Pb and Cu. **Figure 4C** illustrates the trend in  
 349 leaf area, which followed a similar pattern to petiole and midrib elongation. The control plants  
 350 showed continuous increases in leaf area, reaching the highest values around day 12–14. In  
 351 contrast, plants treated with Cd and Cu exhibited reduced leaf expansion toward the later stage  
 352 of exposure. This decline in leaf area may reflect both inhibited cell enlargement and potential  
 353 oxidative damage induced by heavy metal accumulation.



354  
 355 **Figure 4.** Effects of heavy metal exposure (Pb, Cd, and Cu) on Velvetleaf growth parameters  
 356 over time. (A) Petiole + midrib length (cm), (B) total leaf number per plant, and (C) leaf area  
 357 (cm<sup>2</sup>) were measured during 18 days of exposure. “None” represents treatment without heavy  
 358 metal addition.

359 Cd exposure resulted in the lowest reduction in both leaf number and total leaf area of  
360 velvetleaf. Meanwhile, in velvetleaf exposed to Pb, the decrease in leaf number occurs over a  
361 longer period, with leaves beginning to wilt and die on the 16th day after exposure. The total  
362 leaf area of velvetleaf also decreased along with the decrease in leaf number, which starts on  
363 day 10 after exposure. Velvetleaf exposed to Cd and Cu tended to experience a lower decrease  
364 in leaf area compared to exposure to Pb. However, there is a similar pattern between velvetleaf  
365 exposed to heavy metals and the control group.

366 Compared to velvetleaf, kangkong tends to be more resistant to heavy metal stress. At  
367 the beginning of exposure, kangkong showed normal growth until the 14th day of exposure. A  
368 decrease in plant length and leaf number of kangkong was seen after the 14th day. The lowest  
369 plant length, leaf number, and leaf area of kangkong occurred in the Cd exposure. Meanwhile,  
370 Cu exposure had a positive effect on kangkong growth, as indicated by higher increases in  
371 length, leaf number, and leaf area compared to kangkong without metals (**Figure 5**). **Figure**  
372 **5A** shows that main stem length increased steadily in all treatments during the experimental  
373 period, though plants subjected to Pb, Cd, and Cu exhibited reduced elongation rates compared  
374 with the control. The control plants reached the greatest stem length by day 18, while Pb and  
375 Cd treatments produced the most pronounced inhibition. This suggests that these metals  
376 interfere with cell division and elongation processes, likely by disrupting hormonal regulation  
377 and nutrient uptake. **Figure 5B** presents total leaf number per plant, which also increased over  
378 time in all treatments. However, the growth rate of new leaves was consistently lower under  
379 heavy metal exposure, particularly for Pb and Cd. Cu exposure caused only moderate  
380 suppression, implying that Kangkong may possess higher tolerance to Cu stress. The reduced  
381 leaf proliferation under Pb and Cd treatments might be linked to impaired photosynthetic  
382 efficiency and limited carbohydrate allocation to developing leaves. **Figure 5C** illustrates the  
383 effect of heavy metals on leaf area expansion. Control plants maintained the largest leaf areas  
384 throughout the experiment, while Pb- and Cd-treated plants showed the smallest. Cu exposure  
385 caused an intermediate response. The decline in leaf area under metal stress reflects the  
386 inhibitory effects of heavy metals on cell enlargement and chlorophyll synthesis, both of which  
387 are essential for optimal photosynthetic performance.

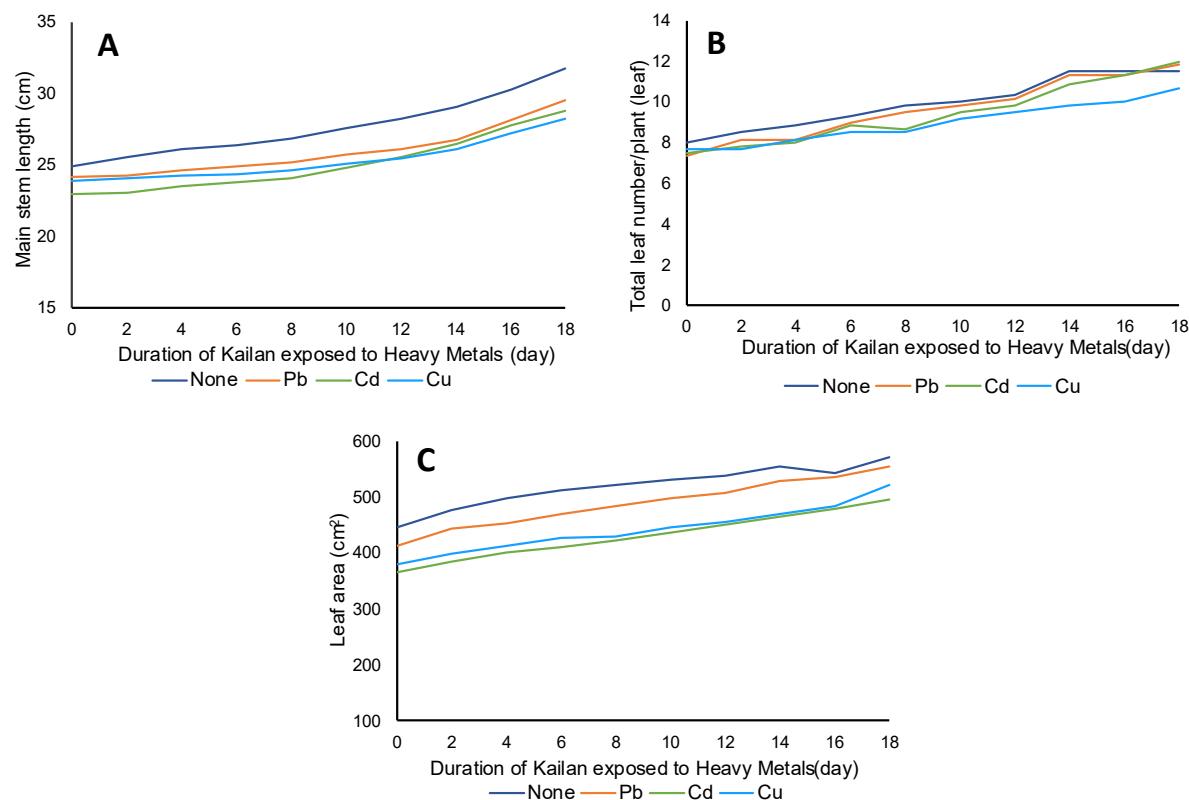


388

389 **Figure 5.** Growth response of Kangkong to heavy metals during exposure time: (A) Plant  
 390 length, (B) Total leaf number, and (C) Total leaf area

391 Unlike velvetleaf and kangkong, kailan showed a different growth response. In the initial  
 392 stages of exposure, there was a suppression of kailan length until day 12. However, after day  
 393 12, kailan length growth began to increase again, indicating a recovery mechanism against  
 394 heavy metal stress. The increase in leaf number and leaf area of kailan leaves also showed a  
 395 similar trend to plant length, where kailan experienced growth inhibition at the initial stage of  
 396 exposure. Exposure to Cd and Cu resulted in lower increases in plant length, leaf number, and  
 397 leaf area compared to Pb. The three types of heavy metals tested (Pb, Cd, and Cu) tended to  
 398 have a negative impact on kailan growth (**Figure 6**). **Figure 6A** shows that main stem length  
 399 increased progressively in all treatments over time, indicating continued growth despite the  
 400 presence of heavy metals. However, plants exposed to Cd and Cu exhibited lower elongation  
 401 rates relative to the control and Pb treatments. The inhibitory effect was more pronounced  
 402 under Cu exposure, suggesting that Cu toxicity more strongly affects stem elongation, possibly  
 403 by impairing cell wall extensibility and interfering with auxin-regulated growth processes.  
 404 **Figure 6B** illustrates the total leaf number per plant, which gradually increased throughout the  
 405 experimental period. The differences among treatments were relatively small, indicating that  
 406 Kailan maintained leaf production under moderate heavy metal stress. However, slight

407 suppression in leaf number was evident under Cd and Cu treatments, particularly after 12 days,  
 408 which may reflect a delayed impact of metal accumulation on leaf initiation and expansion.  
 409 **Figure 6C** shows the trend in leaf area, where the control plants maintained the largest leaf  
 410 area throughout the 18-day exposure period. Pb-treated plants showed a smaller but relatively  
 411 stable reduction compared with the control, whereas Cd and Cu caused greater declines in leaf  
 412 expansion. This reduction in leaf area under Cd and Cu stress may result from disturbances in  
 413 photosynthetic efficiency, chlorophyll degradation, and decreased turgor pressure factors  
 414 commonly linked with heavy metal-induced oxidative stress.



415  
 416 **Figure 6.** Growth response of Kailan to heavy metals during exposure time: (A) Plant length,  
 417 (B) Total leaf number, and (C) Total leaf area

418 The analysis of variance reveals no significant differences in the increase in leaf length  
 419 and leaf area in velvetleaf and kailan following exposure to Pb, Cd, and Cu metals. It was  
 420 different for kangkong, where there were significant differences in plant length between  
 421 kangkong exposed to Cu and kangkong exposed to Pb and Cd. Metal exposure gives  
 422 significant differences in the increase in leaf number of velvetleaf and kailan. Velvetleaf  
 423 exposed to Cd has the lowest leaf number and is significantly different from other metals.  
 424 Meanwhile, exposure to Cu in the leaf number of kailan showed a significant difference

425 compared to Pb and Cd. However, exposure to Pb, Cd, and Cu metals showed substantial  
426 differences in the increase in leaf area in all plants. (**Table 1**).

427 Exposure to heavy metals Pb, Cd, and Cu caused a decrease in the dry weight of kailan.  
428 However, it did not have a negative effect on the dry weight of velvetleaf. Meanwhile, in  
429 kangkong, only exposure to Pb caused a decrease in dry weight. The analysis of variance  
430 showed that the dry weight of kangkong exposed to Pb was significantly different from that  
431 exposed to Cd and Cu (**Figure 7**). Identification and comparison of morphological plants  
432 showed in **Table 2**.

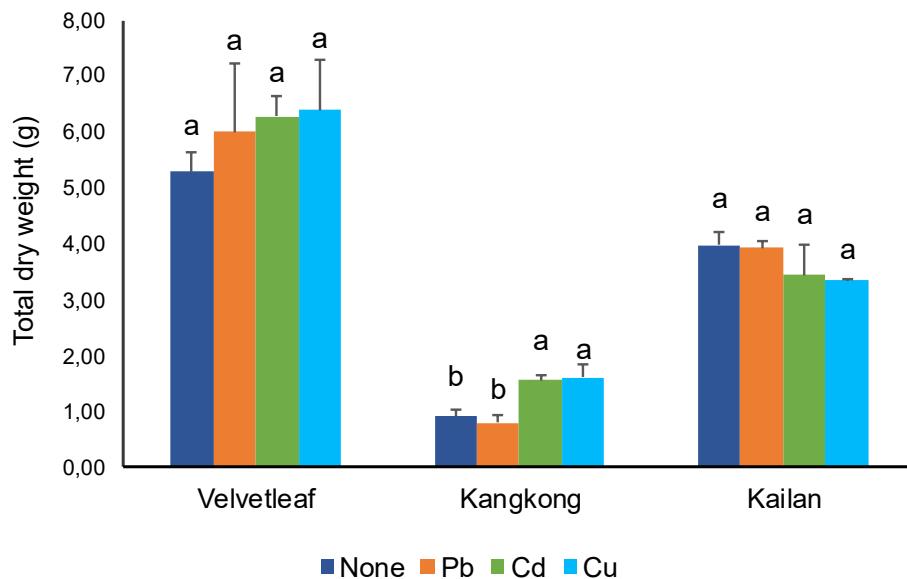
433

434 **Table 1.** Increase in plant length, leaf number, and total leaf area in various types of metal

Parameter	Treatment	Velvetleaf	Kangkong	Kailan
<b>Increase in plant length (cm)</b>	Without metal	7.90 a	36.93 ab	6.63 a
	Pb	7.22 a	35.40 bc	5.40 a
	Cd	7.48 a	28.50 c	5.80 a
	Cu	9.57 a	43.47 a	4.30 a
<b>Increase in leaf number (leaves/plant)</b>	Without metal	-1.33 bc	6.00 a	3.50 ab
	Pb	0.17 a	5.50 a	4.50 a
	Cd	-1.50 c	4.00 a	4.50 a
	Cu	-0.17 ab	7.50 a	3.00 b
<b>Increase in total leaf area (cm<sup>2</sup>)</b>	Without metal	156.23 a	111.08 a	123.91 a
	Pb	204.13 a	112.68 a	142.73 a
	Cd	136.78 a	123.64 a	131.75 a
	Cu	154.84 a	147.85 a	140.49 a

435

436 Values followed by different letters in the same column indicate significant differences  
437 based on DMRT at  $p < 0.05$ . Negative values indicate a decrease.



438  
439  
440

**Figure 7.** Total dry weight of plants on various types of metal. Different letters on each bar indicate significant differences based on DMRT at  $p < 0.05$ .

441 **Table 2.** Morphological Responses of Plants to Heavy Metal Exposure

Plant species	Pollutants Tested	Morphological effects	Ref.
Velvetleaf, Kangkong, Kailan	Pb, Cd, Cu	Chlorosis, reduced leaf size, stunted growth, leaf curling, necrosis	This study
<i>Spinacia oleracea, Amaranthus grain, Mentha spicata</i>	Cr, Ni, Zn, Cu, Pb	Growth inhibition, yellowing, wilting, leaf deformation	(Singh et al., 2024b)
Malabar spinach, 13 leafy species	Pb, Cd, As, Cu	Growth retardation, biomass reduction, low Cd accumulation in select spinach varieties	(Cui et al., 2022)
Lettuce, radish, carrot	Fe, Cu, Cr, Zn	Lettuce: chlorosis; Radish: reduced root length; Carrot: mild symptoms	(Qureshi et al., 2016)

General vegetables (review)	As, Cd, Hg, Pb	Overall yield reduction, leaf necrosis, root blackening	(Manwani, Devi, et al., 2022)
Pumpkin, spinach, gourd varieties	Cr, Mn, Fe, Co, Ni, Cu, Zn, Cd, Pb	Leaf curling, reduced chlorophyll, thin leaves, early senescence	(Jolly et al., 2024)

442

#### 443 **4.2 Heavy metal uptake in plants**

444 Heavy metal content in the three types of plants showed the highest accumulation of  
 445 metals in plant tissues, namely Pb, followed by Cd and Cu (**Figure 8**). In terms of distribution,  
 446 metal absorption in the underground organs (roots) was much higher than in above-ground  
 447 organs (leaves). Pb uptake in roots was found to be highest in kailan at 2873.21 mg/kg,  
 448 velvetleaf at 1473.88 mg/kg, and kangkong at 363.20 mg/kg.

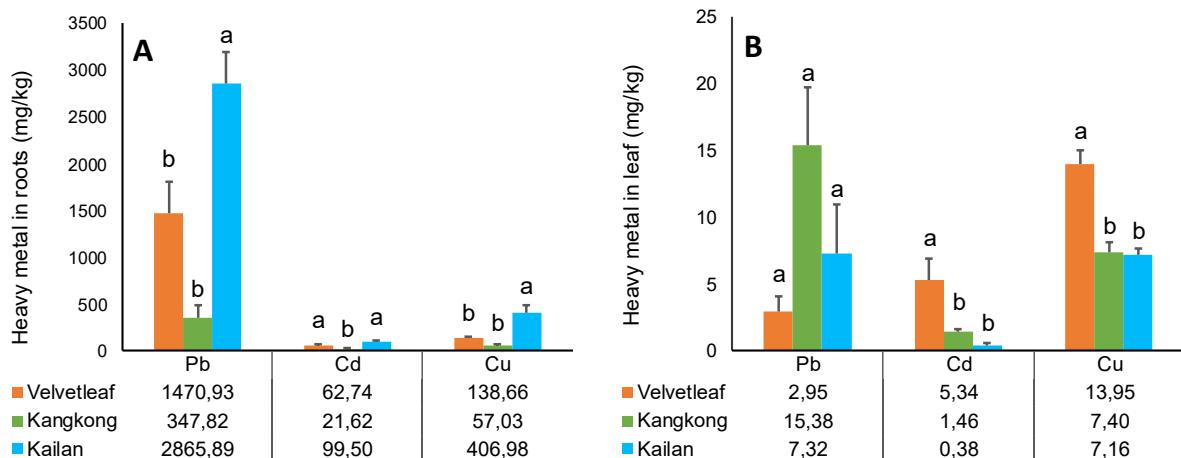
449 In contrast to roots, the highest Pb concentration in leaves was found in kangkong,  
 450 while Cd and Cu were found to be highest in velvetleaf leaves. The DMRT analysis ( $p < 0.05$ )  
 451 showed that there were significant differences between the concentrations of Pb, Cd, and Cu  
 452 in kailan roots and other plants. The results also showed significant differences between the  
 453 concentrations of Cd and Cu in velvetleaf leaves and the leaves of other plants.

454 The Bioconcentration Factor (BCF) for the three types of plants exposed to Pb, Cd, and Cu  
 455 were greater than 1. Kailan had the highest BCF value, followed by kangkong, and  
 456 velvetleaf. Among the three heavy metals tested (Pb, Cd, and Cu), the highest BCF values were  
 457 recorded in kailan, namely 2873.21 Pb, 99.87 Cd, and 414.13 Cu. The DMRT test results ( $p <$   
 458 0.05) showed that the BCF values of kailan showed a significant difference compared to  
 459 velvetleaf and kangkong. Meanwhile, the BCF values of velvetleaf and kangkong did not differ  
 460 significantly (**Table 3**).

461 The Translocation Factor (TF) values for the three plant species for Pb, Cd, and Cu  
 462 metals were greater than 1. The TF values for kailan for Cd and Cu were the lowest compared  
 463 to other plants, indicating that heavy metal translocation from the roots to the leaves was very  
 464 low. The test results showed that there were significant differences in TF for Cd and Cu in  
 465 kailan and velvetleaf. Meanwhile, TF for Pb did not differ significantly among the three plant  
 466 species (**Table 3**). In addition, **Table 4** showed the heavy metals concentration in plants  
 467 comparing with the WHO/FAO standard. This study showed that the concentration of metals  
 468 are higher than limit concentration in mg/kg plants.

469

470



471

472 **Figure 8.** Heavy metal concentrations in plants (A) in roots (B) and leaves. Different letters on  
 473 each bar indicate significant differences based on DMRT at  $p < 0.05$ .

474 **Table 3.** Bioconcentration factor (BCF) and translocation factor (TF) values for Pb, Cd, and  
 475 Cu metals in various types of plants

	Pb	Cd	Cu
<b>BCF</b>			
Velvetleaf	29.48 b	1.36 b	3.05 b
Kailan	2873.21 a	99.87 a	414.13 a
Kangkong	181.60 b	11.54 b	32.21 b
<b>TF</b>			
Velvetleaf	0.0028 a	0.0976 a	0.1027 a
Kailan	0.0028 a	0.0041 b	0.0192 b
Kangkong	0.0074 a	0.0677 ab	0.1442 a

476 Values followed by different letters in the same column indicate significant differences based on DMRT at  $p <$   
 477  $0.05$ .

478

479 **Table 4.** Comparison the heavy metals concentration in plants vs FAO/WHO standard

Heavy metals	Plant	Concentration in leaf	Max concentration (FAO/WHO, 2016) (mg/kg)

Pb	Velvetleaf Kangkong Kailan	2.95* 15.28* 7.32*	0.3
Cd	Velvetleaf Kangkong Kailan	5.34* 1.46* 0.38*	0.2
Cu	Velvetleaf Kangkong Kailan	13.95 7.4 7.16	40

480 \*Exceeding the maximum level

481

482

483 **4.3 Health Risk Assessment**

484 **Tables 5–7** show the Health Risk Assessment (HRA) results, which indicate that all  
485 three vegetables pose severe health risks due to Pb and Cd contamination in the edible parts.  
486 The Hazard Index for kale (3097.6) was the highest, followed by velvetleaf (1421.2) and water  
487 spinach (349.1), clearly exceeding the safe threshold of 1. Pb contributed the most to the HQ  
488 and HI, suggesting that Pb contamination is the dominant factor affecting human health. Cd  
489 also showed a significant contribution, particularly in kale. Cu posed a negligible risk as its HQ  
490 values were below 1. The results emphasize the urgent need for monitoring irrigation water  
491 and selecting crops with limited translocation of metals to edible parts to ensure food safety.

492 **Table 5.** Estimate Daily Intake of heavy metals in each plants

Plant	Metal	Concentration (mg/kg)	EDI (mg/kg/day)
Velvetleaf	Pb	1473.88	4.91
	Cd	5.50	0.018
	Cu	2.50	0.0083
Kangkong	Pb	363.20	1.21
	Cd	1.20	0.004
	Cu	1.50	0.005

Kailan	Pb	2873.21	9.58
	Cd	99.87	0.333
	Cu	414.13	1.38

493 \*Estimated Daily Intake (EDI)

494 **Table 6.** Analysis of Hazard Quotient (HQ)

Plant	Metal	EDI (mg/kg/day)	RfD	HQ
Velvetleaf	Pb	4.91	0.0035	1403
	Cd	0.018	0.001	18
	Cu	0.0083	0.05	0.166
Kangkong	Pb	1.21	0.0035	345
	Cd	0.004	0.001	4
	Cu	0.005	0.05	0.1
Kailan	Pb	9.58	0.0035	2737
	Cd	0.333	0.001	333
	Cu	1.38	0.05	27.6

495 \*Estimated Daily Intake (EDI); Reference Dose (RfD); Hazard Quotients (HQ)

496 **Table 7.** Hazzard Index

Plant	HI	Interpretation
Velvetleaf	1421.2	Very high health risk
Kangkong	349.1	Very high health risk
Kailan	3097.6	Extremely high health risk

#### 497 4.4 Challenges and Limitations

498 Despite providing valuable insights into the morphological responses, bioaccumulation, and  
499 translocation of heavy metals in leafy vegetables, this study has several challenges and  
500 limitations that should be acknowledged. First, the experimental setup was conducted under  
501 controlled pot-scale conditions, which, although useful for isolating variables, may not fully  
502 replicate field-scale dynamics. Factors such as fluctuating temperature, rainfall, microbial  
503 activity, and soil heterogeneity can alter metal bioavailability and plant uptake under real  
504 agricultural conditions. Future studies should therefore validate these findings across multiple  
505 growing seasons and varied soil types to confirm ecological consistency.  
506 Second, the analysis primarily focused on total metal concentrations without differentiating  
507 between free ionic and complexed forms. This limits the interpretation of true bioavailability,

508 as metal speciation strongly influences plant uptake and toxicity. Incorporating speciation-  
509 aware analysis and rhizosphere measurements such as pH, redox potential, and dissolved  
510 organic carbon would provide a more mechanistic understanding of plant-metal interactions.  
511 Third, while this research focused on morphological and accumulation parameters, the  
512 molecular and biochemical pathways underlying plant tolerance and detoxification were not  
513 investigated. Future research integrating transcriptomic or proteomic profiling could elucidate  
514 gene-level defense mechanisms and help identify biomarkers of metal stress.  
515 Fourth, the health risk assessment used standard reference consumption rates and exposure  
516 assumptions, which may vary across populations. Region-specific dietary data and chronic  
517 exposure modeling should be incorporated to refine risk estimations and enhance applicability  
518 to local food systems. Finally, current predictive and statistical approaches, though effective  
519 for experimental analysis, could be enhanced through hybrid data-driven models that integrate  
520 environmental sensing and optimization algorithms, such as Al-Biruni Earth Radius  
521 Optimization and Vision Graph Neural Networks. These advanced computational tools could  
522 improve the precision of contamination prediction and enable spatial mapping of risk zones.

## 523 5. DISCUSSION

524 Each type of plant has a different response to heavy metal exposure. The results showed  
525 that the growth of plants exposed to heavy metals tends to be lower than that of plants without  
526 heavy metals. Among the three types of plants tested, kailan showed initial stress to the toxicity  
527 of heavy metals Pb, Cd, and Cu, where plant length and leaf number were significantly  
528 inhibited at the beginning of exposure. Heavy metal exposure causes morphophysiological,  
529 biochemical, and molecular changes in plants, thereby inhibiting plant growth and productivity  
530 (Noor et al., 2022; Raza et al., 2020). This is evident from the decrease in plant height, number  
531 of branches, leaf area and biomass, to the decrease in wet weight and dry weight (Hu et al.,  
532 2023; Waheed et al., 2022). After 12 days of exposure, kailan growth increased again,  
533 indicating a recovery or adaptation phase of plants in a heavy metal-contaminated environment  
534 (Ghori et al., 2019).

535 When exposed to heavy metals, some plants experience initial stress characterised by  
536 the production of Reactive Oxygen Species (ROS), which can cause cell damage (Liu et al.,  
537 2023; Pande et al., 2022). To overcome this, plants activate various defence mechanisms by  
538 enhancing the antioxidant system and producing proteins, hormones, metabolites, and  
539 chelating molecules such as phytochelatin and metallothionein to reduce oxidative damage  
540 (Ghori et al., 2019; Liu et al., 2023). Phytochelatin and metallothionein chelate toxic metal

541 ions, forming complex compounds that are transported to vacuoles for storage, thereby  
542 reducing the concentration of heavy metals in the cytosol (Faizan et al., 2024). This response  
543 is crucial for enhancing metal detoxification capacity, repairing damage, and helping plants  
544 recover from initial stress and adapt to contaminated environments.

545 The concentration of heavy metals in kailan tissue was found to be very high (Pb  
546 2873.21 mg/kg, Cd 99.87 mg/kg, and Cu 414.13 mg/kg). It indicates that high heavy metal  
547 content in the environment causes plants to be unable to limit the amount of heavy metals  
548 absorbed. The ability of plants to limit the absorption of heavy metals in the environment is  
549 limited, among other things, by their absorption capacity. The capacity of vacuoles to store  
550 heavy metal complexes is limited, causing saturation and a decrease in the efficiency of metal  
551 absorption, especially in contaminated environmental conditions (Choppala et al., 2014; Teng  
552 et al., 2024). Results in this study are consistent with previous reports, which found that the  
553 increase in heavy metal concentrations in plants correlates with higher heavy metal  
554 concentrations in the environment and more prolonged exposure durations (Chen et al., 2015;  
555 Ghori et al., 2019).

556 In contrast to kailan, exposure to heavy metals did not have a significant effect on the  
557 growth of velvetleaf and kangkong. Neither plant showed initial signs of stress during  
558 exposure. This indicates that the concentrations of heavy metals used in the study were still  
559 tolerable for the plants. These plants are effectively able to avoid stress by limiting the entry of  
560 metals through root exudates and maintaining metal ion homeostasis (Ejaz et al., 2023; Ghori  
561 et al., 2019). This exudate plays a role in limiting metal absorption by stimulating peroxidase  
562 enzyme activity, which thickens the root cell walls (Noor et al., 2022). The root apoplast acts  
563 as a physical barrier that inhibits the movement of metal ions into plant tissues (Ejaz et al.,  
564 2023; Singh et al., 2024a).

565 In velvetleaf, a decrease in plant growth was observed after 10 to 14 days of exposure,  
566 including without metal. This indicates that the decrease in velvetleaf growth was not caused  
567 by heavy metal exposure, but rather because the plants entered the generative stage (flowering  
568 phase). It can be seen from the similarity in the pattern of growth reduction in the control group  
569 that was not exposed to metal. Heavy metal exposure also had no significant effect on the dry  
570 weight of velvetleaf and kangkong. It indicates that velvetleaf and kangkong have better  
571 tolerance and adaptation mechanisms to heavy metal exposure than kailan.

572 Based on the type of heavy metal, exposure to Pb and Cd resulted in lower plant length  
573 and slowed leaf growth in all three plants. Cd exposure had the most adverse effect on plant  
574 growth, both in velvetleaf, kailan, and kangkong. Pb and Cd are non-essential metals that are

575 toxic to plants. However, Cd has a higher density than Pb, so even at low concentrations, it  
576 causes toxicity that disrupts metabolism and inhibits plant growth. Cd is easily absorbed by  
577 plant roots and accumulates with exposure time, then transported to leaves and fruit (Ran et al.,  
578 2024). High Cd concentrations in plants caused a decrease in leaf stomatal density, limiting  
579 stomata and mesophyll conductance, resulting in slow growth due to disrupted photosynthesis  
580 (Guo et al., 2023; Hu et al., 2023). Increased Cd concentrations and exposure time in the three  
581 plant accelerate leaf yellowing (chlorosis) due to chlorophyll pigment degradation, leading to  
582 plant death (**Figure 9**). Previous studies have indicated a direct correlation between Cd  
583 exposure and chlorophyll breakdown, where increased Cd concentration caused a reduction in  
584 chlorophyll content in spinach (*Spinacia oleracea L.*) by up to 68% (Rydzyński et al., 2019),  
585 and 9–20% in Hybrid pennisetum (Song et al., 2019)

586 Meanwhile, Cu has a positive effect on the growth of velvetleaf and kangkong. Cu is  
587 an essential micronutrient that plays a role in various metabolic processes and physiological  
588 reactions in plants, including participating in the electron transport chain of photosynthesis and  
589 respiration, oxidative metabolism, cell wall metabolism, and hormone signaling (Nazir et al.,  
590 2019; Shabbir et al., 2020). Plants require Cu in low concentrations, but it becomes toxic at  
591 higher concentrations. There was a slight different between the essential and harmful  
592 concentrations of Cu in organisms. The presence of Cu in the growing medium encourages  
593 plants to absorb Cu because of their need as a micronutrient, but plants cannot control the  
594 amount of Cu absorbed, causing it to exceed the limit (Kumar et al., 2021; Xu et al., 2024).  
595



596  
597 **Figure 9.** Chlorosis on the leaves of Velvetleaf, Kangkong, and Kailan

598 **Table 2** summarizes the morphological alterations observed in various vegetable species  
599 exposed to different heavy metals, highlighting both interspecific variability and pollutant-  
600 specific responses. In this study, velvetleaf, kangkong, and kailan exhibited visible stress  
601 symptoms such as chlorosis, leaf curling, and growth inhibition under Pb, Cd, and Cu  
602 contamination. These morphological changes indicate disruptions in photosynthetic efficiency

603 and nutrient translocation caused by metal toxicity. Similar manifestations have been reported  
604 in *Spinacia oleracea*, *Amaranthus* grain, and *Mentha spicata* subjected to Cr, Ni, Zn, Cu, and  
605 Pb exposure, where leaf deformation and wilting were attributed to oxidative damage and  
606 impaired water balance (Singh et al., 2024b).

607 Consistent with these findings, *Basella alba* (Malabar spinach) and other leafy vegetables  
608 showed reduced biomass and visible growth retardation under Pb, Cd, As, and Cu exposure,  
609 suggesting that metal accumulation interferes with cell division and elongation processes (Cui  
610 et al., 2022). Root vegetables such as lettuce, radish, and carrot demonstrated organ-specific  
611 sensitivity, where chlorosis, root shortening, and mild necrotic symptoms were evident  
612 depending on the dominant metal present (Qureshi et al., 2016). A broader review by Manwani  
613 et al. (2022) confirmed that exposure to As, Cd, Hg, and Pb generally results in yield reduction,  
614 leaf necrosis, and blackening of roots, reinforcing that morphological deterioration is a reliable  
615 indicator of physiological stress.

616 Furthermore, studies on pumpkin, spinach, and gourd varieties exposed to multiple metals (Cr,  
617 Mn, Fe, Co, Ni, Cu, Zn, Cd, and Pb) demonstrated cumulative toxicity manifested through leaf  
618 thinning, early senescence, and reduced chlorophyll content (Jolly et al., 2024). Collectively,  
619 these results emphasize that morphological abnormalities serve as rapid, visible markers of  
620 heavy metal stress and can be used as preliminary diagnostic criteria before conducting  
621 biochemical or molecular analyses. The present study therefore aligns with previous reports,  
622 confirming that both leaf deformation and growth suppression are universal adaptive responses  
623 among leafy vegetables subjected to heavy metal contamination.

624 The ability of plants to accumulate pollutants from growing media, both water and soil,  
625 is calculated using the Bioconcentration Factor (BCF). Several studies have used BCF values  
626 to evaluate the potential for metal bioaccumulation by plants. The BCF for Pb, Cd, and Cu in  
627 the three types of plants tested were greater than 1, indicating that all three plants have a high  
628 potential for heavy metal accumulation (Dogan et al., 2018; Sipos et al., 2023). Based on the  
629 BCF values, the order of plant ability to accumulate Pb, Cd, and Cu is  
630 kailan<kangkong<velvetleaf. All plants are capable of absorbing heavy metals at high  
631 concentrations, thereby increasing the risk of heavy metal exposure through consumption.

632 Based on the distribution of heavy metals in plant parts, higher concentrations of heavy  
633 metals were detected in underground organs (root tissue) than in above-ground organs (leaf  
634 tissue) in all plant tested. Higher heavy metal accumulation in roots is a tolerance strategy of  
635 plants to reduce the effects of toxicity on photosynthetic tissues (Bonanno & Cirelli, 2017).  
636 The highest accumulation of Pb, Cd, and Cu was found in kailan roots, 2-3 times higher than

637 velvetleaf and kangkong. However, the heavy metals translocated to the kailan leaves were  
638 lower than those of velvetleaf and kangkong. The highest accumulation of heavy metals in the  
639 leaves was found in kangkong leaves. It indicates that kangkong tends to transfer more heavy  
640 metals from underground organs to above-ground organs.

641 In kangkong, 88–95% of heavy metals accumulate in the roots, and 4–11% are  
642 translocated to the leaves. Most heavy metals (90–99%) in velvetleaf also accumulate in the  
643 roots, with only 0.2–10% translocated to the leaves. Meanwhile, in kailan, heavy metal  
644 accumulation in the roots reaches 98–99%, while the concentration in the leaves is very low  
645 (0.25–1.73%). The Translocation Factor (TF) showed that all three plants have TF greater than  
646 1. The order of plants based on TF values was kangkong>velvetleaf >kailan, indicating that  
647 kangkong transferred more heavy metals to leaf tissue than velvetleaf and kailan. All three  
648 plant species have  $BCF > 1$  and  $TF < 1$ ; thus, they can be categorised as phytostabilizers, where  
649 the plants are capable of retaining heavy metals in the roots and preventing their mobilisation  
650 to leaf tissues (Hidayati & Rini, 2020).

651 The concentrations of heavy metals Pb, Cd, and Cu in the three plants tested exceeded  
652 the maximum levels set by FAO/WHO for leafy vegetables, namely 0.3 mg/kg for Pb, 0.2  
653 mg/kg for Cd, and 40 mg/kg for Cu. The comparison between the measured heavy-metal  
654 concentrations in plant tissues and the FAO/WHO permissible limits shows a clear pattern of  
655 excessive accumulation for Pb and Cd across all three vegetable species. Lead concentrations  
656 in velvetleaf (2.95 mg/kg), kangkong (15.28 mg/kg), and kailan (7.32 mg/kg) exceed the  
657 recommended maximum limit of 0.3 mg/kg by 9.8- to 51-fold. Kangkong demonstrates the  
658 highest Pb uptake, suggesting strong bioaccumulation potential, possibly due to its aquatic  
659 growth habit that enhances metal absorption from contaminated irrigation water or sediment.

660 Cadmium levels follow a similar trend, with velvetleaf (5.34 mg/kg), kangkong (1.46  
661 mg/kg), and kailan (0.38 mg/kg) all exceeding the FAO/WHO limit of 0.2 mg/kg (FAO/WHO,  
662 2016). The concentration in velvetleaf is particularly high, reaching more than 26 times the  
663 permissible level. These results indicate a substantial health risk because Cd is known for its  
664 high mobility in soil–plant systems and can accumulate in edible tissues even at relatively low  
665 environmental concentrations. In contrast, copper concentrations in all plant samples remain  
666 below the FAO/WHO limit of 40 mg/kg. Even though Cu is an essential micronutrient,  
667 excessive levels can become toxic; however, the measured values (7.16–13.95 mg/kg) suggest  
668 that Cu contamination is not a primary concern at the study site. The distinction between toxic  
669 (Pb, Cd) and non-toxic (Cu) levels highlights the influence of metal speciation, soil chemistry,  
670 and plant physiology on uptake behavior.

671 Food security under conditions of heavy metal contamination can be achieved through  
672 integrated control at multiple stages of the food production chain. At the irrigation level, regular  
673 monitoring and regulation of water quality are essential to prevent chronic input of Pb and Cd  
674 into agricultural systems. When complete water treatment is not feasible, the selection of  
675 vegetable species with low translocation potential becomes critical. The present study  
676 demonstrates that although all tested vegetables accumulated metals ( $BCF > 1$ ), differences in  
677 translocation behavior strongly influence food safety outcomes. Species with lower TF values  
678 restrict metal movement to edible tissues, thereby reducing direct dietary exposure. However,  
679 low TF alone is insufficient; it must be complemented with agronomic mitigation strategies  
680 such as soil pH adjustment, organic amendments, and metal-immobilizing materials to reduce  
681 bioavailable fractions. From a public health perspective, security is achieved not by plant  
682 tolerance alone, but by reducing human exposure below acceptable risk thresholds. Therefore,  
683 integrating crop selection, water quality management, and regulatory enforcement is essential  
684 to ensure safe vegetable production in contaminated environments.

685 The HRA revealed alarming implications for human consumption. The Target THQ  
686 values for Pb and Cd greatly exceeded the safe threshold ( $HQ > 1$ ) in all three plants, particularly  
687 in kale, which had a Hazard Index of 3097.6. Velvetleaf and kangkong also posed severe health  
688 risks, with HIs of 1421.2 and 349.1, respectively. Pb was the dominant contributor to risk,  
689 followed by Cd, while Cu presented negligible risk ( $HQ < 1$ ). These results highlight that even  
690 crops with higher tolerance to heavy metal stress, like velvetleaf and kangkong, can accumulate  
691 metals to hazardous levels, emphasizing the necessity of monitoring irrigation water quality  
692 and implementing mitigation strategies before cultivation.

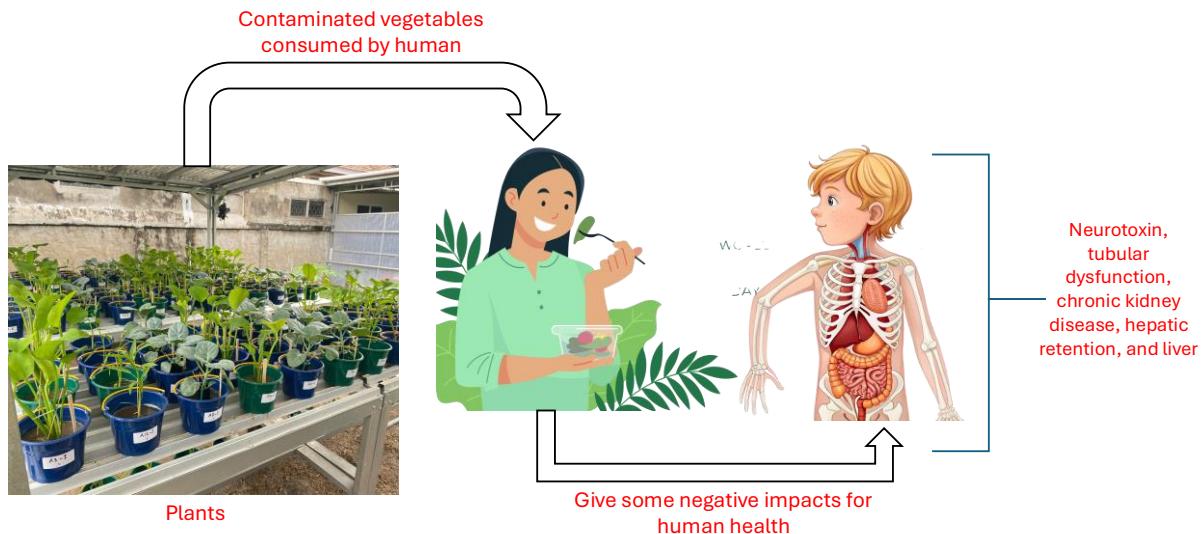
693 Several studies have highlighted the risks of human exposure to heavy metals and their  
694 associated health impacts. For instance, in China, rice samples contained Cd concentrations  
695 9.35 times above the Chinese maximum permissible level, emphasizing the risk of dietary  
696 exposure (Cai et al., 2019). Similarly, in Iran, Pb in cow meat showed the highest Hazard  
697 Quotient (HQ) among the metals analyzed, while Cd in cow kidney posed the greatest  
698 carcinogenic risk (CR). Although the overall Hazard Index (HI) was greater than 1, children  
699 were found to be more susceptible, indicating that frequent consumption of contaminated meat  
700 could pose significant health concerns (Zeinali et al., 2019).

701 In vitro studies further support the harmful effects of heavy metals, particularly when  
702 combined. Human gastric epithelial cells exposed to Cd (5  $\mu$ M) and Cu (10  $\mu$ M)  
703 simultaneously exhibited a 40% loss in cell viability, a 3.5-fold increase in reactive oxygen  
704 species (ROS) indicative of oxidative stress, 36% cell cycle arrest in the S phase, and a 22%

705 apoptosis rate (Wang et al., 2021). These findings indicate that co-exposure to Cd and Cu is  
706 more detrimental than exposure to either metal individually, highlighting the synergistic  
707 toxicity of heavy metals (Wang et al., 2021). Biomonitoring data also reflect chronic exposure  
708 levels in human populations. Overall, these studies demonstrate that lead, cadmium, and copper  
709 are strongly linked to adverse health effects through multiple exposure routes, including food,  
710 air, and occupational contact. Even low-level, chronic exposure can lead to oxidative stress,  
711 organ damage, and carcinogenic outcomes, particularly when metals act synergistically. These  
712 findings underscore the critical importance of monitoring metal contamination in food sources,  
713 irrigation water, and the environment to minimize cumulative health risks in vulnerable  
714 populations.

715 The present findings have direct implications for public health because leafy vegetables  
716 are widely consumed and can meaningfully contribute to dietary exposure when irrigation  
717 water contains bioavailable Pb, Cd, and Cu. Lead is a developmental neurotoxin; even low  
718 prenatal and early-life exposures are linked to decrements in intelligence quotient and adverse  
719 neurocognitive outcomes, while adult exposures are associated with hypertension and kidney  
720 disease (Ramírez Ortega et al., 2021). Cadmium exhibits very long biological half-lives in  
721 humans and preferentially accumulates in the kidney, elevating risks of tubular dysfunction  
722 and chronic kidney disease; diet is a dominant exposure route globally (Charkiewicz et al.,  
723 2023). Although copper is an essential micronutrient, sustained intakes above updated health-  
724 based guidance values raise concern for hepatic retention and potential liver injury,  
725 underscoring the importance of controlling cumulative exposures from food (More et al.,  
726 2023).

727 **Figure 10** showed the pathway of heavy-metal contamination from vegetables to  
728 human health impacts. Irrigation-water contaminants such as lead (Pb), cadmium (Cd), and  
729 copper (Cu) can be taken up by plants and accumulate in edible tissues; consumption elevates  
730 non-carcinogenic risk (HQ and HI). Documented outcomes include neurodevelopmental  
731 deficits and hypertension for Pb, renal tubular dysfunction and chronic kidney disease for Cd,  
732 and hepatic effects at excessive Cu intake. Arrows indicate the progression from contaminated  
733 plants to exposure and health impacts; mitigation can target water quality, soil and cultivar  
734 management, and household preparation to reduce bioaccessibility.



735

736 **Figure 10.** Conceptual pathway from heavy-metal-contaminated vegetables to human health  
737 outcomes

738 Risk characterization for leafy-vegetable consumption should therefore use established  
739 non-cancer metrics based on the hazard quotient (HQ) and hazard index (HI). HQ values  
740 greater than 1 for individual metals, or HI values greater than 1 for mixtures, indicate potential  
741 concern for non-carcinogenic effects and should trigger mitigation. Because consumers  
742 experience co-exposure to multiple metals and foods, reliance on single-metal assessments can  
743 underestimate total risk. Mixed-metal ingestion may also worsen intestinal barrier integrity and  
744 perturb the gut microbiome, amplifying systemic inflammation and metabolic risk, which  
745 suggests that risk estimates based solely on total concentrations may understate overall health  
746 burdens.

747 The findings of this study complement recent developments that combine  
748 environmental monitoring, artificial intelligence, and sustainable agricultural management.  
749 Advanced computational models have demonstrated a strong ability to improve decision-  
750 making in food and crop systems. For instance, the reuse of nutrient-rich water using Recurrent  
751 Neural Networks (RNN) and Natural Language Processing techniques has shown potential for  
752 optimizing nutrient recovery and supporting healthier food practice (K et al., 2025). Similarly,  
753 a comparative study using Support Vector Machines (SVM) and Decision Tree algorithms for  
754 predicting hydroponic tomato growth revealed that data-driven models can effectively capture  
755 non-linear relationships between environmental factors and crop yield (B et al., 2024). In  
756 addition, real-time irrigation optimization in coffee plantations using Bi-Directional RNNs and  
757 Internet of Things (IoT) sensors demonstrated the value of integrating continuous data streams  
758 to enhance water use efficiency and crop productivity (Shahriar et al., 2021). Research on

759 lettuce farming using Random Forest algorithms also indicated that this model outperforms  
760 conventional Decision Tree techniques in predicting plant performance and managing  
761 cultivation parameters (Jegan et al., 2024) .

762 Future research should prioritize multi-season field validation across contrasting soils  
763 and climatic regimes to determine whether the pot-scale responses observed here persist under  
764 on-farm irrigation practices. Studies should incorporate speciation-aware exposure,  
765 distinguishing free ions from complexed forms, together with rhizosphere measurements of  
766 pH, redox potential, and dissolved organic carbon. This approach will help disentangle  
767 bioavailability from total concentration and clarify mechanisms that govern uptake,  
768 bioconcentration, and translocation. Longer observation windows that span full growth cycles,  
769 including a post-exposure depuration phase in clean water, are also recommended to assess  
770 whether edible tissues can reduce accumulated metals and to quantify any agronomic costs.  
771 Because real wastewaters contain mixtures, future designs should shift from single-metal tests  
772 to factorial or response-surface studies that span realistic multi-metal ranges, for example  
773 Pb×Cd×Cu with Zn or Ni as co-contaminants. In parallel, screening of cultivars and landraces  
774 within each species is needed to identify low-translocation phenotypes that maintain yield and  
775 quality under stress. Mechanistic depth can be added through ionomics and multi-omics,  
776 including transcriptomics, proteomics, and metabolomics, to resolve antioxidant responses,  
777 chelation pathways, and vacuolar sequestration. Spatial imaging such as micro-XRF or LA-  
778 ICP-MS can verify tissue-level sequestration targets suggested by physiological measurements.  
779 Translational work should assemble integrative mitigation packages that combine soil  
780 amendments, for example liming, silicon, phosphate, organic matter, or biochar, with foliar  
781 nutrition and microbe-assisted strategies. Candidate consortia include plant growth-promoting  
782 and metal-immobilizing microbes, mycorrhizae, and mineral additives such as iron or  
783 manganese oxides and layered double hydroxide-based sorbents. The goal is to suppress  
784 bioavailability and leaf translocation without penalizing yield. On the consumer side,  
785 experiments that quantify how washing, blanching, pickling or fermentation, and common  
786 cooking methods alter both total concentrations and gastrointestinal bioaccessibility will refine  
787 health-risk estimates in ways that are directly relevant to household practice. These datasets  
788 should feed into probabilistic exposure models that use locally stratified intake distributions by  
789 age, sex, and season, and that report both uncertainty and variability. The same models should  
790 be used to derive irrigation-water trigger values, defined as concentration–time safe operating  
791 envelopes that keep edible-leaf metals below regulatory limits.

792 In addition, next study also should considerate environmental monitoring and precision  
793 agriculture increasingly rely on advanced sensing, data-analysis and modelling techniques to  
794 characterise contamination, plant stress and crop performance (Maruthai et al., 2025; Sridharan  
795 Sivasubramanian et al., 2025). Finally, methodological enhancements will strengthen  
796 reproducibility and policy uptake. Standardized imaging for longitudinal leaf-area tracking,  
797 photographic documentation of all experimental stages, preregistered analysis plans, and a  
798 priori power calculations will improve inference on treatment effects. Socio-economic and  
799 policy integration, including cost–benefit and life-cycle assessments of on-farm versus water-  
800 side mitigation, hotspot mapping, and co-designed risk communication with farmers and local  
801 regulators, will help convert experimental insights into practical guidance. This should include  
802 crop-selection recommendations for settings where water treatment is infeasible and decision  
803 frameworks that link routine water monitoring to actionable farm-level responses.

## 804 6. CONCLUSIONS

805 This study revealed that vegetables exposed to Pb, Cd, and Cu-contaminated irrigation  
806 water exhibit different morphological responses. Kailan showed initial growth inhibition, while  
807 velvetleaf and kangkong tolerated exposure for longer periods. Quantitatively, kailan  
808 accumulates the highest concentrations. All plants displaying  $BCF > 1$  and  $TF < 1$  are classified  
809 as phytostabilizers. Despite root retention, Pb and Cd concentrations in leaves consistently  
810 exceeded FAO/WHO safety limits, resulting in very high health risk indices for kailan,  
811 followed by velvetleaf and kangkong. Therefore, effective mitigation strategies are needed,  
812 including monitoring and managing irrigation water quality, and selecting vegetable varieties  
813 that are capable of root sequestration without leaf accumulation. This strategy is necessary to  
814 reduce the risk of contamination and support the development of safe vegetable cultivation for  
815 human health. Future research should prioritize multi-season field trials across diverse soil  
816 types and climatic conditions to validate whether the pot-scale responses observed in this study  
817 are consistent under real on-farm irrigation practices. Subsequent studies should also  
818 incorporate speciation-sensitive analyses that distinguish between free ionic and complexed  
819 metal forms, alongside comprehensive rhizosphere assessments of pH, redox potential, and  
820 dissolved organic carbon dynamics to better elucidate metal bioavailability and plant uptake  
821 mechanisms. In addition, food security in heavy metal–affected irrigation systems can be  
822 achieved through a combination of preventive water management, selection of low-  
823 translocation crops, and agronomic mitigation to suppress metal bioavailability. Although plant  
824 tolerance allows survival under contamination, safety for human consumption requires

825 minimizing metal transfer to edible tissues and maintaining exposure levels below health-based  
826 limits. These findings provide a scientific basis for risk-informed crop selection and irrigation  
827 management to protect public health in peri-urban agricultural systems.

828 Future research should extend this work beyond pot-scale conditions through multi-  
829 season field trials across different soil types and climatic settings to validate the consistency of  
830 plant responses under real irrigation practices. Further studies should incorporate metal  
831 speciation and rhizosphere processes, including pH, redox potential, and dissolved organic  
832 carbon, to better elucidate bioavailability-driven uptake mechanisms. In addition, experiments  
833 involving realistic multi-metal mixtures and longer exposure periods covering complete crop  
834 cycles are needed to reflect actual wastewater irrigation scenarios. Screening of cultivars with  
835 low translocation potential and the application of mitigation strategies such as soil  
836 amendments, biochar, silicon-based materials, or microbial-assisted approaches should also be  
837 prioritized. Finally, future work should integrate region-specific dietary data and probabilistic  
838 health-risk models to improve the accuracy and relevance of food safety assessments for local  
839 populations.

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## 844 **8. Data Availability Statements**

845 All raw data, laboratory notebooks, and photographic documentation are archived under the  
846 institution's data retention policy and are available upon reasonable request.

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