

1 **Enhancing Zinc and Copper Stabilization Using Date Palm Waste Compost and the**  
2 **Phytoremediation Potential of *Medicago sativa***

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40 **Abstract**

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42 Heavy metal (HM) pollution poses a serious threat to plant growth and productivity. This study  
43 investigated the potential of date palm waste compost to enhance *Medicago sativa* tolerance to Zn  
44 and Cu stress and support phytoremediation. Plants were grown in sandy soil with or without 50%  
45 compost and irrigated with nutrient solutions containing high levels of Zn and Cu. Compost  
46 amendment significantly improved growth, increasing leaf, stem, and root biomass by 30%, 27%,  
47 and 30%, respectively. Despite elevated soil Zn and Cu levels, metal accumulation in leaves  
48 decreased, indicating reduced bioavailability. Compost also enhanced nutrient uptake (Ca, K, Mg,  
49 Fe) in both leaves and roots. Root bioconcentration factors increased, while shoot translocation  
50 factors decreased, suggesting improved phytostabilization. Compost treatments increased  
51 microbial colonies, with Zn and Cu addition boosting bacterial counts by 60% and 70%,  
52 respectively, though Cu reduced fungal numbers by 70%. Microbial diversity also shifted, notably  
53 in bacterial communities, due to the compost's high organic matter and carbon content. PCA  
54 analysis revealed distinct mineral distribution patterns across plant tissues under compost  
55 treatment. These findings highlight the potential of date palm compost to improve soil quality,  
56 support beneficial microbes, and enhance *M. sativa* phytoremediation capacity under heavy metal  
57 stress.

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59 **Key words:** Date palm waste compost, heavy metal stress, *Medicago sativa*, microbial biomass,  
60 phytoremediation

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75 **1.Introduction**

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Soil pollution with heavy metal (HM) has become the world's most severe environmental hazard, which induce growth reduction (Kumpiene et al., 2008). HM can reach the food chain through contaminated soil, water, and air, and their mobility among environmental compartments is also a great threat to human health (Emami Bistgani et al., 2019). The majority of HMs bind to fine-grained particles after entering the riverine environment and deposit in the sediment, causing adverse biological impacts (Bibi et al., 2007) (Brady et al., 2014). The availability of heavy metals (HMs) in soil is influenced by soil pH and the plant's absorption capacity (Shi et al., 2016). Copper (Cu) and zinc (Zn) are common soil pollutants that cause growth suppression, leaf chlorosis, and reduced photosynthesis in plants (Hattab et al., 2016). To prevent HM accumulation in plants and the food chain, various remediation approaches have been proposed, including physical, chemical, and biological methods (Silva et al., 2020). However, inorganic chemical methods may have long-term environmental impacts (Garau et al., 2007; Farrell and Jones, 2010). In contrast, phytoremediation offers a cost-effective, eco-friendly alternative with multiple benefits, such as plant biomass conversion to bioenergy, biodiversity preservation, and soil stabilization (Silva et al., 2020). Plants act as reservoirs for metals, potentially transporting them through the food chain (Hattab et al., 2016).

One of the many phytoremediation methods is phytostabilization, which involves using plants that can withstand metals to store the metals in their roots and rhizosphere. *Medicago* is a leguminous genus that contains economically important species. Among 83 species, *M. sativa* is an economically important leguminous plant that can thrive and tolerate a variety of environmental conditions (Raklami et al., 2021). Due to its quick growth, metal tolerance, its vigor roots to accumulate and ability to create a symbiosis with rhizobacteria, *M. sativa* has received interest in remediation strategies (Raklami et al., 2021). These plants serve as metal excluders, collecting metals mostly in the roots with low accumulation and transfer to the surface (Raklami et al., 2021). Moreover, it has become evident over the past decade that soil and rhizosphere microorganisms can significantly support phytoremediation efforts by enhancing HM bioavailability and facilitating plant uptake (Ahmad et al., 2019). Plants naturally interact with microorganisms, which can influence metal mobility, availability, and uptake. Microbial groups like mycorrhizal fungi and nitrogen-fixing bacteria help plants tolerate metal-contaminated soils by producing growth-promoting compounds and improving stress tolerance (Doornbos et al., 2012). To support plant

107 growth in such environments, amendments are often used, a strategy known as “assisted  
108 phytostabilization. HMs are immobilized in soil through precipitation, sorption, and complexation  
109 reactions (Kiikkilä et al., 2001). Organic amendments like compost are effective tools to reduce  
110 HM bioavailability and improve soil quality (Akram et al., 2018)Pulford and Watson, 2003).  
111 However, excessive use of compost can also lead to HM accumulation in soil and plants, posing  
112 risks to human health and the environment (Ghosh et al., 2015).

113 The date palm (*Phoenix dactylifera* L.) as an perennial fruit tree and alfalfa (*M. sativa* L.)  
114 as a perennial feed crop dominate the agriculture system in south Tunisia oases. Date palm wastes  
115 have a high potential for use as plant growth culture material in hydroponic systems. Compost has  
116 also been used for soil HMs immobilizing by moving speciation from highly bioavailable forms  
117 (i.e. free metals) to fractions associated with carbonates, metal oxides or organic matter (OM)  
118 (Mohammadi 2011, (Amirjani, 2011)), which are substantially less bioavailable (Walker and  
119 Bernal, 2008). These amendments are known to immobilize the HMs due to the humic acids in  
120 them, which can interact with a wide range of metal(loid)s (Madejón et al., 2018). Compost has  
121 long been used as an OA to immobilize HMs during soil remediation. While previous studies have  
122 demonstrated the potential of various composts such as those derived from leaves, straw, olive  
123 mill waste or municipal organic matter to enhance phytoremediation, these typically involve  
124 different organic amendments and soil conditions. In contrast, our study uniquely explores how  
125 date palm compost influences both metal immobilization and rhizosphere microbial dynamics,  
126 particularly under Zn and Cu contamination. For example, green waste compost application in  
127 lettuce reduced shoot Cu and Zn uptake by 22–59% and 16–25%, respectively (Ullah et al., 2025).  
128 Similarly, compost from rice straw improved soil pH, organic matter, and potassium availability  
129 while lowering HM bioavailability (Tang et al., 2020) and Olive-mill waste compost also enhanced  
130 shoot growth and decreased Cu, Zn, and Mn concentrations in both shoots and soil (Walker et al.  
131 2004). The novelty of our study lies in using *date palm waste compost*, abundant in arid regions  
132 like Tunisia, to improve phytoremediation outcomes. Unlike previous studies, we focus on how  
133 compost-mediated shifts in microbial biomass and diversity contribute to HM immobilization and  
134 plant stabilization. Specifically, we assess its ability to enhance Zn and Cu exclusion capacity of  
135 *M. sativa* and promote stable vegetation on contaminated soils. Our objectives were: (i) to evaluate  
136 *M. sativa* phytoremediation performance using date palm compost, and (ii) to study how Zn and  
137 Cu additions affect rhizosphere microbial communities in compost-amended soil.

138 **2. Materials and methods:**

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140 **2.1. Plant material and growth conditions**

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142 *M. sativa* seeds were obtained from the Institute of Arid Regions in Gabes. Following collection,  
143 the seeds underwent a 30-minute stirring process in a 4% NaClO solution, followed by three  
144 washes with deionized water. Subsequently, the sterilized seeds were placed in petri dishes and  
145 subjected to dark conditions at a temperature of 25°C for germination. After a week, the emerging  
146 *Medicago* saplings were transplanted into distinct pots. These pots were filled with a sandy soil  
147 derived from desert dune-sand to provide an environment conducive to further development. This  
148 soil was supplemented with certified high-quality compost obtained from date palm waste in one  
149 portion (CMP, 50% of the pot volume) (Table 1). This proportion was chosen based on its ability  
150 to enhance soil fertility, support microbial activity, and aid in the stabilization of heavy metals,  
151 particularly for phytoremediation. The 50% compost level balances enriching the soil with organic  
152 matter without negatively affecting plant growth or nutrient uptake. Additionally, this ratio has  
153 been shown to improve soil properties such as moisture retention, porosity, and microbial  
154 diversity, which are critical for *M. sativa* growth in metal-contaminated soils. The date palm leaves  
155 were first gathered from the Cheneni and Chenchou Oasis (Gabes, south region of Tunisia). It was  
156 physically crushed to a particle size of 5-10 cm before being soaked in tap water for 5–7 d in a 4  
157 m x 4 m basin with a depth of 2 m. The swelling of date palm waste improved their biodegradation  
158 during the composting process. The physicochemical properties of date palm waste compost are  
159 presented in Table 1. The moisture content was measured by drying samples at 95 °C for 24 h.  
160 Organic matter content in soil was calculated by multiplying Corg by 1.72, whereas OM in organic  
161 amendments was determined by ashing at 430 °C. Soil pH was measured in saturated paste  
162 extracts, while pH of organic amendments was determined in 1:10 (w/v) amendment-to-water  
163 extracts. Electrical conductivity (EC) was measured in 1:5 (w/v) aqueous extracts. Minerals were  
164 determined using atomic absorption spectrophotometer assay (Method 2.3). Total nitrogen was  
165 analyzed following the methods described by Walker and Bernal (2004). Greenhouse conditions:  
166 temperature  $25 \pm 2$  °C and light intensity (PAR): 500–700  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . Plants were irrigated every  
167 4 d with a complete nutrient solution. The metallic stress was imposed after a month of  
168 acclimatization by adding the doses of Zn and Cu in the irrigation solution to 6 and 4 mM,  
169 respectively. To avoid osmotic shock, the Zn or Cu dose was gradually increased by 2 mM each

170 day until the appropriate concentrations were achieved. The stability and uptake of Zn and Cu in the  
171 soil–plant system was confirmed by analyzing metal concentrations in soil and plant tissues using  
172 atomic absorption spectrophotometer (Method 2.3). The experimental design was conducted with  
173 8 seedlings in five pots for each treatment. Four weeks after beginning the treatments, these plants  
174 were harvested. The leaves were weighed and then separated into two groups. The initial group  
175 was preserved at 80°C for subsequent biochemical analyses. Meanwhile, the second group  
176 underwent a drying process at 80°C for 48 hours, after which it was finely powdered to facilitate  
177 ion analyses.

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## 179 **2.2. Soil and compost processing, and characterization**

180 The pH of the soil suspension was read with the aid of a pre calibrated pH meter (*ISO 10390*). 20  
181 g compost was stirred in distilled water to measure the EC by a conductivity meter (*ISO 11265*).  
182 Dry matter content was measured through the gravimetric method, drying the sample at  $105 \pm 2^\circ\text{C}$   
183 in hot air oven until fulfilling constant mass. Moisture content was determined as the weight  
184 difference between fresh and dry matter. After calcining a dried sample at  $550^\circ\text{C}$  for 4 hours in a  
185 muffle furnace, the organic OM was measured. The amount of OC was calculated according to  
186 numerous researchers,  $\text{OC \%} = \text{OM \%}/1.8$  (Kolka et al., 2008). The total nitrogen (N) was  
187 determined using the Afnor standard Kjeldahl method (Sáez-Plaza et al., 2013).

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## 189 **2.3. Analysis of mineral contents**

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191 Two hundred mg of each *M. sativa* sapling (leaves, stem and roots) was digested in solution  
192 containing  $\text{HNO}_3/\text{H}_2\text{O}$  for evaluation of macro and micro-elements. The concentrations of  
193 minerals elements (Ca, Na, K, Mg, Zn and Cu) were determined with a standard of 1% nitric acid  
194 using an atomic absorption spectrophotometer (Abdelgawad et al., 2015, Alotaibi et al., 2021, Saleh  
195 et al., 2020). The mineral composition of soil and soil mixed with compost was measured using  
196 atomic absorption spectrophotometer.

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## 198 **2.4. Heavy Metal Translocation Index**

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200 **2.4.1. Translocation factor (TF):** This factor indicates the transfer of Zn or Cu from the soil to  
201 the root, and from the root to the shoot (Phusantisampan et al., 2016):

202 
$$TF = C_{\text{shoot}} / C_{\text{root}}$$

203 where  $C_{\text{root}}$  and  $C_{\text{shoot}}$  is the Zn or Cu concentration in soil, shoot or root.

204 **2.4.2. Bioconcentration coefficient (BCFR):** it implies the accumulation of heavy metal by  
205 roots (Phusantisampan et al., 2016):

206 
$$BCFR = C_{\text{roots}} / C_{\text{soil}}$$

207 where  $C_{\text{soil}}$  and  $C_{\text{root}}$  indicate is the concentration of Zn or Cu in soil or root.

208

## 209 **2.5. Bacteria and fungi count**

210 Microbial counts of bacteria and fungi were conducted from 1 g of soil or compost sample  
211 suspended in 9 mL of sterile 0.9% NaCl solution. A tenfold serial dilution series (ranging from  
212  $10^{-1}$  to  $10^{-6}$ ) was prepared to reduce microbial concentrations to countable levels. For bacterial  
213 enumeration, 100  $\mu\text{L}$  aliquots from the  $10^{-4}$  dilution were plated onto Plate Count Agar (PCA).  
214 The plates were incubated at  $28^{\circ}\text{C}$  for 48 hours, after which colony-forming units (CFUs) were  
215 counted. For fungal enumeration, a preliminary test was performed to determine the optimal  
216 dilution range. Based on these trials, the  $10^{-2}$  dilution was selected. From this dilution, 100  $\mu\text{L}$   
217 aliquots were plated onto Potato Dextrose Agar (PDA), supplemented with chloramphenicol to  
218 suppress bacterial growth. Plates were incubated at  $25^{\circ}\text{C}$  for 75 hours. These dilution factors ( $10^{-4}$   
219 for bacteria and  $10^{-2}$  for fungi) was selected based on preliminary trials to ensure the optimal  
220 countable range of colonies. Incubation times and temperatures were chosen to maximize the  
221 growth of both bacterial and fungal communities, as recommended by previous studies (Lionel  
222 Ranjard , Sylvie Nazaret , Francois Gourbiere et al., 2000). The soil microbial count represents the  
223 number of colony forming units (CFUs) per gram of dry soil (Lionel Ranjard , Sylvie Nazaret ,  
224 Francois Gourbiere et al., 2000).

## 225 **2.6. Statistical analyses:**

226 The statistical analyses were carried out using the R statistics software (ggplot2, agricolae\_1.3-5).  
227 One-way analysis of variance (ANOVA) was performed, using Tukey's Test (P 0.05) as a post-  
228 hoc test for mean separations. Prior to performing the one-way analysis of variance (ANOVA),  
229 data normalization was applied to ensure comparability across treatments and eliminate any  
230 potential scale-related biases. The number of replicates per treatment was consistently maintained  
231 at five ( $n = 5$ ) for all experiments, ensuring the reliability of the data. Each experiment was repeated

232 at least three times to ensure robust and reproducible results. All data were measured in appropriate  
233 units, which are explicitly stated in the results section. Pearson correlation analysis was also  
234 performed on all experimental data using the `corrplot_0.92` package in R, and hierarchical  
235 clustering was constructed to group similar data points. Principal component analysis (PCA) was  
236 also carried specifically to reduce the dimensionality of the data, allowing for the visualization of  
237 underlying trends and interactions. By using R (`ggplot2`, `agricolae_1.3-5`), we were able to  
238 visualize how different treatments related to each other and to other experimental factors, helping  
239 to highlight the key influences on the system. Additionally, correlation analysis was used to  
240 quantify the strength and direction of relationships between key variables.

241

### 242 **3. Results:**

#### 243 **3.1. Zn and Cu effect on the study soils:**

244 In comparison to the sandy soil, the pH of the soil in composted pots was dramatically increased  
245 (around neutral, pH 7.4, Table 2). These increases can be explained by increased Zn and Cu doses  
246 that raised the pH of the soil from 6.8 to 7.8 and 8, respectively. In pots treated with 50 % of date  
247 palm waste compost, increased Zn lowered the soil pH, while higher Cu increased pH of former  
248 soil. The highest value of EC (1.74 dS/m) was recorded for soil mixed by compost and Cu. The  
249 highest moisture content (23%) was recorded for soil combined with compost. The high doses of  
250 Zn and Cu did affect soil moisture. When compost was applied instead of just soil, organic matter  
251 increased to 46%. In all types of soils, Zn and Cu reduced organic matter, but it still higher in soil  
252 mixed with compost (29% and 32%). Total N increased by 35% in soil amendment with palm  
253 waste compost. Compared to the single effect of Zn or Cu, their interaction with compost boosted  
254 total nitrogen by 50% and 60%, respectively. Date palm waste compost increased the OC carbon  
255 to 25%. But the addition of Zn and Cu decreased the OC in target soils.

256

#### 257 **3.2. The impact of compost, Zn and Cu on soil mineral composition**

258 The compost application to the soil enriched it with high concentrations of Ca, K, Mg, Fe, and Zn.  
259 (Figure 1). The application of compost resulted in a significant increase in the levels of Ca, K, Na,  
260 and Mg by 20%, 205%, 50%, and 15% respectively. However, the Cu content appeared to remain  
261 unchanged with the use of compost. As a result, the introduction of a high Zn concentration led to  
262 an increase in the Zn content in sandy soil, whereas in compost, the Zn level was lower. Notably,

263 the presence of high Zn did not lead to a reduction in the levels of K and Fe in compost compared  
264 to sandy soil. Following a similar trend, the inclusion of Cu results in a greater increase in the  
265 content of Ca, K, Mg, and Fe in sandy soil compared to compost.

266

### 267 **3.3. The effect of compost, Zn and Cu on biomass accumulation of *M. Sativa*:**

268 The application of compost sharply increased biomass of leaves, stem and roots of plants by 30%,  
269 27% et 30%, respectively (Figure 2). But Zn and Cu addition decreased the biomass of all  
270 *Medicago* organs in both types of soil. In fact, the highest concentration of Zn caused a significant  
271 decrease in leaves biomass by 50% as compared to control. The addition of compost mitigates the  
272 decrease in leaf biomass. Cu decreased the leaves biomass by 70% in sandy soil, but compost  
273 decreased it by 50% as compared to control. The fresh weight of stems reduced by 60% in response  
274 to high Zn and Cu concentrations, whereas the reduction did not reach 40% in plants grown in soil  
275 amended with compost. In addition, roots biomass showed the greatest decline in response to  
276 increasing dosages of Zn and Cu.

277

### 278 **3.4. The influence of compost, Zn and Cu on *M. sativa* mineral nutrition**

279 Fe, Mg and K concentrations in leaves of plants cultivated with compost increased by 47% 17%,  
280 and 25%, respectively (Figure 3). The absorption of mineral elements was altered by the uptake of  
281 Zn and Cu concentrations. For instance, Ca, K, Mg, Fe and Na content reduced in all *Medicago*  
282 organs in response to high doses of Cu and Zn. Compost reduced mineral level, while the Zn and  
283 Cu accumulation was increased. The findings demonstrated that roots significantly accumulated  
284 more Cu and Zn than roots of plants cultivated in soil.

285

### 286 **3.5. Translocation Index:**

287 TF and coefficients represent *Medicago*'s ability to take up and accumulate Zn and Cu into shoots  
288 and roots. TF values of *Medicago* saplings in sandy soil treated with Zn were less than 1, and  
289 declined but remained higher level in soil mixed with compost (0.78, Table 3). However, Cu  
290 reduced TF values for roots, except for Cu treatment. TF values below 1 indicate that Zn or Cu is  
291 retained in the roots, preventing translocation to the shoots, which is beneficial for  
292 phytoremediation. In *Medicago* saplings grown in sandy soil with Zn, TF values were below 1,  
293 and further reduced in compost-amended soil (0.78, Table 3), suggesting that compost helps limit

294 metal mobility within the plant. Compost treatment decreased BCFR to 0.39. Compared to all  
295 types of Zn treated soil (BCFR values >1), a high dose of Cu decreased BCFR to 0.39 in soil mixed  
296 with date palm waste compost. Compost treatment decreased BCFR to 0.39, indicating reduced  
297 metal transfer from roots to shoots. In contrast, all Zn-treated soils had BCFR values greater than  
298 1, while a high dose of Cu reduced BCFR to 0.39 in compost-amended soil, suggesting that  
299 compost limits metal translocation in the plant.

300

### 301 **3.6. Bacteria and fungi count:**

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303 Compost treatments had more microbial colony forming than the control (Figure 4), which was  
304 sandy soil (the data was not presented because no microbial colony forming was detected).  
305 Furthermore, adding Zn and Cu to soil mixed with compost significantly enhanced microbial  
306 colony forming by 60% and 70%, respectively. Furthermore, Zn had no effect on the colony  
307 forming of the fungi. As a result of the Cu application, the total fungus count decreased by 70%.

308

### 309 **3.7. Principal Component Analysis (PCA) indicated specific organ and treatment responses**

310 PCA was employed to investigate the variation in mineral composition and microbial community  
311 structure among different plant organs and treatment conditions. PCAs of samples taken from  
312 plants cultivated in two types of soil contaminated with Zn and Cu revealed a very comparable  
313 distribution of the seven elements in the roots and stem of the *Medicago* seedling cultivated under  
314 the different four treatments. The mineral composition of *Medicago* seedling leaves, on the other  
315 hand, moved from other organs in plants grown in soil mixed with compost (Figure 5). In addition  
316 to mineral composition, PCA was also used to explore the relationships between environmental  
317 variables, bacterial communities, and fungal populations (Figure 6). A clear separation of bacterial  
318 and fungal diversity was observed between soil treatments with compost and its interactions with  
319 Zn and Cu, with the heavy metals being separated along Principal Component 1 (PC1). This  
320 suggests that compost application significantly influenced microbial community composition in  
321 the soil, particularly in the presence of heavy metals. Notably, bacterial diversity showed a distinct  
322 separation based on OM and OC content, likely due to the high OM and OC levels in the compost.  
323 In the organic and control treatments, bacterial and fungal populations were highly comparable.  
324 This indicates that compost not only serves as a source of nutrients but also plays a crucial role in  
325 shaping the soil microbiome, particularly the bacterial communities. Bacterial communities

326 showed greater variability in compost-treated soils compared to fungal populations, indicating a  
327 stronger response to the compost and heavy metals. In contrast, microbial populations in the  
328 organic and control treatments were similar, highlighting the impact of compost and heavy metals  
329 on microbial diversity. These PCA results underscore the complex interactions between soil  
330 amendments, metal contamination, and microbial communities, emphasizing the need to consider  
331 both mineral composition and microbial diversity in evaluating soil amendments' effectiveness.

332

### 333 **3.8. Chemical characteristics and biological activity correlations:**

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335 According to Pearson's correlation coefficients between biological and chemical factors (Figure  
336 7), pH was the strongest predictor, with substantial correlations between all variables. MO and OC  
337 were negatively connected ( $P < 0.05$ ), while Cu and Zn were positively correlated ( $P < 0.01$ ). MO,  
338 OC, Ca and K are all favorably associated with total fresh weight. HMs concentrations were highly  
339 found inversely associated to both bacterial and fungal biomass. In compost treated soil, however,  
340 both bacterial and fungal biomass were adversely linked with MO, OC and N ( $P < 0.05$ ).

341

## 342 **4. Discussion**

343

344 Date palm waste compost has attracted considerable research interest due to its beneficial effects  
345 on soil structure, functioning, and its role as an effective fertilizer (Ali, 2008). In this study, the  
346 use of date palm waste compost significantly improved the growth of *M. sativa*. A previous study  
347 found that *M. sativa* exhibited a higher survival rate when planted in compost-amended soil  
348 compared to control soil (Marchand et al., 2017). Green compost individually or in combination  
349 with arbuscular mycorrhizal fungi as an effective strategy to improve alfalfa productivity under  
350 stress conditions (Ben-Laouane et al., 2020). In agreement with our results, soil amendments have  
351 been shown to enhance plant growth and yields in various species, such as barley (Agegnehu et  
352 al., 2016) and poplar trees (Guarino et al., 2020). Furthermore, increased application rates of date  
353 palm waste compost positively influenced the growth characteristics of *M. sativa*, likely due to  
354 improved nutrient mineralization and organic matter decomposition (Rady et al., 2016). The  
355 increase in *Medicago* yield appeared to be linked to mineral nutrient absorption. According to our  
356 findings, applied compost can operate as a nutrient reservoir, allowing nutrients to gradually  
357 release to roots of plant or into the soil (Yahaghi et al., 2019). Compost, rich in nutrients, humus,

358 and microorganisms, positively affects soil biota metabolism, nutrient uptake, soil properties, and  
359 enzymatic activity, which enhances plant growth and productivity (Farrell and Jones, 2010). In our  
360 study, compost, either alone or combined with Zn or Cu, significantly improved *Medicago* growth,  
361 suggesting it can mitigate Zn and Cu uptake and toxicity. The increased organic matter OM,  
362 nutrient concentrations, and OC in date palm compost were linked to improved *M. sativa* growth.  
363 Our study uniquely explores how date palm compost influences both metal immobilization and  
364 rhizosphere microbial dynamics. Similarly, rice straw compost improved soil pH, organic matter,  
365 and potassium availability while reducing heavy metal bioavailability. Olive-mill waste compost  
366 also enhanced shoot growth and decreased Cu, Zn, and Mn concentrations in both shoots and soil  
367 (Tang et al., 2020, Walker et al. 2004).

368  
369 Compost also indirectly boosts plant growth by influencing soil microbial activity (Rady et al.,  
370 2016). *M. sativa* ability to phytostabilize and phytoextract Cu and Zn has been well-documented  
371 (Baldantoni et al., 2014). TF and BCFR are critical indicators of metal uptake, with  $TF > 1$  and  
372  $BCFR > 1$  signaling efficient metal translocation and accumulation. According to Meeinkuirt et  
373 al. (2012), TF values of 1 and  $BCFR > 1$  indicate good metal tolerance. Our results showed BCFR  
374 values above 1, indicating effective metal uptake, while TF values for Zn in sandy soil were below  
375 1, suggesting limited translocation. The addition of compost further reduced TF, implying better  
376 Zn retention in roots. This is consistent with the role of compost in enhancing metal  
377 immobilization. While *M. sativa* did not meet the hyperaccumulator threshold ( $TF > 1$ ,  $BCFR >$   
378  $10$ ), its ability to concentrate metals in roots and improve soil retention through compost suggests  
379 strong potential for phytostabilization, similar to findings in other studies (e.g., Baldantoni et al.,  
380 2014). This suggests the potential for phytostabilization, where compost helps stabilize Zn in roots  
381 and limits its transport to the stem and leaves (Taepayoon et al., 2022), (García Martín et al.,  
382 2020) Concerning the phytomanagement of Cu, it's likely that *M. sativa* has a higher ability to  
383 tolerate and accumulate Cu than other Cu-bioaccumulators like *Brassica juncea* (Qadir et al.,  
384 2004) and could be used for phytoremediation (Rosca et al., 2021). Moreover, Brassica species are  
385 also effective for phytoattenuation, particularly for reducing the bioavailability of HMs in  
386 contaminated soils through phytoextraction (Meers et al., 2010).

387 *M. sativa* responded differently to Cu and Zn in compost-amended soils. BCFR values were lower  
388 in these soils, and TF approached 1, indicating limited metal translocation. The roots accumulated  
389 10 times more Cu than *M. indica*, confirming ability of *M. sativa* to extract Cu (Bélanger, 1992).  
390 This is likely due to metal binding in root cell walls and storage in vacuoles, mechanisms used to  
391 avoid metal toxicity (Krzesłowska, 2011). Although *M. sativa* transferred Cu to aerial parts, the  
392 low leaf Cu content suggests it acts as a Cu excluder. The addition of date palm waste compost  
393 enhanced this exclusion. Soil pH, OM content, and nutrient availability influence Cu and Zn  
394 phytoavailability (Tariq Rafiq et al., 2014). The compost increased pH (7.5–7.7), reducing metal  
395 bioavailability by enhancing metal sorption (Sukreeyapongse et al., 2002; Kazlauskaitė-  
396 Jadzevičė et al., 2013). The high OM content in the compost further decreased metal availability,  
397 promoting phytostabilization (Cornu et al., 2016; Stewart et al., 2008). These findings indicate that  
398 *M. sativa*, in compost-amended soils, enhances phytostabilization, especially for Cu, by improving  
399 soil quality and reducing metal toxicity.

400 Compost significantly enhanced soil microbial diversity, particularly among bacteria and fungi,  
401 crucial for nutrient cycling and plant growth. These microbial shifts are key indicators of soil  
402 quality and fertility changes, often occurring faster than organic matter content. Compost, rich in  
403 beneficial microorganisms, boosts nutrient uptake, soil structure, and enzymatic activities,  
404 promoting plant productivity (Farrell et al., 2010; Rady et al., 2016). The increased microbial  
405 counts result from the carbon substrate and microorganisms in compost, with date palm waste  
406 providing nutrients that support beneficial microbes (Ren et al., 2018). Despite the presence of Zn  
407 and Cu, microbial biomass remained stable, suggesting compost improved microbial resistance to  
408 heavy metal stress (Azarbad et al., 2013). In compost-enriched soils, Zn and Cu had a minimal  
409 impact on microbial community structure. The higher microbial counts in contaminated soils likely  
410 stem from compost's high nitrogen and carbon content. However, the microbial balance shifted:  
411 bacterial resistance improved, while fungal communities became more sensitive. These shifts have  
412 ecological implications, affecting nutrient competition and soil health (Fones and Preston, 2013;  
413 Hudek et al., 2013). Thus, compost not only supports plant growth under contamination but also  
414 reshapes microbial consortia, enhancing phytoremediation by improving plant–microbe  
415 interactions (Farrell et al., 2010; L. Wood et al., 2016; Kamran et al., 2017). This study shows that  
416 applying date palm waste compost to contaminated soils improves soil health, promotes microbial

417 diversity, and helps immobilize metals, reducing their bioavailability. Compost-assisted  
418 phytoremediation is a promising strategy for restoring contaminated soils while maintaining  
419 ecological stability.

420 Overall, the present study highlights the viability of integrating locally available organic waste,  
421 such as date palm compost, into phytomanagement strategies for metal-contaminated soils. This  
422 approach not only enhances plant growth and soil microbial activity but also contributes to metal  
423 stabilization, offering a low-cost and environmentally friendly alternative to conventional  
424 remediation methods. The use of agricultural waste within a circular bioeconomy framework  
425 aligns with sustainable development goals and has strong potential in arid and semi-arid regions,  
426 where resource recovery and soil rehabilitation are critical. On the other hand, this study was  
427 conducted under controlled conditions and for a limited duration, which may constrain the  
428 applicability of results to long-term, field-scale scenarios. While changes in microbial abundance  
429 were observed, deeper functional insights (*e.g.*, metagenomics) are needed. Nonetheless, our  
430 findings support the combined use of organic compost and phytoremediation as a promising  
431 strategy for soil restoration. Future research should focus on field validation, economic feasibility,  
432 and life-cycle analysis to advance this sustainable approach.

433

## 434 **5. Conclusion:**

435 The use of date palm waste compost amendment to *Medicago sativa* saplings reduced Zn  
436 translocation and increased its retention by roots. The compost enhanced Cu phytoextraction by  
437 roots and its immobilization, likely due to its high organic matter content, organic carbon, and  
438 lower pH. The microbial consortia in the compost, particularly fungi, also contributed to improved  
439 phytoremediation outcomes. This compost-based strategy, which benefits from enhancing soil  
440 structure and microbial diversity while stabilizing metals, presents a promising and cost-effective  
441 approach for soil remediation. Its applicability extends beyond *M. sativa*, offering a sustainable  
442 solution for contaminated soils, especially in arid and semi-arid regions with abundant date palm  
443 waste.

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447 **6. References:**

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627 **Figures captions:**

628 **Figure 1:** The effect of Zn and Cu on mineral composition sandy soil and soil mixed with compost. SS: Sandy Soil,  
629 SC: Sandy Soil + Compost, SS+Zn: Sandy Soil treated with Zn, SS+Cu: Sandy Soil treated with Copper (Cu), SC+Zn:  
630 Compost-amended Soil treated with Zinc, SC+Cu: Compost-amended Soil treated with Cu .Values are represented by  
631 mean  $\pm$  standard deviation of at least three independent replicates. Within the same organs, different letters on the bars  
632 indicate significant differences at  $p < 0.05$ .

634 Figure 2: The effect of compost, Zn, and Cu on the growth parameters (roots (g/plant), stems (g/plant), leaves  
635 (g/plant), and shoot/root ratio) of *Medicago sativa*. Treatments include SS: Sandy Soil, SC: Sandy Soil + Compost,  
636 SS+Zn: Sandy Soil treated with Zn, SS+Cu: Sandy Soil treated with Copper (Cu), SC+Zn: Compost-amended Soil  
637 treated with Zinc, and SC+Cu: Compost-amended Soil treated with Cu. Values are represented as the mean  $\pm$   
638 standard deviation of at least three independent replicates. Within the same species, different letters on the bars  
639 indicate significant differences at  $p < 0.05$ .

640 Figure 3: The effect of compost, Zn, and Cu on the mineral composition (mg/g DW) of the roots, stems, and leaves  
641 of *Medicago sativa*. Treatments are the same as described in Figure 2. Values are represented as the mean  $\pm$  standard  
642 deviation of at least three independent replicates. Within the same organs, different letters on the bars indicate  
643 significant differences at  $p < 0.05$ .

644 Figure 4: The effect of compost, Zn, and Cu application on bacterial and fungal biomass (CFU/gDW). The  
645 treatments are as follows: SS: Sandy Soil, SC: Sandy Soil + Compost, SS+Zn: Sandy Soil treated with Zn, SS+Cu:  
646 Sandy Soil treated with Copper (Cu), SC+Zn: Compost-amended Soil treated with Zinc, SC+Cu: Compost-amended  
647 Soil treated with Cu. Values are represented as the mean  $\pm$  standard deviation of at least three independent  
648 replicates. Within the same treatment, different letters on the bars indicate significant differences at  $p < 0.05$ .

649 Figure 5: Principal Component Analysis (PCA) Biplot depicting the relationship of various parameters from the  
650 leaves (A), stems (B), and roots (C) of *Medicago sativa*.

651 Figure 6: Correlation analysis of variance between Organic Matter (OM), Organic Carbon (OC), Nitrogen (N), pH,  
652 heavy metals, essential elements, and total fresh weight of *Medicago sativa* saplings. The blue and red colors  
653 represent positive ( $0 < r < 1$ ) and negative ( $-1 < r < 0$ ) correlations, respectively.

654 Figure 7: Correlation analysis of variance between Organic Matter (OM), Organic Carbon (OC), Nitrogen (N), pH,  
655 heavy metals, essential elements, and microbial and fungal biomass. The blue and red colors represent positive  
656 ( $0 < r < 1$ ) and negative ( $-1 < r < 0$ ) correlations, respectively.

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Parameters	Date palm waste	Soil
pH	7.7±0.3	6.8 ±0.04
EC (mS/cm)	5.9±0.04	0.92± 0.03
Moisture (%)	9.9±0.52	2.12±0.1
Dry matter (%)	63±0.50	18.7±4.0
OM(%)	25.7±0.2	4.4±0.2
OC(%)	52.6±0.3	10±0.23
N (%)	0.6±0.01	0.1±0.02
K (mg/g dw)	18.2±0.2	15.6± 0.2
Ca ( mg/g dw )	20.9± 1.1	8.6± 0.03
Mg ( mg/g dw )	3.0± 0.3	3.4± 0.07
Na ( mg/g dw )	10.9±1.0	3.5 ±0.1
Fe ( mg/g dw )	1.75± 0.1	2.0±0.00

**Table 1** : Main Physical and Chemical Properties Measured in Date Palm Waste Compost and Soil: pH, EC, Moisture, Organic Matter, and Nutrient Content

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pH, EC (Electrical Conductivity), Moisture (%) (Moisture Content), Dry Matter (%) (Dry Matter Content), OM (%) (Organic Matter Content), OC (%) (Organic Carbon Content), N (%) (Nitrogen Content), K (mg/g dw) (Potassium Content), Ca (mg/g dw) (Calcium Content), Mg (mg/g dw) (Magnesium Content), Na (mg/g dw) (Sodium Content), and Fe (mg/g dw) (Iron Content).

719 **Table 2:** Physico-chemical properties of sandy soil and compost-amended soil treated with zinc  
 720 (Zn) and copper (Cu).  
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	SS	SC	SS+Zn	SS +Cu	SC+ Zn	SC +Cu
<b>pH</b>	6,85±0.1c	7,43±0.00b	7,91±0.04a	8,01±0.05a	7,79±0.00b	7,59±0.01b
<b>EC (dS/m)</b>	0,92±0.01b	1,61±0.02a	0,98±0.01b	1,05±0.00b	1,68±0.00a	1,74±0.00a
<b>Moisture (%)</b>	2.14±0.00d	23,06±0.2a	2,26±0.00d	3,00±0.01c	14,68±0.2b	18,12±0.2b
<b>OM (%)</b>	3.12±0.01d	46,5±0.3a	4,04±0.02c	3,11±0.01d	29,41±0.5b	32,16±0.4a
<b>Total azote (%)</b>	0.18±0.00c	0,35±0.00b	0,2±0.00c	0,22±0.00b	0,91±0.00a	0,87±0.00a
<b>OC(%)</b>	1.73±0.01d	25,83±0.2a	2,24±0.00c	1,72±0.00d	17,16±0.01b	17,87±0.3b

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 727 OC: Organic Carbon, TN: Total Nitrogen, OM: Organic Matter, SS: Sandy Soil, SC: Sandy Soil + Compost, SS+Zn:  
 728 Sandy Soil treated with Zn, SS+Cu: Sandy Soil treated with Copper (Cu), SC+Zn: Compost-amended Soil treated  
 729 with Zinc, SC+Cu: Compost-amended Soil treated with Cu. The superscript lowercase letters (a- d) indicate  
 730 statistically significant differences between the samples. The statistical significance of the relative abundances was  
 731 determined by Tukey's post hoc test. with  $p < 0.05$  (n=5)  
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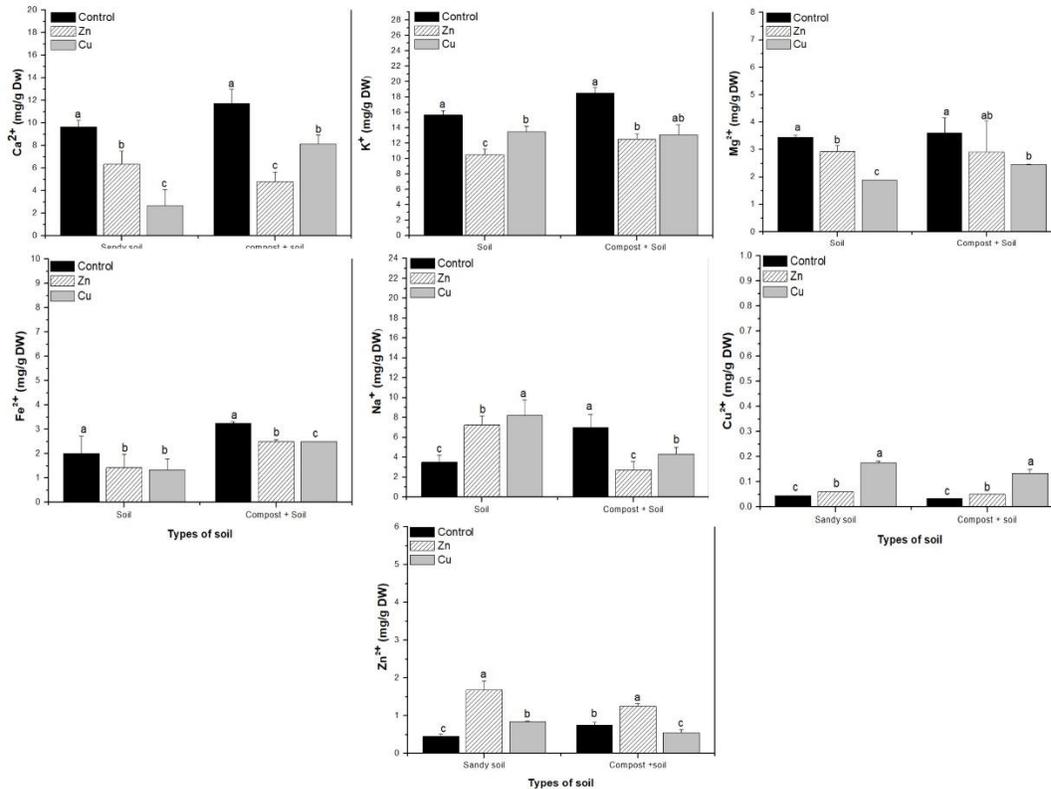
752 **Table 3:** Translocation factor (TF) and bioconcentration coefficient for roots (BCFR) in the  
 753 *Medicago satvina*.

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 756 **TF:** Ratio of metal concentration in shoots to roots, indicating the plant's ability to move metals upwards.  
 757 **BCFR:** Ratio of metal concentration in roots to soil, showing how much metal is absorbed by the roots. The  
 758 superscript lowercase letters (a-d) indicate statistically significant differences between the samples. The statistical  
 759 significance of the relative abundances was determined by Tukey's post hoc test, with  $p \leq 0.05$  (n=5).  
 760

		TF	BCFR
<b>Sandy Soil</b>	<b>Zn</b>	1.39±0.10a	1.00±0.17a
	<b>Cu</b>	0.31±0.07c	0.42±0.02b
<b>Soil + Compost</b>	<b>Zn</b>	0.70±0.07b	1.27±0.13a
	<b>Cu</b>	0.15±0.01d	0.39±0.04b

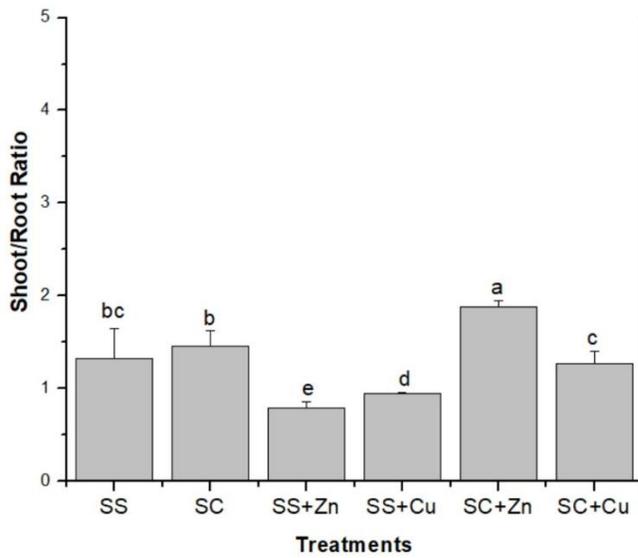
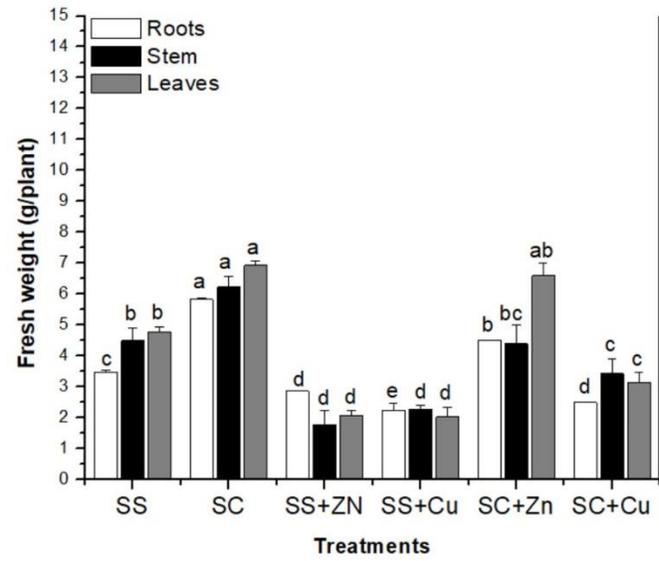
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Figure 1 :



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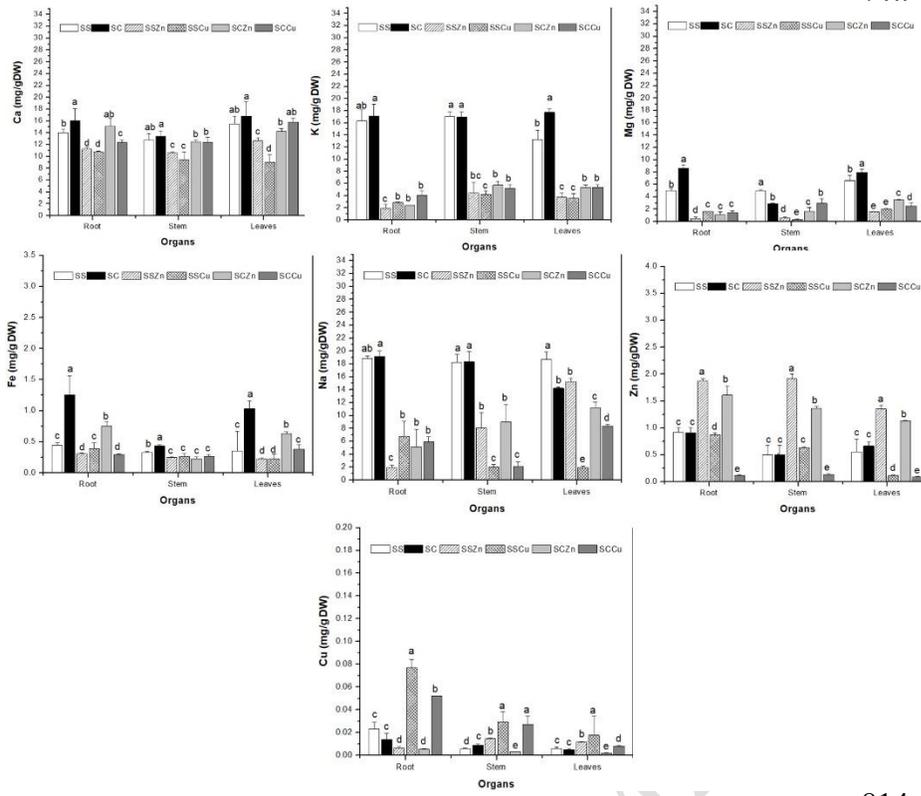
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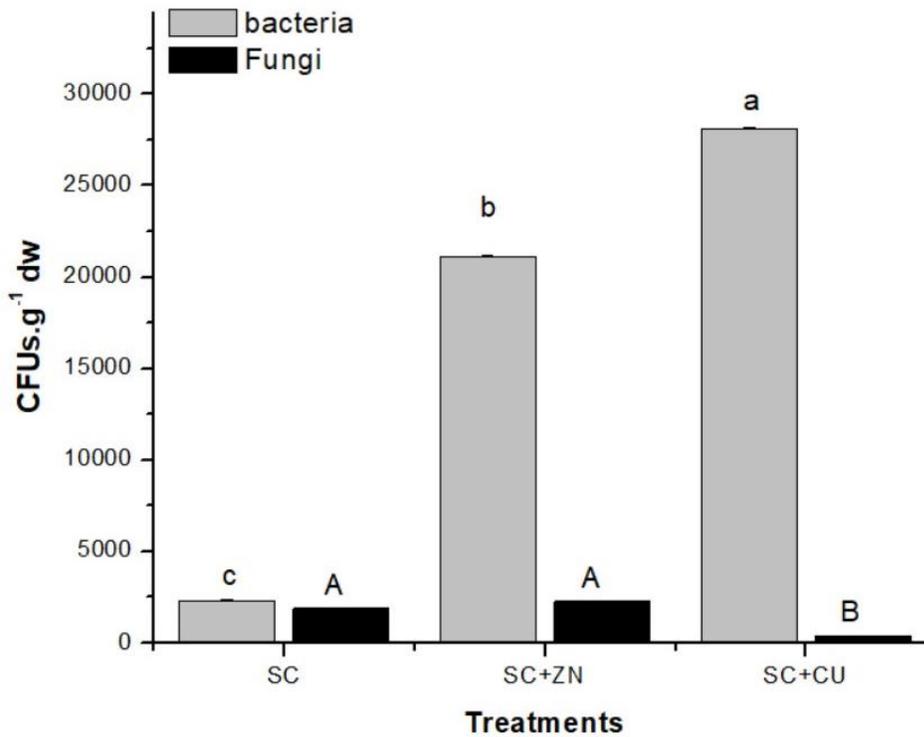
Figure 3 :



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Figure 4:

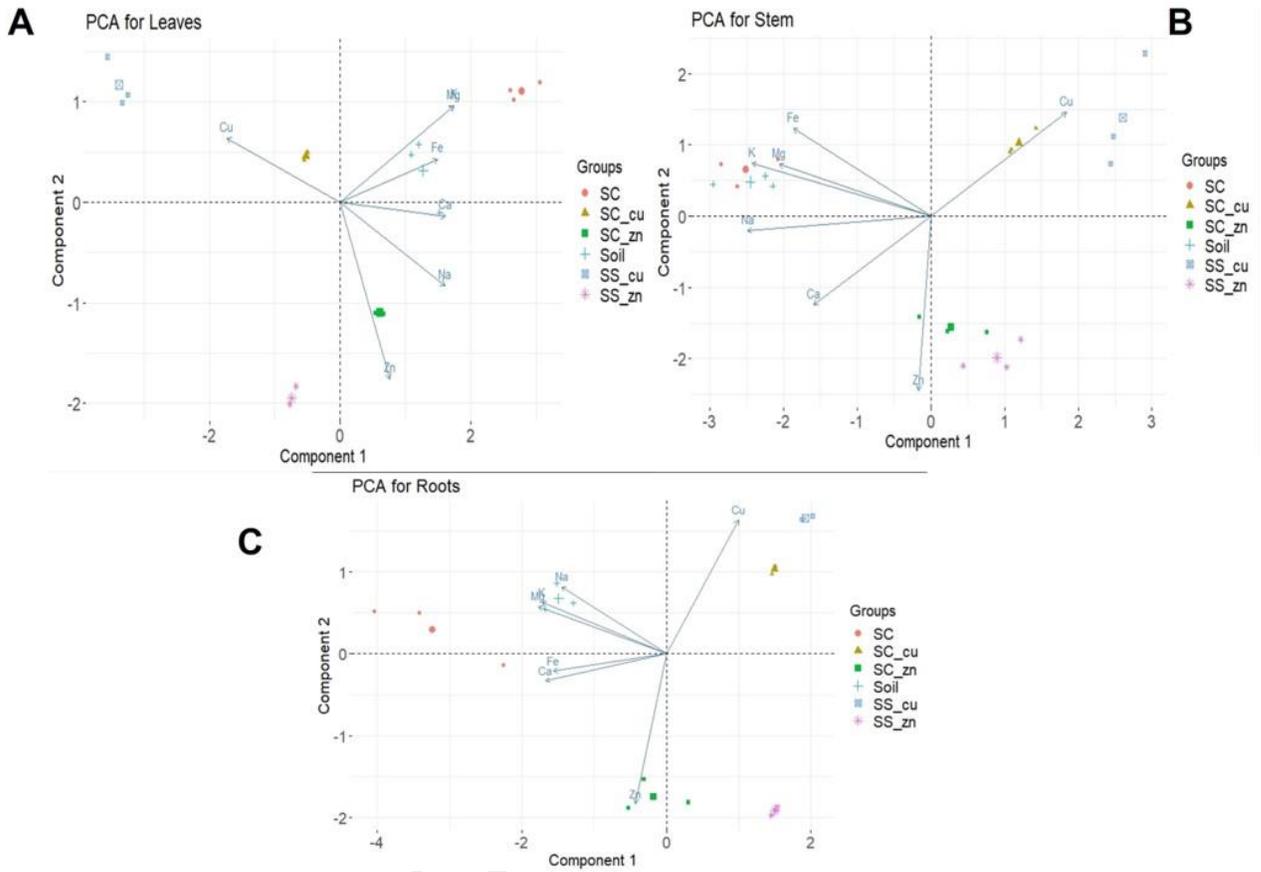


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Figure 5 :



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