

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19

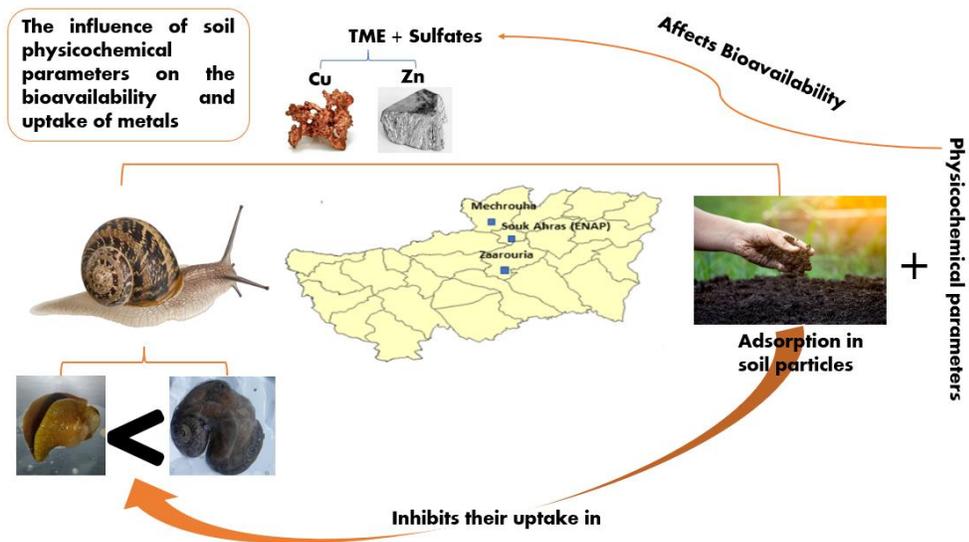
Biomonitoring of soil quality in the Souk Ahras region (Northeast Algeria) using *Cornu aspersum* as a bioindicator: Trace metals and sulfates analysis

Roumaissa BENDJEBBAR^{1*}, Houda BERROUK¹, Choukri BAROUR¹,
Noomene SLEIMI², Amina MERGHAD¹

¹ Laboratory of Aquatic and Terrestrial Ecosystems (LEAT), Department of Biology,
Faculty of Natural and Life Sciences, Mohamed Chérif Messaâdia University, BP1553,
Annaba Road, Souk Ahras, Algeria.

² Research Laboratory RME – Resources, Materials, and Ecosystems, Faculty of
Sciences, Bizerte,
University of Carthage, Tunisia.

r.bendjebbar@univ-soukahras.dz*
h.berrouk@univ-soukahras.dz
c.barour@univ-soukahras.dz
noomene.sleimi@gmail.com
a.merghad@univ-soukahras.dz



20 **Abstract.** The present study aims to assess the impact of physicochemical
21 parameters on the availability of trace metal elements (TME) in soil by
22 analyzing the accumulation of copper (Cu), zinc (Zn) and sulfates in both
23 soil and edible tissues of *Cornu aspersum* using a passive biomonitoring
24 approach at three sites in Souk Ahras province: Zaarouria, Mechrouha and
25 ENAP (National Paint Enterprise). Statistical analysis of the data indicated
26 that adsorption of TME and sulfates on soil particles inhibits their uptake
27 by other terrestrial organisms such as snails. In addition, physicochemical
28 parameters significantly influenced the distribution of TME in the soil,
29 revealing marked spatial differences in the accumulation of Cu in the foot
30 and viscera as well as Zn especially in the viscera. Furthermore, a
31 significantly higher accumulation of sulfates was observed in the foot
32 compared to the viscera especially at the ENAP site. These observations
33 could confirm the potential of *C. aspersum* species as an effective
34 bioindicator of metal pollution in soil.

35 **Keywords.** *Cornu aspersum*, bioindicator, bioavailability, accumulation,
36 metal pollution, soil.

37 **1. introduction**

38 One of the main areas of current research is environmental pollution
39 caused by the release of different types of pollutants into water, air

40 and soil (Baroudi 2020, N'Doua et al. 2024). These different
41 pollutants originate from the expansion of human activities and
42 chemical emissions, notably trace metal elements (TME) associated
43 with industrial and agricultural needs (Dossou et al. 2022, Amlan et
44 al. 2023; Hidouri et al. 2024). TME occur naturally in the
45 environment and are considered among the most harmful pollutants,
46 as they are non-biodegradable and their concentration is constantly
47 increasing in different ecosystem compartments (Ibrahim et al.
48 2019, Ning et al. 2024). Due to their excessive presence in the
49 ecosystem, heavy metals can also have a direct impact on the life
50 cycles of animal and plant organisms, both terrestrial and aquatic.
51 This dynamic could lead to a reduction in their density and
52 abundance. This creates an imbalance in the food chain and poses a
53 threat to biodiversity. As a result, the ecosystem becomes more
54 vulnerable to environmental changes (Ibrahim et al. 2023, Shaffique
55 et al.2024). High concentrations of TME can cause serious problems
56 for organisms, particularly due to the increased sensitivity of
57 invertebrates to TME compared to vertebrates (Monchanin et al.

58 2021, Benhamdoun et al. 2024), such as disruption of the
59 physiological balance in earthworms, as well as in snails and beetles
60 resulting in an increase or decrease of certain oxidative stress
61 biomarkers such as AchE, GST, MDA and antioxidant enzymes.
62 This is due to the accumulation of TME in their tissues, which is
63 directly caused by the presence of these metals in their natural
64 environment, be it water, plants or soil (Du et al. 2019, Bennour and
65 Soltani 2020, Larba et al. 2023, Bouzahouane et al. 2024; Ogbeide
66 and Amayanbo 2024).

67 Currently, assessing the bioavailability of metals in soil and
68 studying the multiple effects and hazards of polluted soils are a
69 crucial issue in ecotoxicology (Chen et al. 2023; Selvanarayanan et
70 al. 2024a) in order to protect potentially exposed populations, such
71 as living fauna and flora organisms (Alsherif et al. 2022, El Mageed
72 et al. 2023, Bici et al. 2024). Recent developments in real-time
73 environmental monitoring using IoT-based systems have shown
74 promise in improving pollution management and water quality
75 assessment (Maruthai et al. 2025; Periasamy et al. 2024). as well as

76 in predicting and classifying water quality using advanced deep
77 learning models such as Attention-based Deep Differential
78 RecurFlowNet (Enkatraman et al. 2025).

79 Soil is considered to be a habitat for numerous microorganisms,
80 plants and terrestrial fauna and their interactions within which
81 diverse life communities develop and reproduce. It also provides a
82 capacity to retain pollutants to prevent their transport especially by
83 water, and subsequent transfer into food chains (Owojori et al. 2022,
84 Soliman et al. 2022, Li et al. 2023, Mwelwa et al. 2023). Different
85 species are able to indicate soil quality, among which we find
86 terrestrial gastropods, due to their direct contact with the soil and
87 their high capacity to accumulate TME, they are considered the most
88 reliable bioindicators (Guessasma et al. 2020, Salih et al. 2021,
89 Cheriti 2022, Nwagu et al. 2022, Owojori et al. 2022, Sargsyan et
90 al. 2022, Ajayi and Oyewole2023, Louzon et al. 2023). Due to its
91 sedentary lifestyle and diet, it is easily contaminated by various
92 pollutants present in the ecosystem (Al-Alam et al. 2024). The
93 assessment of metal bioavailability in soils can be performed using

94 this indicator in situ by active bioindication (by confining sentinel
95 snail species from in vitro breeding, thus making them with a known
96 biological history) or passive bioindication (based on direct
97 sampling of wild snails from their habitats, which therefore have an
98 unknown biological history) (Al-Alam et al. 2022, Sahraoui 2022).
99 In a context where environmental conservation is a global concern,
100 it is crucial to understand the mechanisms of metal contamination in
101 ecosystems and the main reasons for it (Selvanarayanan et al.
102 2024b). How does this contamination affect human health,
103 ecosystems and local biodiversity?
104 This study aims to monitor the distribution of TME in the soil of
105 three different sites in the Souk Ahras province which are Zaarouria,
106 Mechrouha and the paint factory (ENAP) under the influence of soil
107 physicochemical parameters. This work is driven by growing
108 environmental concerns over heavy metal contamination in regions
109 affected by industrial and agricultural activities, and the need for
110 effective biomonitoring tools to assess soil quality. We focused our

111 survey on measuring rates of two metals, copper (Cu) and Zinc (Zn)
112 in the foot and viscera of *Cornu aspersum* as accumulation organs.
113 In addition, a comparative analysis of sulfate concentrations was
114 carried out between sites and snails' tissues of both foot and viscera
115 using a passive biomonitoring approach which allowed us to assess
116 the level of snail contamination by TME at the investigated sites and
117 thereby evaluate soil quality.

118 **2. Materials and methods**

119 **2.1. Sampling sites**

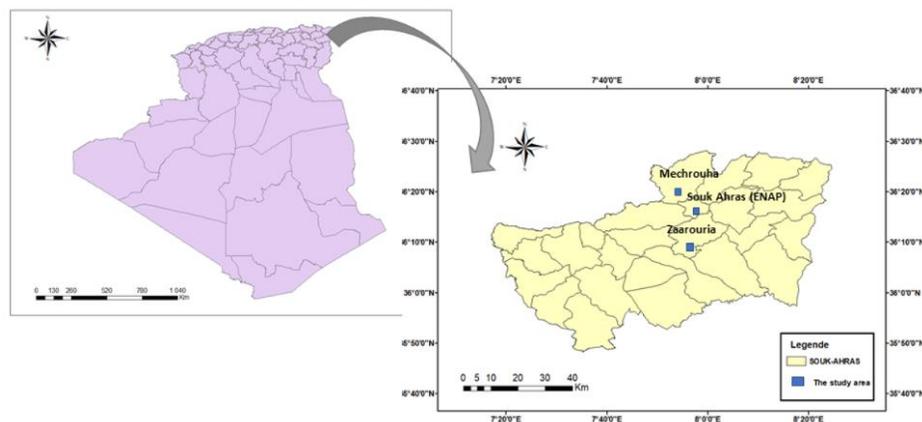
120 This study was carried out in the geographical region of the extreme
121 northeast of Algeria, which includes three different sampling sites
122 in Souk Ahras province, bounded to the northwest by the cities of
123 El Tarf and Guelma and to the southwest by Oum El Bouaghi city,
124 with Tébessa to the southeast and Tunisia to the east. Our sampling
125 area is characterized by its mountainous terrain, it is also
126 characterized by its topographical diversity, with altitudes ranging
127 from 1000 m in the north to 650 m in the south. The area is divided
128 into two geographical regions: in the north, mountainous and forests

129 are distributed, characterized by a subhumid climate and siliceous
130 or limestone soils, with an average annual rainfall of approximately
131 730 mm. On the other hand, the south is composed of high plains
132 and pastures, with a semi-arid climate and brownish agricultural
133 soils, often covered with a layer of limestone, with an average
134 annual rainfall of 350 mm per year (Boudy 1955).

135 In addition, the sampling sites were selected according to a gradient
136 of soil contamination compared to their proximity (distances) to
137 factories of different industries. An area without any industrial
138 activity was also selected, taking into account their accessibility, the
139 ease of sampling and the abundance of *C. aspersum* species. Three
140 monitoring sites were selected, as illustrated in fig.1:

- 141 ○ ENAP: National Enterprise of Paints of Souk Ahras located
142 near the national road N° 16 (36° 17' 21" N, 7° 56' 19" E),
143 the factory is responsible for manufacturing paints by using
144 several raw materials: binders, pigments, solvents and glues.

- 145 ○ Mechrouha: is located 21 km northwest of Souk Ahras city
146 and 58 km southeast of Guelma ($36^{\circ} 21' 0''$ N, $7^{\circ} 49' 60''$ E),
147 this region is characterized by its rugged terrain, it consists
148 mainly of mountains, contains stunning natural sites and is
149 known for its dense forest cover.
- 150 ○ Zaarouria: is located 10 km west of Souk Ahras city ($36^{\circ} 13'$
151 $38''$ N, $7^{\circ} 57' 28''$ E), it comprises a natural mountainous
152 area, an urban and an industrial area characterized by the
153 presence of a brick factory. It is also distinguished by its
154 variety of trees, including vineyards.



155
156
157
158

Figure 1: Map showing the sampling sites of *C. aspersum* in Souk Ahras province (northeastern Algeria)

159 **2.2. Experimental approach**

160 The use of sentinel bioindicator species has become a widely
161 recognized phenomenon for assessing soil quality in recent decades.
162 The bioindication method used in this study is passive bioindication.
163 This approach is based on the principle that terrestrial gastropods
164 due to their sensitivity to environmental changes can serve as
165 reliable indicators of soil quality, thus allowing the monitoring of
166 concentrations of various analyses including metals (Dupont et al.
167 2023, Larba et al. 2023)

168 **2.3. Biological model**

169 The biological material used is a terrestrial gastropod, the *C.*
170 *aspersum* snail (Müller 1774) randomly collected from three sites in
171 the Souk Ahras province (northeastern Algeria). The snails (with an
172 average weight of 8 ± 0.5 g) were placed in aestivation under dry
173 conditions, then they were woken up by spraying with
174 demineralized water and fed with fresh lettuce for two days before
175 being deprived of food for another two days to ensure that their
176 intestinal tract was empty.

177 **2.4. Soil physicochemical characteristics**

178 A physicochemical analysis was carried out on three soil samples
179 collected from three different sites to determine common soil
180 characteristics, in particular: hydrogen potential (pH) according to
181 Baize and Jabiole (1995), electrical conductivity (EC) according
182 to(Okalebo et al. 2002), salinity (Rhoades et al. 1982), nitrogen (N)
183 based on Kjeldahl method (1883), phosphorus (P) according to
184 Olsen et al. (1954), carbon (Walkley and Black 1934), total
185 limestone content (Drouineau 1942), organic matter (OM)
186 (Bonnefont et al. 1980), cation exchange capacity (CEC) according
187 to Thomas (1982) and soil texture(Lag et al. 2008). This analysis
188 was carried out at the Horizon private laboratory in Annaba city.

189 **2.5. TME extraction by hot acid mineralization**

190 After snails' dissection and drying of the viscera and feet of *C.*
191 *aspersum* in an oven at 65 °C for 48 to 72 hours, the dry organic
192 matter was ground to a fine powder using a ceramic mortar and
193 pestle. 100 mg of dry matter was digested in 3 ml of an acid mixture

194 consisting of: HNO₃: H₂SO₄: HClO₄ in a volume ratio of 10: 1:
195 0.5 at 110 °C for 2 h in Teflon bombs.

196 The extracts were recovered in 50 ml of 0.5% (v/v) HNO₃, then
197 filtered through Whatman N° 1 filter paper and injected into the
198 atomic absorption spectrometer (PinAAcle 900T, PerkinElmer,
199 Waltham, MA, USA) to determine the content of the two metals (Cu
200 and Zn). The Cu and Zn standards used to calibrate the instrument
201 were prepared using PerkinElmer solutions of 1000 mg L⁻¹
202 (PerkinElmer, Waltham, MA, USA). The concentration range
203 varied depending on the nature of element being analyzed and its
204 expected abundance in the mineralized samples. The calibration
205 ranges for Cu and Zn were 0-5 mg L⁻¹ and 0-1 mg L⁻¹ respectively
206 (Sleimi et al. 2022, Bankaji et al. 2023, Bouzahouane et al. 2024).

207 **2.6. Soil analysis by X-ray fluorescence spectrometry**

208 The analyses were carried out at the common service unit for
209 research “Atomic Absorption Spectrometer” and the Research
210 Laboratory RME-Resources, Materials and Ecosystems, University
211 of Carthage, Tunisia.

212 The studies presented here were carried out using an X-ray
213 fluorescence (XRF) spectrometer. This analyzer offers the highest
214 performance in XRF analysis, providing the lowest possible
215 detection limits for nearly forty elements. It is specifically designed
216 for the analysis of solid metals, alloys, ores and soils. This last
217 feature was implemented for our study. The analysis time lasts 30 s
218 per sample. The results are expressed in parts per million (ppm), and
219 they are particularly positive, allowing the direct use of the cooper
220 and zinc concentrations.

221 Soil samples (5 cm deep) were collected from the ENAP, Zaarouria
222 and Mechrouha sites. Once collected, the soil to be analyzed must
223 first be dried in an oven at 65 °C, then ground and sieved to a mesh
224 size of 2 mm. They are then placed in capsules and covered with
225 Mylar film. For analysis, the samples are placed in an automatic
226 feeder of the XRF spectrometer (Téreygeol et al. 2010, Guessasma
227 et al. 2020, Weindorf and Chakraborty 2020, Kim et al. 2023).

228

229 **2.7. Sulfates analysis**

230 For the quantification of sulfates, a photometric method was
231 employed, sulfates are precipitated in a hydrochloric acid medium,
232 forming barium sulfate. After mineralization of our samples and
233 preparation of stock solutions, a precise dosing technique was
234 followed. In test tubes, 2.5 ml of the solution (test sample) was
235 combined with 17 ml of distilled water, 0.5 ml of HCL and 2.5 ml
236 of the 20% BaCl₂ + PVP solution. These mixtures were vigorously
237 stirred at a constant speed for exactly 1 minute, followed by a resting
238 period of 15 minutes. The spectrometer cuvettes were then filled and
239 the turbidity measured 30 seconds later at a wavelength of 650 nm.
240 The sulfates transformed into suspended barium sulfate in a range
241 solution were distributed in 6 tubes: 0-15-21-27-30-90 mg L⁻¹ of
242 SO₄⁻². This detailed methodology ensures a meticulous analysis of
243 the sulfates present in the sample (Gboko et al. 2022).

244 **2.8. Statistical analysis**

245 In the present study, all our statistical analyses were performed
246 using R, version 4.2.2 (R Core Team 2022, Ihaka and Gentleman

247 1996) for MacOS (<http://cran.r-project.org>). Shapiro-Wilk test was
248 used to test variables normality. Data were expressed as mean \pm
249 standard error (se). The comparisons between sites for the soil
250 physico-chemical characteristics and between tissues for trace metal
251 elements (TME) and sulfates were assessed by the nonparametric
252 Kruskal-Wallis (KW) rank sum test. The KW test was followed by
253 the nonparametric pairwise Dunn's test (with Bonferroni adjusted p-
254 value) to find post-hoc statistical differences at $\alpha= 0.05$ as
255 significant level. Correlations between the analysed variables were
256 also calculated by using Spearman's nonparametric correlation
257 (with Bonferroni adjusted p-value). Finally, in order to characterise
258 the soil samples according to their physicochemical characteristics
259 and their possible impacts on the TME and sulfates concentrations
260 in the snail's tissues; we carried out also a multivariate analysis by
261 applying a principal component analysis (PCA) as ordination
262 technique. Several R packages (libraries) were also used in our
263 statistical analyses and to plot data results such as 'FactoMineR' (Lê
264 et al. 2008), 'ggplot2' (Wickham 2016), 'dunn.test' (Dinno 2017),

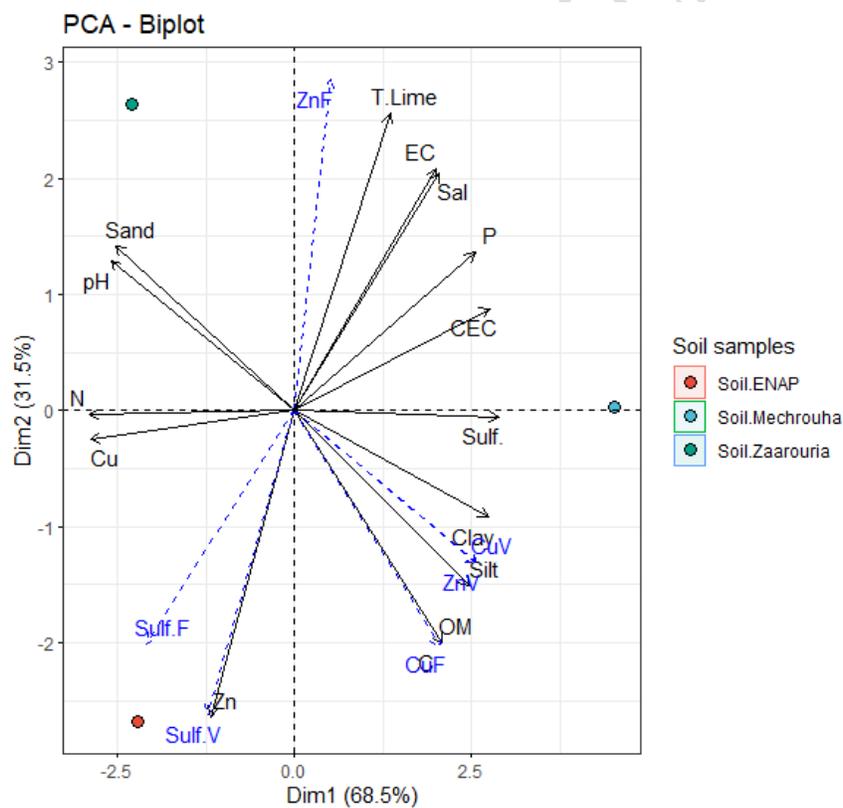
265 'factoextra' (Kassambara and Mundt 2020), 'Hmisc' (Harrell 2023),
266 'ggcorrplot' (Kassambara 2023a), 'ggpubr' (Kassambara 2023b),
267 'psych' (Revelle 2023) and 'PMCMRplus' (Pohlert 2023).

268 **3. Results**

269 **3.1. TME and sulfates analysis across distinct sites**

270 We used the Principal Component Analysis (PCA) because it is a
271 real versatile statistical method. In our study we applied it as an
272 ordination technique and it allowed us to reduce the dimensionality
273 of our dataset (based on 21 variables) and to preserve the most
274 important patterns or relationships between the analyzed variables
275 according to the target sites. The PCA provided an approximation
276 of our original data on *C. aspersum* snails' TME: copper (CuF and
277 CuV) and zinc (ZnF and ZnV) as well as sulfates (Sulf. F and Sulf.
278 V) within three distinct sites in the Souk Ahras province (Zaarouria,
279 Mechrouha and ENAP). In our case, the PCA enabled us the
280 assessment of correlations between snails' TME (blue variables on
281 the biplot, Fig. 2) and with other soil variables (TME and texture-
282 physicochemical parameters): copper (Cu), zinc (Zn), sulfates
283 (Sulf), sand, silt, clay, cation exchange capacity (CEC), organic

284 matter (OM), total lime (T, Lime), carbon (C), phosphorus (P),
 285 nitrogen (N), salinity (Sal), electrical conductivity (EC) and pH. In
 286 a synthetic way, the PCA results indicate that the first two axes
 287 explain 100% of the total inertia (data variance) present in our
 288 dataset, indicating that the biplot represents the entirety of the
 289 variability (Fig. 2).



290

291 **Figure 2: Principal component analysis (PCA) biplot of the three sampled site**
292 **characterization (Dim 1: 68.5% and Dim 2 = 31.5%). The biplot shows the PCA**
293 **scores of the fifteen explanatory variables as black arrows. Points on the same**
294 **side as a given variable should be interpreted as having a high contribution on**
295 **it. In this study, the six TME measured in snails' tissues of *C. aspersum* were**
296 **treated as supplementary variables (the six blue arrows).**

297 On the one hand, the 1st axis (Dim 1) alone, explained 68.5% of the
298 total variation and showed strongly positive correlation especially
299 with: Sulf ($r = 1$; $\cos^2 = 1$), CEC ($r = 0.95$; $\cos^2 = 0.91$), clay ($r =$
300 0.95 ; $\cos^2 = 0.90$), P ($r = 0.88$; $\cos^2 = 0.78$), silt ($r = 0.85$; $\cos^2 =$
301 0.73), OM ($r = 0.72$; $\cos^2 = 0.52$), C ($r = 0.72$; $\cos^2 = 0.52$) and Sal
302 ($r = 0.71$; $\cos^2 = 0.71$). In addition, the 1st axis is also negatively
303 correlated with the following variables: N ($r = -1$; $\cos^2 = 1$), Cu ($r =$
304 -1 ; $\cos^2 = 0.99$), pH ($r = -0.90$; $\cos^2 = 0.80$) and sand ($r = -0.87$; \cos^2
305 $= 0.76$). On the other hand, the 2nd axis (Dim 2) just explained 31.5%
306 of the total variation is and it was also positively correlated with the
307 variables: T, Lime ($r = 0.88$; $\cos^2 = 0.78$), EC ($r = 0.72$; $\cos^2 = 0.52$)
308 and Sal ($r = 0.71$; $\cos^2 = 0.50$) and as well as negative correlations
309 with the variable Zn ($r = -0.91$; $\cos^2 = 0.83$).

310 Moreover, PCA showed different patterns of correlations between
311 analyzed variables and the results revealed an important
312 differentiation between sites due to fluctuations in the
313 concentrations of various TME measured values from the soil as
314 well as in the foot and viscera of *C. aspersum*.

315 The 1st axis generally distinguished Zaarouria and ENAP sites from
316 the Mechrouha one which is characterized by silty-clay soil and high
317 concentrations values, mostly of Sulf, CEC, OM, C, P, Sal, CuV,
318 ZnV and CuF. By contrast, it exhibited low values for Cu, Sand, N
319 and pH concentrations of Mechrouha in comparison with the two
320 other sites. Furthermore, the 2nd axis mainly differentiated the
321 Zaarouria site from the ENAP one. The Zaarouria site which is
322 characterized by sandy texture and high values of pH, ZnF and T.
323 Lime. By opposition, the ENAP site is marked by very high
324 concentrations of Zn, Sulf. F and Sulf. V.

325 The PCA results also highlighted the relationship between the soil's
326 physicochemical parameters and the distribution of sulfates as well
327 as TME in the soil and the edible tissues of *C. aspersum*. The

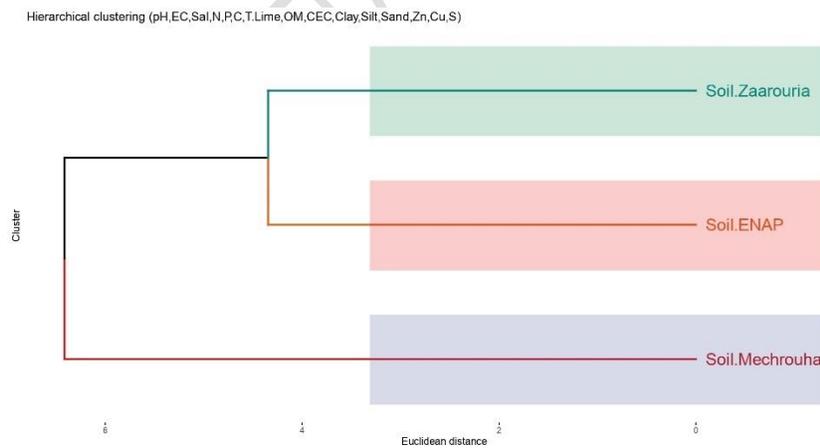
328 findings indicated that the soil Zn is completely independent of
329 T.Lime, Sal and EC. Conversely, these latter physicochemical
330 parameters could promote the accumulation of ZnF. Additionally,
331 ZnV appeared negatively correlated with the soil pH.

332 Likewise, soil sulfates appeared positively correlated with OM,
333 CEC, P, C and the silty-clay textures. In contrast, T. Lime, EC and
334 Sal could inhibit the absorption of sulfates in the foot and viscera
335 (Sulf. F and Sulf. V) in *C. aspersum species*. Copper exhibited a
336 tendency to be present in higher concentrations in soils rich in N and
337 sandy texture compared to silty-clay ones. Finally, the PCA biplot
338 showed that copper absorption by snails' tissues (CuF and CuV)
339 could be more pronounced in silty-clay soils rich in OM and C than
340 in sandy soils.

341 **3.2. Hierarchical Cluster Analysis (HCA)**

342 The objective of HCA is to evaluate the similarities between soil
343 samples according to the bioavailability of various TME in the soil
344 as well as the physicochemical parameters analyzed across the three
345 geographic sites, the cluster analysis dendrogram presented a

346 plausible classification (in concordance with the PCA results). It
 347 clearly separated the Mechrouha site (cluster 1, high sulfates level
 348 according the PCA biplot), characterized by silty-clay soil, from the
 349 two others. The cluster 2 included two relatively homogeneous sites:
 350 Zaarouria and ENAP, mainly characterized by high N and Cu levels
 351 (according to the PCA biplot). Thus, the hierarchical analysis
 352 revealed heterogeneity levels among the soil samples, reflecting the
 353 north-south geographic transect. The observed heterogeneity could
 354 support the TME and sulfates distributions both in soil and in snails
 355 (Fig. 3, see PCA biplot Fig. 2).



356
 357
 358
 359

Figure 3: Hierarchical classification of sampling sites of *C. aspersum* (Zaarouria, Mechrouha and ENAP) based on soil physicochemical parameters and TME.

360 **3.3. TME and sulfates assessment in the snail's foot and**
361 **viscera**

362 The mean of the CuF concentrations (in dry weight) ranged from
363 $46.91 \pm 2.83 \mu\text{g/g}$ to $78.58 \pm 5.06 \mu\text{g/g}$ at Zaarouria and Mechrouha
364 sites; respectively (Fig. 4); with a minimum of $36.44 \mu\text{g/g}$ at
365 Zaarouria site and a maximum of $140.33 \mu\text{g/g}$ at ENAP site. The
366 mean of the CuV concentrations ranged from $49.52 \pm 2.06 \mu\text{g/g}$ to
367 $99.55 \pm 8.04 \mu\text{g/g}$ at Zaarouria and Mechrouha sites, respectively;
368 with a minimum of $37.45 \mu\text{g/g}$ at Zaarouria site and a maximum of
369 $146.66 \mu\text{g/g}$ at Mechrouha site. The nonparametric Kruskal-Wallis
370 rank sum test revealed significant differences between sites only
371 in viscera tissues (CuV, $p < 0.05$).

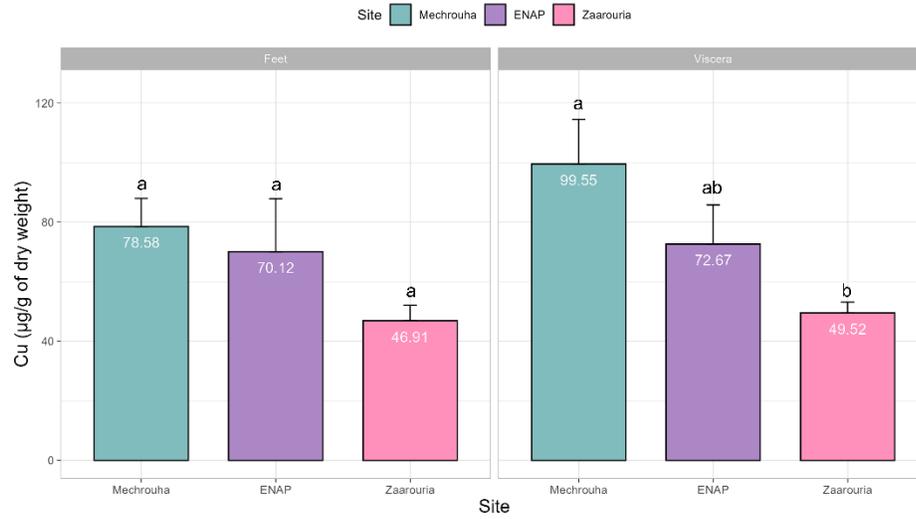
372 Besides the Cu assessment, the mean of the ZnF concentrations
373 ranged from $68.85 \pm 1.21 \mu\text{g/g}$ to $73.51 \pm 1.81 \mu\text{g/g}$ at ENAP and
374 Zaarouria sites, respectively (Fig. 5); with a minimum of $61.60 \mu\text{g/g}$
375 at Mechrouha site and a maximum of $84.10 \mu\text{g/g}$ at Zaarouria site.

376 In addition, the mean ZnV concentrations ranged from 161.61 ± 8.38
377 $\mu\text{g/g}$ and $327.11 \pm 32 \mu\text{g/g}$ at Zaarouria and Mechrouha sites,
378 respectively; with a minimum of $123.68 \mu\text{g/g}$ at Zaarouria site and

379 a maximum of 472.41 $\mu\text{g/g}$ at Mechrouha site. The Kruskal-Wallis
380 rank sum test also revealed significative differences among sites
381 only in viscera tissues (ZnV, $p < 0.05$).

382 In this study we have also performed the nonparametric Mann-
383 Whitney rank sum test to compare, within each site, TME and
384 sulfates concentrations between viscera and feet tissues. The
385 statistical results highlighted significant differences in sulfates
386 concentrations between snails' viscera and feet tissues only for
387 ENAP site (Fig. 6). Regarding to the TME data, the Mann-Whitney
388 test also revealed, in each site, no significant differences in Cu
389 concentrations between viscera and feet tissues. In contrast,
390 significant differences in Zn concentrations were highlighted among
391 viscera and feet tissues within each site. All these results will be
392 explained in the discussion section.

TME: Cu assessment



393

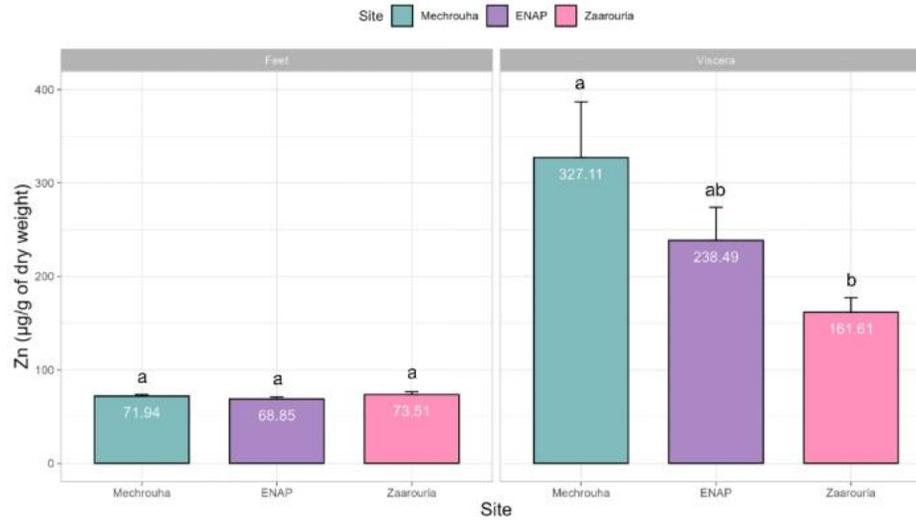
394

395

396

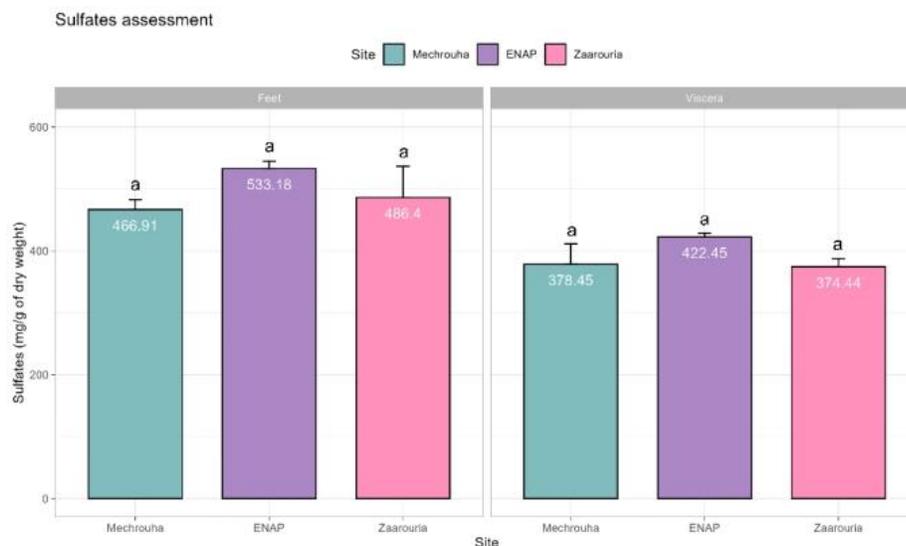
Figure 4: Variations in copper (Cu) concentrations measured in foot and viscera tissues of *C. aspersum*. Data are expressed as mean ± standard error (SE); the lowercase letters indicate significant differences (Dunn's test) among sites (n= 5).

TME: Zn assessment



397

398 **Figure 5: Variations in zinc (Zn) concentrations measured in foot and viscera**
399 **tissues of *C. aspersum*. Data are expressed as mean \pm standard error (SE); the**
400 **lowercase letters indicate significant differences (Dunn's test) among sites (n= 5).**



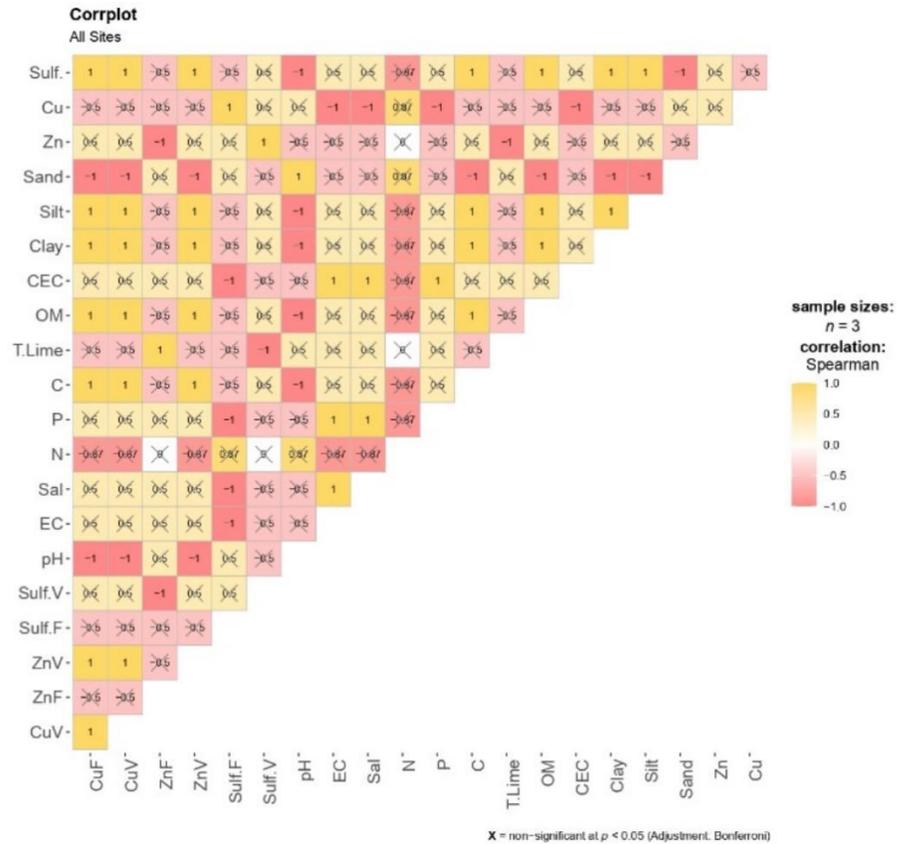
401
402 **Figure 6: Variations in Sulfates (Sulf) concentrations measured in foot and viscera**
403 **tissues of *C. aspersum*. Data are expressed as mean \pm standard error (SE); the**
404 **lowercase letters indicate significant differences (Dunn's test) among sites (n= 5).**

3.4. Correlation analysis

406 The results of the Spearman's correlation analysis showed that 68%
407 of the correlations, calculated between the twenty-one studied
408 variables, were not statistically significant. All correlation
409 coefficients are shown in fig.7 as a corrplot; the latter provided a
410 visual exploratory tool on the correlation matrix relating to TME,

411 sulfates and physicochemical parameters concentrations (measured
412 in snails' tissues and soil samples) and it determined the statistical
413 significance of the correlations.

414 The corrplot highlighted that only 67 correlation coefficients were
415 statistically significant ($p < 0.05$). As an example: the results clearly
416 indicated highly significant positive correlations ($r = 1$) between
417 several variables, such as Sulf. -CuV, ZnV-CuV, Zn-Sulf.V, Sal-
418 EC, P-N and CuF-ZnV. On the other hand, the corrplot also
419 highlighted highly significant negative correlations ($r = -1$) between
420 many variables such as pH-CuF, Sulf.V-ZnF, EC-Sulf.F, C-pH, Zn-
421 ZnF and Sulf. -Sand.



423 **Figure 7: Spearman's correlation carried out on TME, sulfates and**
 424 **physicochemical parameters measured in both snails' tissues and soil samples**
 425 **in all sites.**

426 **4. Discussion**

427 **4.1. Impacts of soil physicochemical parameters on the**
 428 **bioavailability of TME and sulfates**

429 Physicochemical parameters are among the main factors influencing
 430 the distribution and mobility of metals in soils. Several studies

431 including those by van Gestel (2008), Laurent et al. (2020) and
432 Kicińska et al. (2022) have demonstrated that an increase in pH
433 reduces the concentrations of certain metals such as zinc, cadmium
434 and lead. Włostowski et al. (2016) highlighted that under acidic soil
435 conditions the concentration of cadmium tends to increase, which
436 promotes its absorption by *Helix pomatia* and its accumulation in
437 the intestinal gland. This accumulation results from various
438 mechanisms, including ingestion of plants that have absorbed
439 cadmium, direct ingestion of soil particles, and cutaneous diffusion
440 through the snail's foot. According to Wei et al. (2021) acidic soils
441 contribute significantly to the increased availability of Cu, Ni and
442 Zn while simultaneously reducing the accumulation of Cd, Cu and
443 Zn. These observations emphasize the influence of soil pH on the
444 mobility and bioavailability of heavy metals as well as their uptake
445 by terrestrial organisms. Our results are consistent with these
446 findings showing that in alkaline soils with elevated pH levels the
447 concentrations of Zn^V, Cu^V and Cu^F tend to decrease. Heavy
448 metals are generally insoluble and less available for plant uptake in

449 alkaline soils, which could be explained by the fact that at high pH
450 levels metals may form insoluble complexes or precipitate with
451 other soil elements thereby reducing their mobility and
452 bioavailability in soil (Yu et al. 2016; Sintorini et al. 2021). Shahid
453 et al. (2017) demonstrated that soil pH plays a critical role in
454 modulating the chemical forms of heavy metals by affecting various
455 physicochemical processes, including adsorption /desorption,
456 precipitation/dissolution and the formation of complexes with soil
457 components. Conversely, the study by Li et al. (2014a) specifically
458 mentions that soil acidity not only promotes the bioavailability of
459 metals to terrestrial species such as *Eisenia fetida* but also increases
460 their accumulation in soils.

461 The PCA analysis from this study revealed that the soil at
462 Mechrouha is richer in sulfates compared to other sites. It is well
463 known that this site experienced several significant fires two years
464 ago, which likely led to substantial changes in soil composition
465 including pH, organic matter content, and nutrient availability.
466 Among the constituents of ash is sulfuric acid, which can increase

467 soil acidity as evidenced by our findings for this site. Clemente et
468 al. (2003) emphasized that soil acidification resulting from the
469 oxidation of sulfides is a key factor in increasing the bioavailability
470 of metals as demonstrated by the concentrations of Cu and Zn
471 observed in the tissues of *C. aspersum* (CuF, CuV and ZnV).
472 Furthermore, soil texture as discussed by Angelaki et al. (2022);
473 Sungur et al. (2023) influences the distribution and availability of
474 metals. Clayey soils exhibit a high adsorption capacity for various
475 metals except for lead, which is in agreement with our results. We
476 observed that CuF, CuV and ZnV tend to accumulate in silty clay
477 soils rather than in sandy soils. The presence of Cu and Zn in the
478 tissues of *C. aspersum* can be attributed to the organic richness and
479 acidity of silty clay soils. These conditions promote increased metal
480 desorption enhancing their solubility and availability not only to
481 plants but also to other soil organisms such as snails (He et al.
482 2020;(AbdelRahman et al., 2022); Zhu et al. 2023). Our results
483 suggest that an increase in certain physicochemical parameters such
484 as T, Lime, EC and salinity reduces the presence of zinc and copper

485 in the soil. This observation contradicts the findings of Benahmed
486 et al. (2016) and Bartkowiak et al. (2020) who reported an increase
487 in zinc concentration in response to salinity, EC and total lime. A
488 possible explanation for our findings could be competition between
489 sodium cations (Na^+) in saline soils and other cations such as
490 calcium (Ca^{2+}) in calcareous soils with zinc (Zn^{2+}) and copper (Cu^{2+})
491 ions for sorption sites on soil particles. This competition could
492 reduce the amount of zinc retained by the soil (Acosta et al. 2011)
493 increasing its concentration in the soil solution, making it more
494 accessible for uptake by plants and other terrestrial organisms. This
495 mechanism could also explain the results for ZnF, whose
496 accumulation is promoted by these same three parameters (T, Lime,
497 Sal and EC). This observation aligns with studies by Zhang et al.
498 (2020), Park et al. (2021) and which confirmed that salinity affects
499 the mobility of metals particularly cadmium by increasing its
500 mobility. Kadkhodaie A. (2012) also observed a significant increase
501 in cadmium in different parts of sunflower and Sudan grass (roots,
502 stems and leaves) in response to increased salinity.

503 In addition, our results also reveal that the parameters T, Lime, Sal
504 and EC could influence the mobility and availability of sulfates
505 inhibiting their uptake by the edible tissues of *C. aspersum* (Sulf.F
506 and Sulf.V). This phenomenon could be explained by the fact that
507 in calcium and sodium rich soils sulfates can form partially insoluble
508 ionic complexes such as gypsum. These less soluble complexes tend
509 to remain bound to the soil matrix. This hypothesis is supported by
510 the study of Zunino and Scrivener (2020) which suggests that
511 sulfates tend to form fewer mobile complexes in the presence of
512 calcium. The study by Salao et al. (2017) confirmed that industrial
513 emissions play a significant role in the concentration of nitrogen in
514 the environment, which corroborates our results highlighting the
515 high levels of nitrogen in the Zaarouria site (urban and industrial).
516 This pollution could be due to the release of nitrogen compounds
517 such as ammonia and nitrogen oxides during industrial processes.
518 These emissions can be deposited in the soil by precipitation, thus
519 increasing the concentration of nitrogen in the soil (Stevenson
520 1965). The application of nitrogen fertilizers can alter the mobility

521 and bioavailability of metals in the soil as confirmed by Li et al.
522 (2019) whose study suggests that increasing nitrogen inputs to the
523 soil could reduce the bioavailability of certain metals thus limiting
524 their uptake by plants and microorganisms. These findings are
525 consistent with our results which show that the presence of nitrogen
526 promotes Cu accumulation in the soil while reducing its transfer to
527 the tissues of *C. aspersum*.

528 **4.2. TME (copper and zinc) in the foot and viscera of snails**

529 The presence of TME in the soil can have a significant impact on
530 organisms such as terrestrial gastropods which are considered
531 bioindicators of pollution. Due to their sedentary lifestyle, these
532 organisms have the ability to accumulate various pollutants
533 including TME, thereby reflecting the contamination status of their
534 environment (Mukhtorova et al. 2023; Rašković et al. 2023). Among
535 the metals detected in the soils of the studied areas are zinc and
536 copper which were also found in the edible tissues of the snails. Our
537 analyses of the TME indicate that Cu and Zn were primarily

538 accumulated in the foot and viscera of snails, revealing significant
539 spatial variation among the three sampling sites for Cu in these parts,
540 as well as for Zn in the viscera. This accumulation was particularly
541 notable at the Mechrouha and ENAP sites compared to the Zaarouria
542 site. The spatial variation of Cu can be explained by the different
543 concentrations of this metal in the soils of three sites which may be
544 influenced by the physicochemical parameters of the soil, leading to
545 differences in absorption by the snail's foot and viscera. The spatial
546 variation of Zn in the viscera only can be attributed to varying levels
547 of Zn in the food sources of *C. aspersum* as well as the specific
548 capacities of the viscera to absorb and accumulate this metal (Mitev
549 et al. 2023) the absence of spatial variation of Zn in the snails' feet
550 can be explained by similar or very close concentrations of Zn in the
551 soils of the three sites. Our results also revealed a notable fluctuation
552 in the distribution of Cu and Zn between the tissues of *C. aspersum*.
553 The viscera exhibit a higher accumulation of metals compared to the
554 foot which supports the findings of Gomot-de Vaufleury and Pihan
555 (2002), Hoang et al. (2008) and Benhamdoun et al. (2024) who

556 confirmed through their studies on metal bioaccumulation in land
557 snails that the viscera accumulate significantly more metals than the
558 foot. For example, Hoang et al. (2008) reported that copper was
559 predominantly accumulated in the viscera, accounting for
560 approximately 60% of the total burden, while the shell contained
561 less than 4%. The foot represented about 40% of the total copper
562 accumulation. This observation can be explained by the particularly
563 active metabolism of the viscera which is responsible for the
564 accumulation, processing and storage of nutrients, leading to higher
565 metal accumulation. In contrast, the foot is primarily composed of
566 muscles dedicated to locomotion. However, this does not exclude
567 the accumulation of metals in the foot or other parts of the snail, as
568 some metals tend to concentrate in the foot or the shell (Aleksander-
569 Kwarczak and Gołas-Siarzewska 2015; Ćirić et al. 2018; Huang et
570 al. 2018). In addition, the mode of exposure to these contaminants
571 (whether by dermal contact or ingestion) plays a significant role in
572 the distribution of metals within various parts of the snail body. This
573 observation aligns with the study by (De-Vaufleury & Bispo, 2000)

574 who supported this theory through their experiments. Sebban et al.
575 (2022) revealed that certain metals exhibit a preference for
576 accumulation in specific tissues, with Zn tending to accumulate
577 more heavily in the viscera, while Cu is found in abundance in the
578 foot of snails. Our results regarding Zn are consistent with this study
579 but they are different for Cu. Indeed, we observed that Cu
580 accumulates almost equally in both the viscera and the foot of the
581 snails with a slight predominance in the viscera in all three study
582 areas. This can be explained by a significant presence of Cu in the
583 snails' food sources rather than in the soils in which they live.

584 **4.3. Sulfates concentration in the feet and viscera of snails**

585 Although sulfur is an essential macronutrient (Maruyama-Nakashita
586 and Ohkama-Ohtsu 2017; Li et al. 2020) high concentrations can
587 cause issues in soil, water and microorganisms (Geurts et al. 2009;
588 Björnerås et al. 2019). There are several potential sources of sulfates
589 in the soil as we hypothesized in our study. We detected sulfates at
590 all three distinct sites, the presence of sulfates at the Zaarouria site

591 could be attributed to vineyards where sulfur-based treatments are
592 commonly used (Hermes et al. 2022). Additionally, this presence
593 could also be explained by household waste as this site is located in
594 an urban area. The study by Malik et al. (2020) showed that organic
595 matter promotes the presence of sulfur in the soil. Therefore, it is
596 reasonable to conclude that household waste which often contains
597 organic matter and nutrients could also be a source of sulfur. For the
598 Mechrouha site, we hypothesize that the presence of sulfates is a
599 result of the wildfires that occurred during the summer of 2022 both
600 at this site and in other areas of Souk Ahras province. This is
601 consistent with the study by Hermes et al. (2021) which shows that
602 wildfire can introduce sulfur into the soil especially in agricultural
603 areas by altering soil chemistry and facilitating the release of sulfur
604 into the environment. Regarding the ENAP site (industrial
605 activities), the presence of sulfates could be attributed to sulfur in
606 raw materials used in paints particularly pigments based on barium
607 sulfates. This site is the only one showing a significant difference in
608 sulfate accumulation between the foot and the viscera of the snails

609 (p-value = 0.0079), with accumulation being much higher in the foot
610 (533.18 mg/g in dry weight). This contradicts the study by Lobo-da-
611 Cunha (2019) which indicated that the digestive gland of mollusks
612 is the primary site for nutrient absorption. Our result could be
613 explained by a much higher concentration of sulfates in the soil
614 compared to what is found in the snails' food sources. Additionally,
615 this difference may be due to a high concentration of calcium (Ca^{2+})
616 and magnesium (Mg^{2+}) ions which compete with sulfates for
617 absorption, resulting in different accumulation levels in the foot and
618 viscera of the snails. In contrast, the accumulation of sulfates in both
619 the foot and viscera of snails across the three study sites (inter-site)
620 showed no significant variation. This could be due to consistent
621 sulfate concentrations across the three study sites and a uniform
622 absorption mechanism by the snails' feet and viscera.

623 **5. Conclusion**

624 The results of this research highlight the potential of utilizing metals
625 concentrations as environmental indicators, while emphasizing the
626 need to consider spatial variations as well as the physicochemical

627 parameters of the soil that influence these indicators at our sites,
628 which in turn affects the level of metal absorption by terrestrial
629 gastropods. We also measured sulfate levels in the foot and viscera
630 of *C. aspersum* based on preliminary soil analysis by X-ray
631 fluorescence spectrometry which revealed the presence of sulfur
632 both in the soil and our target snail species. The results demonstrate
633 that physicochemical parameters significantly influence the
634 availability of TME in the soil leading to differences in their
635 accumulation within edible tissues. Specifically, it was observed
636 that copper and zinc accumulate much more in the viscera than in
637 the foot of *C. aspersum* while sulfate shows the opposite pattern
638 with higher concentrations in the foot. These results confirm that *C.*
639 *aspersum* is an excellent bioindicator of soil pollution.
640 Accumulation in the different tissues of edible snails including our
641 study species *C. aspersum* could have harmful effects on human
642 health through the food chain. This study presents certain
643 limitations, including the lack of seasonal monitoring, limited
644 geographic scope, and a restricted range of analyzed contaminants.

645 Future research should broaden the study area, incorporate a wider
646 array of pollutants and bioindicator species, and include temporal
647 monitoring to better understand bioaccumulation dynamics.
648 However, such proposed work may also face constraints such as
649 logistical challenges, environmental variability, and limited
650 accessibility to target species.

651 **Acknowledgements**

652 The authors would like to express their sincere gratitude to the
653 Research Laboratory RME – Resources, Materials and Ecosystems,
654 University of Carthage, Faculty of Sciences, Bizerte, Tunisia, for
655 their essential support in providing access to the necessary
656 equipment. Special thanks are also extended to all the laboratory
657 staff for their invaluable assistance throughout this work.

658 **Bibliographic references**

659 AbdelRahman, M. A. E., Metwaly, M. M., Afifi, A. A., D'Antonio, P. and Scopa,
660 A. (2022). Assessment of soil fertility status under soil degradation rate using
661 geomatics in West Nile Delta, Land, 11, 1256,
662 <https://doi.org/10.3390/land11081256>.

663 Acosta, J. A., Jansen, B., Kalbitz, K., Faz, A. and Martínez-Martínez, S. (2011).
664 Salinity increases mobility of heavy metals in soils, *Chemosphere*, 85, 1318–
665 1324, <https://doi.org/10.1016/j.chemosphere.2011.07.046>.

666 Ajayi, A. A. and Oyewole, B. O. (2023). Giant African land snails (*Achatina*
667 *achatina* and *Archachatina marginata*) as bioindicators of heavy metal
668 pollution, *Afr. J. Environ. Sci. Technol.*, 17, 80–88,
669 <https://doi.org/10.5897/AJEST2022.3131>.

670 Al-Alam, J., Millet, M., Khoury, D., Rodrigues, A., Akoury, E., Tokajian, S. and
671 Wazne, M. (2024). Biomonitoring of PAHs and PCBs in industrial, suburban,
672 and rural areas using snails as sentinel organisms, *Environ. Sci. Pollut. Res.*,
673 31, 4970–4984, <https://doi.org/10.1007/s11356-023-31493-6>.

674 Al-Alam, J., Millet, M., Khoury, D., Rodrigues, A., Harb, M., Akoury, E.,
675 Tokajian, S. and Wazne, M. (2022). Snails as temporal biomonitors of the
676 occurrence and distribution of pesticides in an apple orchard, *Atmosphere*, 13,
677 1185, <https://doi.org/10.3390/atmos13081185>.

678 Aleksander-Kwaterczak, U. and Gołas-Siarzewska, M. (2015). Comparative
679 analysis of *Helix pomatia* L. shells found in soils with varying degrees of
680 contamination (southern Poland), *Geol. Geophys. Environ.*, 41, 299,
681 <https://doi.org/10.7494/geol.2015.41.4.299>.

682 Alsherif, E. A., Al-Shaikh, T. M. and Abdelgawad, H. (2022). Heavy metal effects
683 on biodiversity and stress responses of plants inhabiting contaminated soil in

684 Khulais, Saudi Arabia, *Biology*, 11, 164,
685 <https://doi.org/10.3390/biology11020164>.

686 Amlan, P. G., Chouti, W. K., Dedjiho, C. A., Fangnon, K. R. and Chitou, N. E.
687 (2023). Évaluation de la pollution chimique des eaux par les éléments traces
688 métalliques (ETM) : cas de la rivière Mekrou (nord-ouest Bénin), *J. Appl.*
689 *Biosci.*, 192, 20331–20346, <https://doi.org/10.35759/JABs.192.4>.

690 Angelaki, A., Dionysidis, A., Sihag, P. and Golia, E. E. (2022). Assessment of
691 contamination management caused by copper and zinc cations leaching and
692 their impact on the hydraulic properties of a sandy and a loamy clay soil, *Land*,
693 11, 290, <https://doi.org/10.3390/land11020290>.

694 Baize, D. and Jabiol, B. (1995). Guide pour la description des sols, INRA, Paris,
695 Techniques et Pratiques.

696 Bankaji, I., Kouki, R., Dridi, N., Ferreira, R., Hidouri, S., Duarte, B., Sleimi, N.
697 and Caçador, I. (2023). Comparison of digestion methods using atomic
698 absorption spectrometry for the determination of metal levels in plants,
699 *Separations*, 10, 40, <https://doi.org/10.3390/separations10010040>.

700 Baroudi, F. (2020). Étude comparative de trois biomoniteurs (conifère, escargot
701 et miel) pour évaluer la variabilité spatio-temporelle de polluants organiques
702 dans l’atmosphère au Liban, PhD thesis, Université de Strasbourg, Chimie
703 analytique, NNT : 2020STRAF023.

704 Bartkowiak, A., Dąbkowska-Naskręt, H., Jaworska, H. and Rydlewska, M.
705 (2020). Effect of salinity on the mobility of trace metals in soils near a soda

706 chemical factory, J. Elem., 25, 501–512,
707 <https://doi.org/10.5601/jelem.2019.24.2.1875>.

708 Benhamdoun, A., Achtak, H. and Dahbi, A. (2024). Bioaccumulation of trace
709 metals in edible terrestrial snails, *Theba pisana* and *Otala* spp., in a dumpsite
710 area in Morocco and assessment of human health risks for consumers,
711 *Environ. Sci. Pollut. Res.*, 31, 42810–42826, [https://doi.org/10.1007/s11356-](https://doi.org/10.1007/s11356-024-33945-z)
712 [024-33945-z](https://doi.org/10.1007/s11356-024-33945-z).

713 Bennour, A. and Soltani, N. (2020). Assessment of soil quality in Annaba area
714 (northeast Algeria) using the earthworm *Lumbricus terrestris*: bioindicative
715 stress responses and heavy metal contamination, *Fresenius Environ. Bull.*, 29,
716 9635–9643.

717 Bici, M., Sahiti, H., Zogaj, M., Halili, J. and Bislimi, K. (2024). Data on heavy
718 metals (Pb, Ni, Zn) in soil and biota (common nettle and Roman snail) around
719 the power plant TC Kosova A in Obiliq (Kosovo), *Jordan J. Biol. Sci.*, 17, 1–
720 7, <https://doi.org/10.54319/jjbs/170101>.

721 Björnerås, C., Škerlep, M., Floudas, D., Persson, P. and Kritzberg, E. S. (2019).
722 High sulfate concentration enhances iron mobilization from organic soil to
723 water, *Biogeochemistry*, 144, 245–259, [https://doi.org/10.1007/s10533-019-](https://doi.org/10.1007/s10533-019-00581-6)
724 [00581-6](https://doi.org/10.1007/s10533-019-00581-6).

725 Bonnefont, J. C., Bonneau, M. and Souchier, B. (1980). Constituants et propriétés
726 du sol, *Revue Géogr. Est*, 20, 263–264, [https://www.persee.fr/doc/rgest_0035-](https://www.persee.fr/doc/rgest_0035-3213_1980_num_20_3_1477_t1_0263_0000_2)
727 [3213_1980_num_20_3_1477_t1_0263_0000_2](https://www.persee.fr/doc/rgest_0035-3213_1980_num_20_3_1477_t1_0263_0000_2).

728 Boudy, P. (1955). *Économie forestière nord-africaine*, vol. 4, Description
729 forestière de l'Algérie et de la Tunisie, Larose, Paris.

730 Bouzahouane, H., Kouki, R., Amri, S., Barour, C., Sleimi, N. and Ouali, K.
731 (2024). Investigating seasonal metal impact on *Stramonita haemastoma*
732 gastropod along the Algerian East Coast: Understanding through various
733 pollution indicators, *Mar. Pollut. Bull.*, 199, 116006,
734 <https://doi.org/10.1016/j.marpolbul.2023.116006>.

735 Chen, F., Wang, Q., Meng, F., Chen, M. and Wang, B. (2023). Effects of long-
736 term zinc smelting activities on the distribution and health risk of heavy metals
737 in agricultural soils of Guizhou province, China, *Environ. Geochem. Health*,
738 45, 5639–5654, <https://doi.org/10.1007/s10653-020-00716-x>.

739 Cheriti, O. (2022). Utilisation des gastéropodes pulmonés terrestres comme
740 traceurs de la pollution métallique des sols dans le bassin du Kebir Rhumel:
741 de l'inventaire à la bioindication, PhD thesis, Université Frères Mentouri
742 Constantine 1, Département de Biologie et Écologie Végétale.

743 Ćirić, J., Cerić, O., Marković, R., Janjić, J., Spirić, D., Popović, M., Pećanac, B.,
744 Baltić, B. and Baltić, M. (2018). Seasonal distributions of heavy metal
745 concentrations in different snail (*Helix pomatia*) tissues from an urban
746 environment in Serbia, *Environ. Sci. Pollut. Res.*, 25, 33415–33422,
747 <https://doi.org/10.1007/s11356-018-3295-1>.

748 Clemente, R., Walker, D. J., Roig, A. and Pilar, M. P. (2003). Heavy metal
749 bioavailability in a soil affected by mineral sulphides contamination following
750 the mine spillage at Aznalcóllar (Spain), *Biodegradation*, 14, 199–205.

751 De-Vaufleury, G. A. and Bispo, A. (2000). Methods for toxicity assessment of
752 contaminated soil by oral or dermal uptake in land snails. 1. Sublethal effects
753 on growth, *Environ. Sci. Technol.*, 34, 1865–1870,
754 <https://doi.org/10.1021/es9907212>.

755 Dinno, A. (2017). Dunn’s Test of Multiple Comparisons Using Rank Sums, R
756 package version 1.5.1.

757 Dossou, D., Bokossa, H. K. J., Adanloknonon, E. A. S., Zounon, Y., Adanloknonon,
758 S. M. I., Johnson, R. C., Fiogbe, E. D. and Edorth, P. (2022). Evaluation of
759 metal contamination of sediments and crabs (*Callinectes amnicola*) from Lake
760 Ahémé in southern Benin, *Int. J. Biol. Chem. Sci.*, 16, 2424–2435.

761 Drouineau, G. (1942). Dosage rapide du calcaire actif des sols. Nouvelles données
762 sur la répartition et la nature des fractions calcaires, *Ann. Agron.*, 2, 441–450.

763 Du, C., Wu, J., Bashir, M. H., Shaukat, M. and Ali, S. (2019). Heavy metals
764 transported through a multi-trophic food chain influence the energy
765 metabolism and immune responses of *Cryptolaemus montrouzieri*,
766 *Ecotoxicol.*, 28, 422–428.

767 Dupont, J. (2023). Le souffle polymétallique de la fonderie Horne : caractérisation
768 des dépôts atmosphériques à l’aide d’approches passives et de biosurveillance,
769 PhD thesis, Université du Québec à Montréal, Montréal.

770 El Mageed, Y. S. M. A., Ghobashy, A. E. F. A., Al-Thomali, A. W., Soliman, M.
771 F., Mohammadein, A. and El-Shenawy, N. S. (2023). Impact of heavy metals
772 contamination on biology, biochemical, and histology of *Eobania vermiculata*
773 and *Monacha obstructa*, *Toxicol. Environ. Health Sci.*, 15, 19–30,
774 <https://doi.org/10.1007/s13530-022-00233-8>.

775 Enkatraman, M., Surendran, R., Senduru Srinivasulu., Vijayakumar K. (2025)
776 “Water quality prediction and classification using Attention based Deep
777 Differential RecurFlowNet with Logistic Giant Armadillo Optimization”, *Global*
778 *NEST Journal*. <https://doi.org/10.30955/gnj.0679>.

779 Gboko, Y. D. A., Natchia, A., Keumean, K. N. and Soro, N. (2022). Suivi de la
780 qualité physico-chimique des eaux du Sassandra à la station hydrométrique de
781 Gaoulou, Sud-ouest de la Côte d’Ivoire, *Afr. Sci.*, 21, 127–141.

782 Geurts, J. J. M., Sarneel, J. M., Willers, B. J. C., Roelofs, J. G. M., Verhoeven, J.
783 T. A. and Lamers, L. P. M. (2009). Interacting effects of sulphate pollution,
784 sulphide toxicity, and eutrophication on vegetation development in fens: A
785 mesocosm experiment, *Environ. Pollut.*, 157, 2072–2081,
786 <https://doi.org/10.1016/j.envpol.2009.02.024>.

787 Gomot-de Vaufleury, A. and Pihan, F. (2002). Methods for toxicity assessment of
788 contaminated soil by oral or dermal uptake in land snails: metal bioavailability
789 and bioaccumulation, *Environ. Toxicol. Chem. Int. J.*, 21, 820–827,
790 <https://doi.org/10.1002/etc.5620210419>.

791 Guessasma, Z., Khaldi, F., Grara, N., Agouni, M., Sleimi, N. and Benslama, M.
792 (2020). Assessment of heavy metal soil contamination in some northeastern
793 Algerian biotopes by using the terrestrial snail, *Helix aspersa*, *Studia Univ.*
794 *Vasile Goldis Arad, Ser. Stiint. Vietii Life Sci. Ser.*, 30, 55–63,
795 <https://www.univ-soukahrass.dz/wpuploads/eprints/2021-992-681d3.pdf>.
796 Harrell, J. F. (2023). Hmisc: Harrell Miscellaneous, R package version 5.1-1.
797 He, G., Zhang, Z., Wu, X., Cui, M., Zhang, J. and Huang, X. (2020). Adsorption
798 of heavy metals on soil collected from *lixisol* of typical karst areas in the
799 presence of CaCO_3 and soil clay and their competition behavior, *Sustain.*
800 *Switz.*, 12, 7315, <https://doi.org/10.3390/SU12187315>.
801 Hermes, A. L., Dawson, T. E. and Hinckley, E. L. S. (2022). Sulfur isotopes reveal
802 agricultural changes to the modern sulfur cycle, *Environ. Res. Lett.*, 17,
803 054032, <https://doi.org/10.1088/1748-9326/ac6683>.
804 Hermes, A. L., Ebel, B. A., Murphy, S. F. and Hinckley, E. L. S. (2021). Fates
805 and fingerprints of sulfur and carbon following wildfire in economically
806 important croplands of California, US, *Sci. Total Environ.*, 750, 142179,
807 <https://doi.org/10.1016/j.scitotenv>.
808 Hidouri, N., Moussaoui, Z., Sleimi, N., Hamed, Y. and Hamzaoui-Azzaza, F.
809 (2024). Soil contamination by trace elements and radioelements and related
810 environmental risks in agricultural soils of the M'Dhilla Basin (southwestern
811 Tunisia). *Environmental Monitoring and Assessment*, 196(11), 1024.

812 Hoang, T. C., Rogevich, E. C., Rand, G. M. and Frakes, R. A. (2008). Copper
813 uptake and depuration by juvenile and adult Florida apple snails (*Pomacea*
814 *paludosa*), *Ecotoxicol.*, 17, 605–615, [https://doi.org/10.1007/s10646-008-](https://doi.org/10.1007/s10646-008-0243-8)
815 [0243-8](https://doi.org/10.1007/s10646-008-0243-8).

816 Huang, F., Peng, L., Zhang, J., Lin, W. and Chen, S. (2018). Cadmium
817 bioaccumulation and antioxidant enzyme activity in hepatopancreas, kidney,
818 and stomach of invasive apple snail *Pomacea canaliculata*, *Environ. Sci.*
819 *Pollut. Res.*, 25, 18682–18692, <https://doi.org/10.1007/s11356-018-2092-1>.

820 Ihaka, R. and Gentleman, R. (1996). R: A language for data analysis and graphics,
821 *Comput. Graph. Stat.*, 5, 299–314.

822 Kadhodaie, A., Kelich, S. and Baghbani, A. (2012). Effects of salinity levels on
823 heavy metals (Cd, Pb, and Ni) absorption by sunflower and sudangrass plants,
824 *Bull. Environ. Pharm. Life Sci.*, 1, 47–53.

825 Kassambara, A. (2023a). ggcorrplot: Visualization of a Correlation Matrix using
826 'ggplot2', R package version 0.1.4.1.

827 Kassambara, A. (2023b). ggpubr: 'ggplot2' Based Publication Ready Plots, R
828 package version.

829 Kassambara, A. and Mundt, F. (2020). factoextra: Extract and Visualize the
830 Results of Multivariate Data Analyses, R package version 1.0.7.

831 Kicińska, A., Pomykała, R. and Izquierdo-Diaz, M. (2022). Changes in soil pH
832 and mobility of heavy metals in contaminated soils, *Eur. J. Soil Sci.*, 73,
833 e13203, <https://doi.org/10.1111/ejss.13203>.

834 Kim, S. H., Bae, S. and Hwang, Y. S. (2023). Comparative bioaccumulation,
835 translocation, and phytotoxicity of metal oxide nanoparticles and metal ions in
836 soil-crop system, *Sci. Total Environ.*, 856, 158938,
837 <https://doi.org/10.1016/j.scitotenv.2022.158938>.

838 Kjeldahl, J. (1883). Neue Methode zur Bestimmung des Stickstoffs in organischen
839 Körpern, *Fresenius' Z. Anal. Chem.*, 22, 366–382,
840 <https://doi.org/10.1007/BF01338151>.

841 Lag, J., Hadas, A., Fairbridge, R. W., Nóvoa Muñoz, J. C., Pombal, X. P.,
842 Cortizas, A. M., Almendros, G., Chesworth, W., Arbestain, M. C., Macías, F.,
843 Martínez, C. A., Chesworth, W., Jaynes, W. F. and Winterkorn, H. F. (2008).
844 Hygroscopicity hygroscopy constant, *Encycl. Soil Sci.*, Springer.

845 Larba, R., Zeraib, A. and Soltani, N. (2023). Assessment of soil metallic
846 contamination in several sites from Northeast Algeria by use of terrestrial
847 gastropod: *Cornu aspersum* (O.F. Müller, 1774) (Helicidae), *Biodivers. J.*, 14,
848 373–383, <https://doi.org/10.31396/biodiv.jour.2023.14.2.373.383>.

849 Laurent, C., Bravin, M. N., Crouzet, O., Pelosi, C., Tillard, E., Lecomte, P. and
850 Lamy, I. (2020). Increased soil pH and dissolved organic matter after a decade
851 of organic fertilizer application mitigates copper and zinc availability despite
852 contamination, *Sci. Total Environ.*, 709, 135927,
853 <https://doi.org/10.1016/j.scitotenv.2019.135927>.

854 Lê, S., Josse, J., Rennes, A. and Husson, F. (2008). FactoMineR: An R Package
855 for Multivariate Analysis, *J. Stat. Softw.*, 25, 1–18, <http://www.jstatsoft.org/>.

856 Li, L., Wu, H., Van Gestel, C. A. M., Peijnenburg, W. J. G. M. and Allen, H. E.
857 (2014). Soil acidification increases metal extractability and bioavailability in
858 old orchard soils of Northeast Jiaodong Peninsula in China, *Environ. Pollut.*,
859 188, 144–152, <https://doi.org/10.1016/j.envpol.2014.02.003>.

860 Li, Q., Gao, Y. and Yang, A. (2020). Sulfur homeostasis in plants, *Int. J. Mol.*
861 *Sci.*, 21, 1–16, <https://doi.org/10.3390/ijms21238926>.

862 Li, R., Tan, W., Wang, G., Zhao, X., Dang, Q., Yu, H. and Xi, B. (2019). Nitrogen
863 addition promotes the transformation of heavy metal speciation from
864 bioavailable to organic bound by increasing the turnover time of organic
865 matter: An analysis on soil aggregate level, *Environ. Pollut.*, 255, 113170,
866 <https://doi.org/10.1016/j.envpol.2019.113170>.

867 Li, X., Liang, H., Zeng, Y., Zheng, X., Ren, Z. and Mai, B. (2023). Trophic
868 transfer of heavy metals in a wetland food web from an abandoned e-waste
869 recycling site in South China, *Sci. Total Environ.*, 890, 164327.

870 Lobo-da-Cunha, A. (2019). Structure and function of the digestive system in
871 molluscs, *Cell Tissue Res.*, 377, 475–503, [https://doi.org/10.1007/s00441-](https://doi.org/10.1007/s00441-019-03085-9)
872 [019-03085-9](https://doi.org/10.1007/s00441-019-03085-9).

873 Louzon, M., de Vaufleury, A. and Capelli, N. (2023). Ecogenotoxicity assessment
874 with land snails: A mini-review, *Mutat. Res. Rev. Mutat. Res.*, 792, 108472,
875 <https://doi.org/10.1016/j.mrrev.2023.108472>.

876 Malik, K. M., Khan, K. S., Akhtar, M. S. and Ahmed, Z. I. (2020). Sulfur
877 distribution and availability in alkaline subtropical soils affected by organic

878 amendments, J. Soil Sci. Plant Nutr., 20, 2253–2266,
879 <https://doi.org/10.1007/s42729-020-00292-0>.

880 Maruthai, S., Rajendran, S., Selvanarayanan, R. and Gowri, S. (2025)
881 “Wastewater Recycling Integration with IoT Sensor Vision for Real-time
882 Monitoring and Transforming Polluted Ponds into Clean Ponds using HG-RNN”,
883 Global NEST Journal. 27(4). <https://doi.org/10.30955/gnj.06758>.

884 Maruyama-Nakashita, A. and Ohkama-Ohtsu, N. (2017). Sulfur assimilation and
885 glutathione metabolism in plants, in Glutathione in Plant Growth,
886 Development, and Stress Tolerance, Springer Int. Publ., pp 287–308,
887 https://doi.org/10.1007/978-3-319-66682-2_13.

888 Mitev, T., Dedov, I. K. and Hristovski, S. (2023). Contents of Zn, Cu, Mn and Cd
889 in soil, plants and Turkish snails *Helix lucorum* Linnaeus, 1758 in Skopje,
890 Republic of North Macedonia, Acta Zool. Bulg., 75, 2.

891 Mohamed, B., Abdelkader, D. and Benchaben, H. (2016). Mobilité du plomb et
892 du zinc issus de retombées atmosphériques dans le sol : Cas de la zone
893 industrielle de Tiaret, Algérie, Eur. Sci. J., ESJ, 12, 131,
894 <https://doi.org/10.19044/esj.2016.v12n18p131>.

895 Monchanin, C., Devaud, J. M., Barron, A., Lihoreau, M. and Barron, A. B. (2021).
896 Current permissible levels of metal pollutants harm terrestrial invertebrates,
897 Sci. Total Environ., 779, 146398,
898 <https://doi.org/10.1016/j.scitotenv.2021.146398>.

899 Mukhtorova, D., Hlava, J., Száková, J., Najmanová, J. and Tlustoš, P. (2023). Can
900 mollusks or insects serve as bioindicators of the risk element polluted area?
901 Gastropods (Gastropoda) versus leaf beetles (Coleoptera: Chrysomelidae),
902 Environ. Sci. Pollut. Res., 30, 78707–78717.

903 Mwelwa, S., Chungu, D., Tailoka, F., Beesigamukama, D. and Tanga, C. (2023).
904 Biotransfer of heavy metals along the soil-plant-edible insect-human food
905 chain in Africa, Sci. Total Environ., 881, 163150,
906 <https://doi.org/10.1016/j.scitotenv.2023.163150>.

907 N'Doua, A. D. Y., Adjessan, M. A. M. and Koffi, K. M. (2024). Détermination du
908 niveau de contamination en métaux lourds (arsenic, cadmium, mercure et
909 plomb) de quatre espèces de poissons consommées par les familles de
910 pêcheurs de Jacquville, Eur. Sci. J., ESJ, 20, 53,
911 <https://doi.org/10.19044/esj.2024.v20n3p53>.

912 Ning, Q., Shao, B., Huang, X., He, M., Tian, L. and Lin, Y. (2024).
913 Bioaccumulation, biomagnification, and ecological risk of trace metals in the
914 ecosystem around oilfield production area: A case study in Shengli Oilfield,
915 Environ. Monit. Assess., 196, 87, [https://doi.org/10.1007/s10661-024-
916 102566](https://doi.org/10.1007/s10661-024-102566).

917 Nwagu, C. U., Olujimi, O. O. and Okoro, G. C. (2022). The use of Giant African
918 Snails (*Archachatina marginata* and *Achatina fulica*) as bio-indicators of
919 heavy metal pollution, IAA J. Sci. Res., 8, 19–41, ISSN: 2736-7319, Available
920 at: <https://www.researchgate.net/publication/364348221>.

921 Ogbeide, O. and Amayanvbo, O. (2024). Effect Of Heavy Metal Runoff from
922 Ikpoba River on The Gonad and Antioxidant Properties of African Snail
923 (*Bulinus Africanus*). African Journal of Health, Safety and Environment,
924 5(2), 10-25.

925 Okalebo, J. R., Gathua, K. W. and Woomer, P. L. (2002). Laboratory methods of
926 soil and plant analysis: A working manual, second edition, Sacred Africa,
927 Nairobi, pp. 21–26.

928 Olsen, S. R. (1954). Estimation of available phosphorus in soils by extraction with
929 sodium bicarbonate (No. 939), US Department of Agriculture, Available at:
930 <http://books.google.com>.

931 Owojori, O. J., Awodiran, M., Ayanda, O. E. and Jegede, O. O. (2022). Toxicity
932 and accumulation of lead and cadmium in the land snail, Archachatina
933 papyracea, in a tropical Alfisol from Southwestern Nigeria, Environ. Sci.
934 Pollut. Res., 29, 44917–44927.

935 Park, H. J., Kim, S. U., Jung, K. Y., Lee, S., Choi, Y. D., Owens, V. N., Kumar,
936 S., Yun, S. W. and Hong, C. O. (2021). Cadmium phytoavailability from 1976
937 through 2016: Changes in soil amended with phosphate fertilizer and compost,
938 Sci. Total Environ., 762, 1–10.

939 Periasamy, S., Subramanian, P. and Surendran, R. (2024) “An intelligent air
940 quality monitoring system using quality indicators and Transfer learning based
941 Lightweight recurrent network with skip connection”, Global NEST, 26(5), pp.
942 1–10.

943

944 Pohlert, T. (2023). PMCMRplus: Calculate Pairwise Multiple Comparisons of
945 Mean Rank Sums Extended, R package version 1.9.7.

946 R Core Team. (2022). R: A language and environment for statistical computing,
947 Available at: <https://www.R-project.org/>.

948 Rašković, B., Poleksić, V., Jarić, I., Skorić, S., Topisirović, G. and Stojnić, B.
949 (2023). Accumulation of metal trace elements in different body parts of
950 terrestrial Roman snail *Helix pomatia* L., 1758 on three polluted sites in Serbia,
951 *Environ. Sci. Pollut. Res.*, 30(8), 21853–21862,
952 <https://doi.org/10.1007/s11356-022-23697-z>.

953 Selvanarayanan, R., Rajendran, S., Pappa, K. C. and Thomas, B. (2024b).
954 “Wastewater Recycling to Enhance Environmental Quality using Fuzzy
955 Embedded with RNNIoT for Sustainable Coffee Farming”. *Global NEST*
956 *Journal*. Available at: <https://doi.org/10.30955/gnj.06346>.

957 Selvanarayanan, R., Rajendran, S., Algburi, S., Khalaf, O.I. and Hamam, H.
958 (2024a). “Empowering coffee farming using counterfactual recommendation
959 based RNN driven IoT integrated soil quality command system,” *Scientific*
960 *Reports* 14, 6269. <https://doi.org/10.1038/s41598-024-56954-x>

961 Revelle, W. (2023). *psych: Procedures for Psychological, Psychometric, and*
962 *Personality Research*, R package version 2(9).

963 Rhoades, J. D. (1982). *Soluble salts, Methods of Soil Analysis: Part 2 Chemical*
964 *and Microbiological Properties*, 2nd ed., American Society of Agronomy,
965 Madison, WI, pp. 849–867.

966 Sahraoui, A. S. and Sahli, L. (2022). Utilisation in situ et ex situ de *Helix aspersa*
967 comme bioindicateur de la contamination des sols par le cadmium et le plomb,
968 Thèse de doctorat, Université Frères Mentouri-Constantine 1.

969 Salao, A. B., Diop, C., Livardjani, F. and Djokhane, A. M. (2017). Évaluation des
970 risques de la pollution atmosphérique (CO, NO_x, SO₂, O₃, PM₁₀ et PM_{2.5})
971 de la ville de Dakar, Sénégal, Rev. Ivoir. Sci. Technol., 30, 1–10,
972 <http://www.revist.ci>.

973 Salih, A. H. S., Hama, A. A., Hawrami, K. A. M. and Ditta, A. (2021). The land
974 snail, *Eobania vermiculata*, as a bioindicator of the heavy metal pollution in
975 the urban areas of Sulaimani, Iraq, Sustainability (Switzerland), 13(24),
976 <https://doi.org/10.3390/su132413719>.

977 Sargsyan, A., Hovhannisyan, G., Simonyan, A., Arakelyan, M., Arzumanyan, M.
978 and Aroutiounian, R. (2022). Application of land snail *Helix lucorum* for
979 evaluation of genotoxicity of soil pollution, Mutat. Res./Genet. Toxicol.
980 Environ. Mutagen., 878, 503500.

981 Sebban, H., Ait Belcaid, H., El Alaoui El Fels, A., Bouriqi, A., Pineau, A. and
982 Sedki, A. (2022). Trace element bioaccumulation in the edible milk snail
983 (*Otala lactea*) and Cabrilla (*Otala punctata*) in Marrakech, Morocco, Appl.
984 Ecol. Environ. Res., 20(1), 875–892,
985 https://doi.org/10.15666/aeer/2001_875892.

986 Shaffique, S., Kang, S. M., Ashraf, M. A., Umar, A., Khan, M. S., Wajid, M., Al-
987 Ghamdi, A. A. and Lee, I. J. (2024). Research progress on migratory water

988 birds: indicators of heavy metal pollution in Inland Wetland resources of
989 Punjab, Pakistan, *Water* (Switzerland), 16(8), 1163,
990 <https://doi.org/10.3390/w16081163>.

991 Shahid, M., Dumat, C., Khalid, S., Niazi, N. K. and Antunes, P. M. (2017).
992 Cadmium bioavailability, uptake, toxicity and detoxification in soil-plant
993 system, *Rev. Environ. Contam. Toxicol.*, 241, 73–137.

994 Sintorini, M. M., Widyatmoko, H., Sinaga, E. and Aliyah, N. (2021). Effect of pH
995 on metal mobility in the soil, *IOP Conf. Ser. Earth Environ. Sci.*, 737(1),
996 012071, <https://doi.org/10.1088/1755-1315/737/1/012071>.

997 Sleimi, N., Bankaji, I., Kouki, R., Dridi, N., Duarte, B. and Caçador, I. (2022).
998 Assessment of extraction methods of trace metallic elements in plants:
999 approval of a common method, *Sustainability* (Switzerland), 14(3), 1428,
1000 <https://doi.org/10.3390/su14031428>.

1001 Soliman, M. M., Hesselberg, T., Mohamed, A. A. and Renault, D. (2022). Trophic
1002 transfer of heavy metals along a pollution gradient in a terrestrial agro-
1003 industrial food web, *Geoderma*, 413, 115748.

1004 Stevenson, F. J. (1965). Origin and distribution of nitrogen in soil, *Soil Nitrogen*,
1005 10, 1–42.

1006 Sungur, A., Temel, E., Everest, T., Soylak, M. and Özcan, H. (2023). Effects of
1007 soil texture on trace metal concentrations and geochemical fractions in the soil
1008 of apple orchards (Çanakkale, NW Turkey), *Arch. Agron. Soil Sci.*, 69, 1–10.

1009 Téreygeol, F., Arles, A., Foy, E., Florsch, N. and Llubes, M. (2010). Dosages par
1010 fluorescence X portable d'ateliers médiévaux de production des métaux non-
1011 ferreux, *ArchéoSciences*, 34, 243–252,
1012 <https://doi.org/10.4000/archeosciences.2802>.

1013 Thomas, G. W. (1982). Exchangeable cations, *Methods of Soil Analysis: Part 2*
1014 *Chemical and Microbiological Properties*, 9, 159–165.

1015 van Gestel, C. A. M. (2008). Physico-chemical and biological parameters
1016 determine metal bioavailability in soils, *Sci. Total Environ.*, 406(3), 385–395,
1017 <https://doi.org/10.1016/j.scitotenv.2008.05.050>.

1018 Walkley, A. and Black, I. A. (1934). An examination of the Degtjareff method for
1019 determining soil organic matter, and a proposed modification of the chromic
1020 acid titration method, *Soil Sci.*, 37, 29–38.

1021 Wei, X., Zhang, P., Zhan, Q., Hong, L., Bocharnikova, E. and Matichenkov, V.
1022 (2021). Regulation of As and Cd accumulation in rice by simultaneous
1023 application of lime or gypsum with Si-rich materials, *Environ. Sci. Pollut.*
1024 *Res.*, 28(6), 7271–7280, <https://doi.org/10.1007/s11356-020-11053-y>.

1025 Weindorf, D. C. and Chakraborty, S. (2020). Portable X-ray fluorescence
1026 spectrometry analysis of soils, *Soil Sci. Soc. Am. J.*, 84(5), 1384–1392,
1027 <https://doi.org/10.1002/saj2.20151>.

1028 Wickham, H. (2016). *Data Analysis*, in: *ggplot2*, Springer, Cham, pp. 145–183,
1029 https://doi.org/10.1007/978-3-319-24277-4_9.

1030 Włostowski, T., Kozłowski, P., Łaskiewicz-Tiszchenko, B. and Oleńska, E.
1031 (2016). Cadmium accumulation and pathological alterations in the midgut
1032 gland of terrestrial snail *Helix pomatia* L. from a zinc smelter area: role of soil
1033 pH, Bull. Environ. Contam. Toxicol., 96, 484–489,
1034 <https://doi.org/10.1007/s00128-016-1748-0>.

1035 Yu, H. Y., Liu, C., Zhu, J., Li, F., Deng, D. M., Wang, Q. and Liu, C. (2016).
1036 Cadmium availability in rice paddy fields from a mining area: The effects of
1037 soil properties highlighting iron fractions and pH value, Environ. Pollut., 209,
1038 38–45.

1039 Ibrahim, O. Z., Tankari, D. B. A., Guero, Y., Maissoro, M. I. F., Feidt, C.,
1040 Sterckeman, T. and Echevarria, G. (2019). Spatial distribution of metallic trace
1041 elements in soils of Komabangou gold zone in Niger, Int. J. Biol. Chem. Sci.,
1042 13(1), 557–573.

1043 Zhang, S., Ni, X., Arif, M., Yuan, Z., Li, L. and Li, C. (2020). Salinity influences
1044 Cd accumulation and distribution characteristics in two contrasting
1045 halophytes, *Suaeda glauca* and *Limonium aureum*, Ecotoxicol. Environ. Saf.,
1046 191, 110230.

1047 Zhu, Q., Ji, J., Tang, X., Wang, C. and Sun, H. (2023). Bioavailability assessment
1048 of heavy metals and organic pollutants in water and soil using DGT: A review,
1049 Appl. Sci., 13(17), 9760.