

1 **Investigation of Cephalexin Removal Using Biochar Derived from *Nephelium***  
2 ***lappaceum* Seeds**

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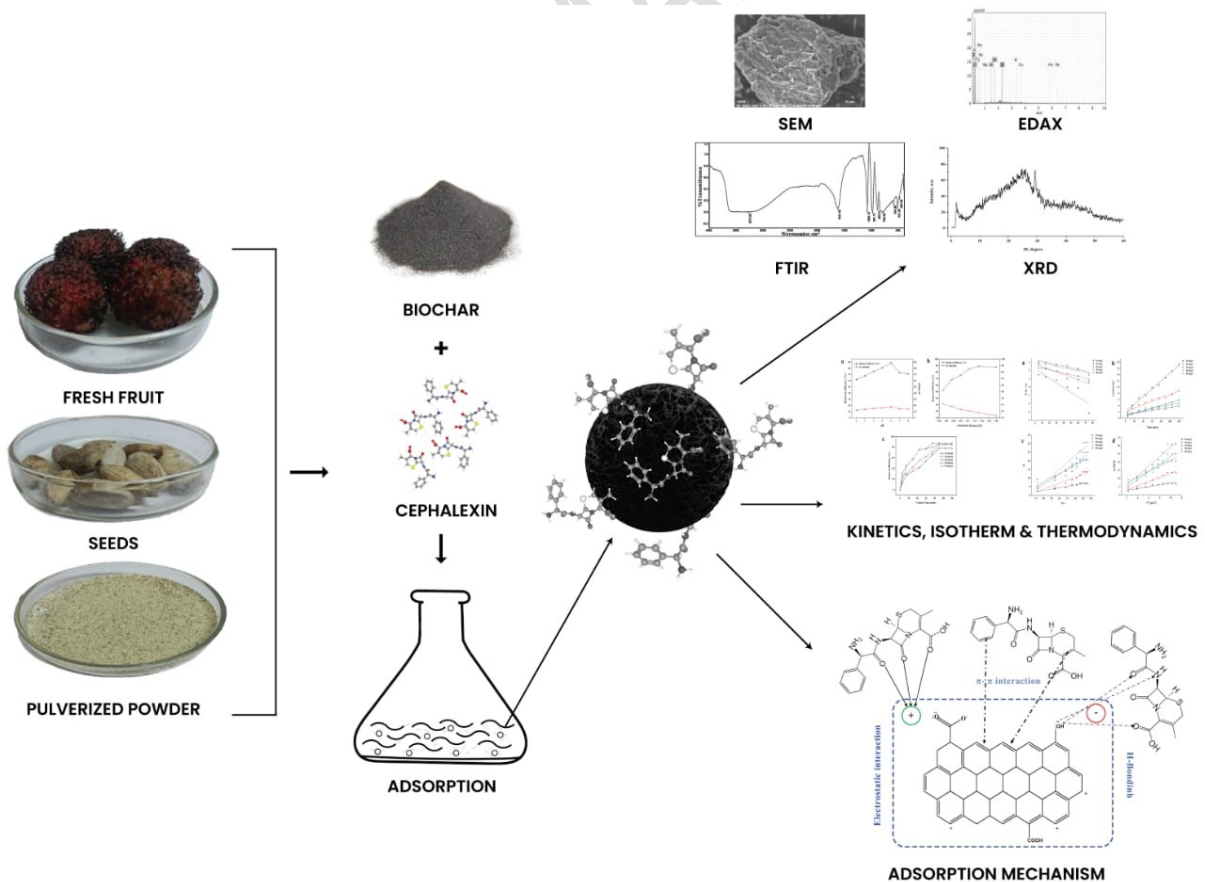
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11  
12 **GRAPHICAL ABSTRACT**



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14

## 15 ABSTRACT

16 The improper disposal of drugs into bodies of water has become a severe environmental and  
17 health issue. Biochar synthesized from different agro residues could be a potential material  
18 for the adsorption of pollutants in aqueous environments. *Nephelium lappaceum* seeds were  
19 pyrolyzed at 600 °C in a pyrolysis reactor incorporating a slow pyrolysis process for  
20 carbonization. Fourier-Transform Infrared (FTIR), Scanning Electron Microscope (SEM),  
21 and X-ray diffraction (XRD) were used to characterize the biochar before and after  
22 adsorption of the pharmaceutical pollutant cephalexin. The effects of several factors,  
23 including initial cephalexin concentration, time of contact, dosage of adsorbent, and pH, were  
24 considered for the sorption. The findings showed that the Freundlich model offered the  
25 greatest fit for adsorption. The better-fitted kinetic model was the pseudo-second-order  
26 kinetic. Based on Langmuir isotherm model, the maximum adsorption capacity of cephalexin  
27 was obtained as 63.69 mg/g. The adsorption process involves hydrogen bonding, surface  
28 complexation, electrostatic and  $\pi$ - $\pi$  EDA interactions. The current study offers a feasible and  
29 optimistic method for using agricultural waste and an alternative adsorbent substance for  
30 recovering highly concentrated cephalexin from water-based solutions.

31 **Keywords:** Drug disposal, Antibiotics, Biomass waste, Isotherms, Kinetics

## 32 1. INTRODUCTION

33 One of the most significant moments in medical history was the discovery of antibiotics, and  
34 according to statistics, more than 200,235 metric tons of antimicrobial agents will be required  
35 to eradicate growth and prevent bacterial diseases (Xu *et al.* 2022). Cephalexin, often known  
36 as CPX and having the technical name 7-(D-a-Amino-a-phenylacetamido)-3-methyl-3-  
37 cephem-4-carboxylic acid monohydrate ( $C_{16}H_{17}O_4 N_3SH_2O$ ) with a molecular mass of 365.40  
38 g/mol, is an antibiotic of the first generation called cephalosporin, with an annual  
39 consumption of 3,000 tons. CPX is one of the world's most widely-used antibiotics,

40 according to reports. It belongs to the class of antibiotics known as beta-lactams and is  
41 discharged into the environment through sewage, hospital wastewater, pharmaceutical  
42 companies, animals, and agriculture. Environmental exposure to  $\beta$ -lactam antibiotic  
43 compounds occurs because of their use as components or intermediates for internal use  
44 (Wang *et al.* 2021). The continuous entry of CPX into the environment and water supplies  
45 will lead to environmental difficulties and issues for humans. To maintain water quality and  
46 safeguard public health, it is crucial to remove antibiotics, including CPX, from contaminated  
47 aqueous media (Arab *et al.* 2022). There are several innovative methods for eliminating  
48 antibiotics, including photocatalysis, fenton oxidation, ozonation, wet oxidation, membrane  
49 technology, adsorption, hybrid technology, aerobic and anaerobic treatment, and  
50 electrochemical oxidation. Among various methods for antibiotic removal, adsorption is  
51 highly effective because of its versatility, high selectivity, and ease of implementation across  
52 diverse water sources, offering several advantages in the process. Adsorption methods are  
53 employed to remove organic, inorganic, heavy metals, and microplastics (Phoon *et al.* 2020,  
54 Varma *et al.* 2024).

55 When organic raw materials are pyrolyzed in a low-oxygen environment to form biochar, a  
56 carbon-rich substance, as indicated above, the feedstock can come from a variety of sources  
57 (Wang *et al.* 2019). Because of its porous nature, substantial surface area, functional group  
58 incorporation, elevated capacity for cation exchange (CEC), and efficient removal of  
59 contaminants, biochar, a sustainable resource, serves as a proficient adsorbent (Karthik *et al.*  
60 2023). Through pore filling, surface sorption, electrostatic interaction,  $\pi$  orbital interactions,  
61 electrostatic interaction, and ion exchange mechanisms, biochar eliminates contaminants (Al-  
62 Gheethi *et al.* 2021). Biochar can also be represented as BC, which can be made from  
63 agricultural and other natural waste components such as fruit seed waste. Many studies have  
64 documented biochar synthesis from seed waste biomass such as watermelon seeds, mango

65 seeds, jackfruit seed waste, *Prosopis juliflora* seed waste, and *Schizizium commune* seed to  
66 obtain low-cost substances that can be used for the elimination of several environmental  
67 contaminants, mainly heavy metals, dyes, and antibiotics (Periyasamy *et al.* 2022, Velusamy  
68 *et al.* 2021, Khadem *et al.* 2023, Diaz-Uribe *et al.* 2022, Kandasamy *et al.* 2022). As an  
69 alternative adsorbant, *Nephelium lappaceum* fruit seeds were used to generate biochar to  
70 eliminate cephalixin.

71 The tropical fruit *Nephelium lappaceum*, also referred to as rambutan, is indigenous to  
72 Southeast Asia and some portions of India under appropriate conditions, particularly in the  
73 southern sections. The fruit, characterized by a hairy, red or yellow outer skin and  
74 translucent, juicy flesh with seeds within the fleshy aril, provides a promising feedstock for  
75 biochar production because of the seeds' high cellulose content and unique chemical  
76 composition. The process involves subjecting the seeds to pyrolysis and thermochemical  
77 decomposition without oxygen, resulting in the transformation of the biomass into a stable,  
78 carbonaceous material with a porous structure. The cation exchange capacity, high surface  
79 area, and functional groups of BC derived from rambutan seeds are all potentially  
80 advantageous features. Despite being consumed globally for its health benefits, the  
81 unpalatable nature of rambutan seeds has led to their disposal. Converting these discarded  
82 seeds into biochar offers a potential solution for effective by-product use (Batoool *et al.* 2022,  
83 Naveen and Muthumari 2024). To overcome some of the shortcomings of conventional  
84 adsorbents with low kinetics and poor adsorption capacity, *Nephelium lappaceum* seed  
85 biochar was employed as an adsorbent for successful antibiotic removal.

86 In this study, biochar was prepared using *Nephelium lappaceum* seeds under pyrolysis.  
87 Biochar was used to adsorb cephalixin. FTIR, SEM with EDAX, and XRD were used to  
88 characterize the synthetic biochar. The impacts of various operating parameters, such as pH,  
89 temperature, contact time, and cephalixin concentration, were also investigated. Furthermore,

90 the adsorption process was determined by analyzing the characteristics of the biochar before  
91 and after cephalixin adsorption, as well as examining the adsorption kinetics, isotherms, and  
92 thermodynamics.

## 93 **2. EXPERIMENTAL**

### 94 **2.1 Collection of Biomass and Biochar Preparation**

95 *Nephelium lappaceum* seeds were collected from a nearby village located near  
96 Tenkasi, Tamil Nadu, India. After thorough cleaning with tap water and deionized water, it  
97 was left to dry for 3 days under sunlight. To remove the last traces of moisture the seeds were  
98 kept in an oven at 50°C for 2 days over night. After drying, it was crushed with a mortar and  
99 pestle and used to make biochar (BC). *Nephelium lappaceum* seeds that had been processed  
100 were pyrolyzed in a furnace carbonized at 600°C for one hour at a heating rate of 3°C/min  
101 with slow pyrolysis. The oxygen-free environment in the pyrolyzer was achieved by purging  
102 inert gases such as nitrogen before and during pyrolysis. The produced BC was removed  
103 from the furnace and then cooled to room temperature in desiccator. For later use, *Nephelium*  
104 *lappaceum* seeds BC were kept in an airtight container (Chakraborty *et al.* 2018,  
105 Venkateswaran *et al.* 2023).

### 106 **2.2 Preparation of Cephalixin Solution**

107 Commercially available cephalixin -250 mg tablets were purchased in their impure form. The  
108 tablets were available in powdered form it was dissolved in the distilled water after removing  
109 their outer covering. The optimal wavelength for cephalixin was determined using  
110 UV/visible spectrometry. A 100 ppm stock solution of cephalixin was prepared in a 250 mL  
111 volumetric flask. From this stock solution, standard solutions with cephalixin concentrations  
112 of 2, 4, 6, 8, 10, 12, and 14 ppm were prepared. The optimized wavelength obtained was 251  
113 nm (Velusamy *et al.* 2021).

### 114 **2.3 Point of Zero Charge**

115 The pH Point of Zero charge (pzc) was ascertained by adding salt with BC. Twenty milliliters  
116 of 0.1M NaNO<sub>3</sub> solutions were prepared by varying the pH from 3 to 12 at every 1 pH  
117 interval, which is represented as the initial pH (pH<sub>i</sub>).After adding 0.20 g of biochar, the  
118 aforementioned solutions were stirred for a whole day.The solution's final pH (pH<sub>f</sub>) was  
119 measured after 24 hours. Plotting the graph of (pH<sub>i</sub>-pH<sub>f</sub>) versus (pH<sub>i</sub>) showed the pzc value  
120 (Adam *et al.* 2016).

#### 121 **2.4 Characterization of Biochar**

122 The morphology of the surface of the sample was analyzed by SEM-EDAX (Supra 55-Carl  
123 Zeiss, Germany). In order to investigate the structure and crystalline character of biochar  
124 before and after adsorption, XRD test was conducted at a voltage of 9 kW and radiation of  $\lambda$   
125 = 1.54 Å (Rikagu Japan). Functional groups present in the BC were determined using FTIR  
126 (Shimadzu 8400s) before and after adsorption within a wavenumber region of 0 to 4000 cm<sup>-1</sup>  
127 (Li *et al.* 2018).

#### 128 **2.5 Batch Adsorption Study**

129 Batch adsorption studies were evaluated using the synthesized BC to remove cephalixin  
130 (CPX). To determine the effects of agitation time (0 to 120 mins), initial cephalixin  
131 concentration (10 to 50 mg/L), BC dosage (0.05 to 0.2 g/L), pH (2-8), and temperature  
132 (303.15 to 333.15K), the removal of CPX was tested under various experimental conditions.  
133 After optimizing the parameters, the adsorption isotherms were determined. 100 mL of CPX  
134 solution was taken in 250 mL conical flask containing BC sample was kept in an orbital  
135 shaker (Neolab Incubator Shaker) with an agitating speed of 150 rpm. A PTFE filter with a  
136 mesh size of 0.45µm was used to filter the sample, and to quantify concentration, a UV-  
137 visible spectrophotometer at a wavelength of 251nm was utilized (Dai *et al.* 2020). In this  
138 study, the adsorbed CPX concentration  $q_{(t)}$  was determined using the following equation:

$$139 \quad q_{(t)} = \frac{V(C_0 - C_t)}{W} \quad (1)$$

140 where V - denotes the sample volume (L), W - represents the adsorbent mass (g), and C<sub>0</sub> is  
141 the initial concentration of CPX in solution (mg/L) and C<sub>t</sub> - represents the concentration of  
142 cephalixin at time t (mg/L).

## 143 2.6 Adsorption Isotherm

144 The adsorption isotherms of CPX, a BC dose of 1.25g/L and pH 6 were maintained  
145 throughout the experiment. After adding BC, the temperature 30° C and contact time was 90  
146 min. Then, the adsorption isotherms of cephalixin by *Nephelium lappaceum* BC were  
147 performed through isotherm models such as Langmuir (Equation2), Temkin (Equation3), and  
148 Freundlich (Equation4).

$$149 \frac{C_e}{q_e} = \frac{1}{q_m} K_L + \frac{C_e}{q_m} \quad (2)$$

$$150 q_e = \frac{RT}{b_T} \ln K_T + \frac{RT}{b_T} \ln C_e \quad (3)$$

$$151 \ln q_e = \ln K_f + \frac{1}{n} C_e \quad (4)$$

152

153 where, K<sub>L</sub> illustrates the Langmuir constant with respect to adsorption energy and q<sub>m</sub>  
154 represents the maximum sorption capacity (mg/g), K<sub>f</sub> indicates the Freundlich sorption  
155 capacity (mg/g), and the symbol n denotes the Freundlich constant, a measure of the  
156 attraction between the adsorbent and the adsorbate that is correlated to surface heterogeneity,  
157 the heat of adsorption is related to the Temkin constant (b<sub>T</sub>), the maximal binding energy  
158 (L/mg) is related to the equilibrium binding constant (K<sub>T</sub>), the ideal gas constant (8.314  
159 J/mol/K) is related to R, and the absolute temperature is related to T (K) (Karthik *et al.* 2020).

## 160 3. RESULTS AND DISCUSSION

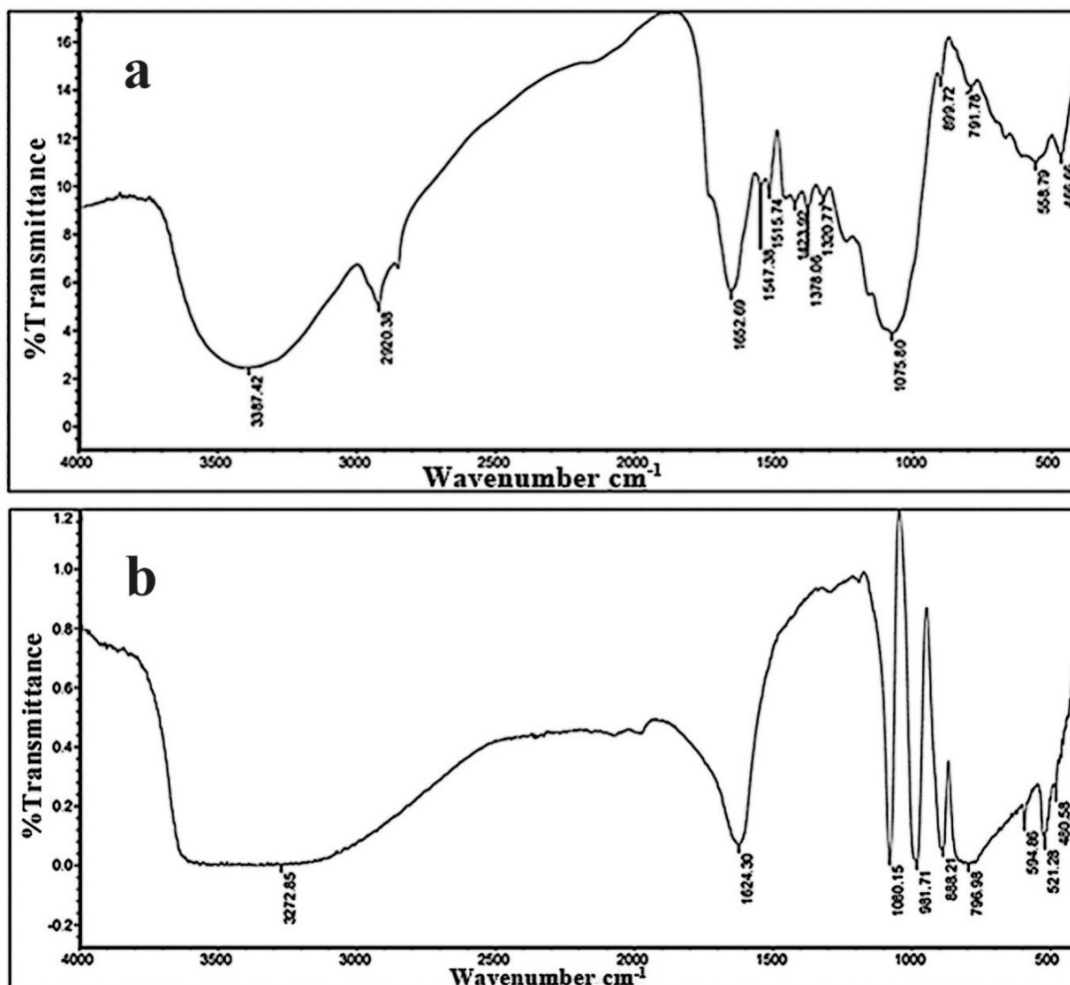
### 161 3.1 Characterization Studies

#### 162 3.1.1. FTIR analysis

163 The FTIR spectra of the produced biochar and the one with and without antibiotics showed  
164 the functional groups on the surface of the biochar (Figure 1). It is imperative to identify  
165 functional groups, which include organic, inorganic, and polymeric functional groups when  
166 synthesizing biochar to understand the adsorption properties of the produced biochar. It  
167 should be noted that the functional groups in the BC atoms may not be an exact match due to  
168 molecular interactions that can affect the BC atoms, as seen from the FTIR spectra of the  
169 samples. The FTIR spectra of the *Nephelium lappaceum* seed-derived biochar and the  
170 corresponding FTIR spectra after CPX adsorption were obtained over the range of 400–4000  
171  $\text{cm}^{-1}$ . The results depict that there are large bands in the region 3400-3300  $\text{cm}^{-1}$ , which is the  
172 characteristic of O-H stretch owing to hydrogen bonding. This peak shifts from a broad peak  
173 at around 3400  $\text{cm}^{-1}$  after adsorption to a narrower range between 3300  $\text{cm}^{-1}$  and 3200  $\text{cm}^{-1}$ .  
174 The C-H group is seen at 2920  $\text{cm}^{-1}$  only in BC which suggests that hydrogen is quite  
175 significantly displaced during the adsorption of CPX. The peaks located at 1652  $\text{cm}^{-1}$  and  
176 1624  $\text{cm}^{-1}$  are the carbonyl C=O group that is present before and after the samples' exposure  
177 to CPX. The small peaks at approximately 1515  $\text{cm}^{-1}$  and 1547  $\text{cm}^{-1}$  can be attributed to  
178 C=C stretching. Also, the shift of the peak at around 1600  $\text{cm}^{-1}$  associated with the C=C  
179 bond of phenol and the new peak at 1080  $\text{cm}^{-1}$  for C-O in the phenol bond after the  
180 adsorption of CPX. The peaks involved in the metal hydroxyl stretch that falls in the 1000-  
181 400  $\text{cm}^{-1}$  region of both spectra is assigned to the metal adsorption region (Velusamy *et al.*  
182 2021, Dai *et al.* 2020). By analyzing the relative intensities and positions of the spectra peaks,  
183 it is shown that there is a good interaction between CPX and the functional groups, which are  
184 parts of the biochar structure. A comparative analysis of the changes in the peak intensities  
185 associated with OH and C-H functional groups demonstrate enhanced interaction between the  
186 molecules and hydrogen abstraction in the course of adsorption (Karthik *et al.* 2020, Martins  
187 *et al.* 2015, Miao *et al.* 2016). The results provided by the spectra, showing the carbonyl and



188 phenol groups on the surface of the samples, as well as their absence after the adsorption  
189 process prove their importance in the process. The metal hydroxyl stretch being present all  
190 the time point out that the areas where the metal is adsorbed are in the process of being  
191 formed. The results of this study also point towards functional groups as significant for the  
192 adsorption of CPX by *Nephelium lappaceum* biochar.

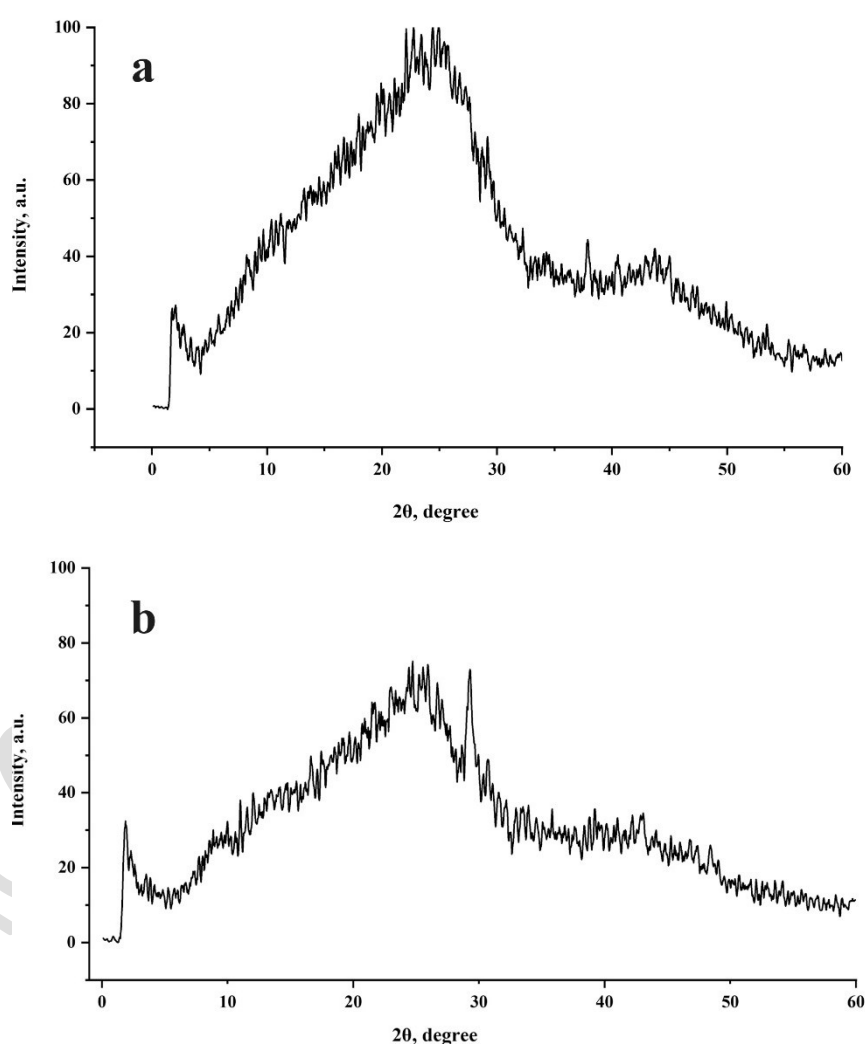


193  
194 **Figure 1.** FTIR graph for before adsorption (a) and after adsorption (b) of cephalixin using  
195 biochar.

### 196 3.1.2. XRD Analysis

197 XRD analysis was made before and after the adsorption of CPX by biochar (BC), as  
198 illustrated in Figure 2. The untreated CPX in BC was characterized by scattering angle found  
199 at  $2\theta = 24.42, 30.61, \text{ and } 40.09$ , which indicates that BC is an anhydrous crystal structure.

200 Similarly, the XRD pattern of CPX-treated BC displayed a broad peak at  $2\theta = 19.92, 24.89,$   
201 and  $43.89$ , which was reported to be crystalline in nature (Mondal *et al.* 2018, Topal *et al.*  
202 2020). The BC showed an increased d value (interlayer space) of  $4.453^\circ$  due to the  
203 interactions of the organic molecules in the BC, which were also confirmed by the FTIR  
204 results. When CPX was treated with BC, the d value was decreased to  $3.641^\circ$ . The surface-  
205 active sites of BC bind to the CPX functional group, resulting in a grafting reaction. A similar  
206 behavior was reported in (Ashiq *et al.* 2019).



207  
208  
209 **Figure 2.** XRD analysis of before adsorption (a) and after adsorption (b) for cephalexin using  
210 biochar.

211 **3.1.3. SEM –EDAX analysis**

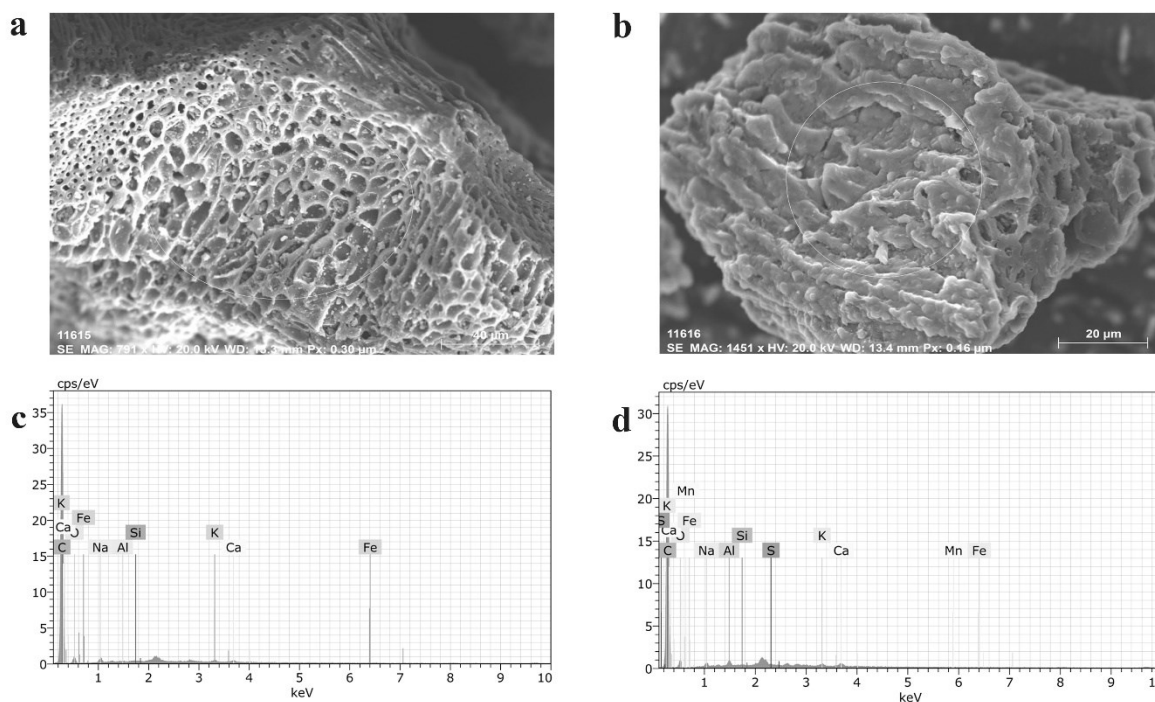
212 The SEM image before and after the adsorption of CPX by *Nephelium lappaceum* BC is  
213 shown in Figure 3a. The sample was examined using SEM to determine whether *Nephelium*  
214 *lappaceum* BC took up CPX. It is also employed to examine BC pore size, shape, and  
215 structure that favors cephalixin adsorption. Figure 3a shows that *Nephelium lappaceum* BC,  
216 before adsorption, was a non-uniform pore over the surface area. In contrast, the SEM image  
217 in Figure 3b represents the clump-like structure of cephalixin 250mg, effectively adhering to  
218 *Nephelium lappaceum* BC's pores. CPX is primarily adsorbed by *Nephelium lappaceum* BC  
219 because of its large pores, as shown in Figure 3b. The results obtained were compared with  
220 the SEM image of the reported soap nut seed biochar (Velusamy *et al.* 2021). The elemental  
221 composition before and after the adsorption of CPX by *Nephelium lappaceum* BC was  
222 analyzed using EDAX. The primary organic components of CPX are carbon and oxygen. The  
223 addition of CPX is implied by variations in the adsorbent's elemental contents, particularly  
224 carbon and oxygen weight percentages, before and after adsorption, as shown in Table 1 and  
225 Figures 3c and 3d. It was shown that the absence of sulphur and manganese in BC before  
226 adsorption. The peak of sulphur and manganese was observed in the EDAX spectra of CPX  
227 adsorption by *Nephelium lappaceum* BC, which confirmed the adsorption process. Oxygen,  
228 carbon, sodium, calcium, potassium and iron atoms are presented before and after adsorption.  
229 In addition, it was also found that the element C decreased by about 2% by weight, Na and  
230 Ca increased by 55% and 54% by weight. O was found to be increased by 2.5 weight % while  
231 other elements did not change much (Chakraborty *et al.* 2018, Topal *et al.* 2020).

232 **Table1.** Elemental composition of BC before and after CPX adsorption.

Map sum spectrum		
	Before adsorption	After adsorption
Components	Mass %	Mass %

C	88.18	86.19
O	10.67	10.94
Na	0.41	0.92
Ca	0.31	0.68
Al	0.04	0.57
K	0.24	0.45
S	-	0.10
Fe	0.11	0.07
Si	0.03	0.05
Mn	-	0.04

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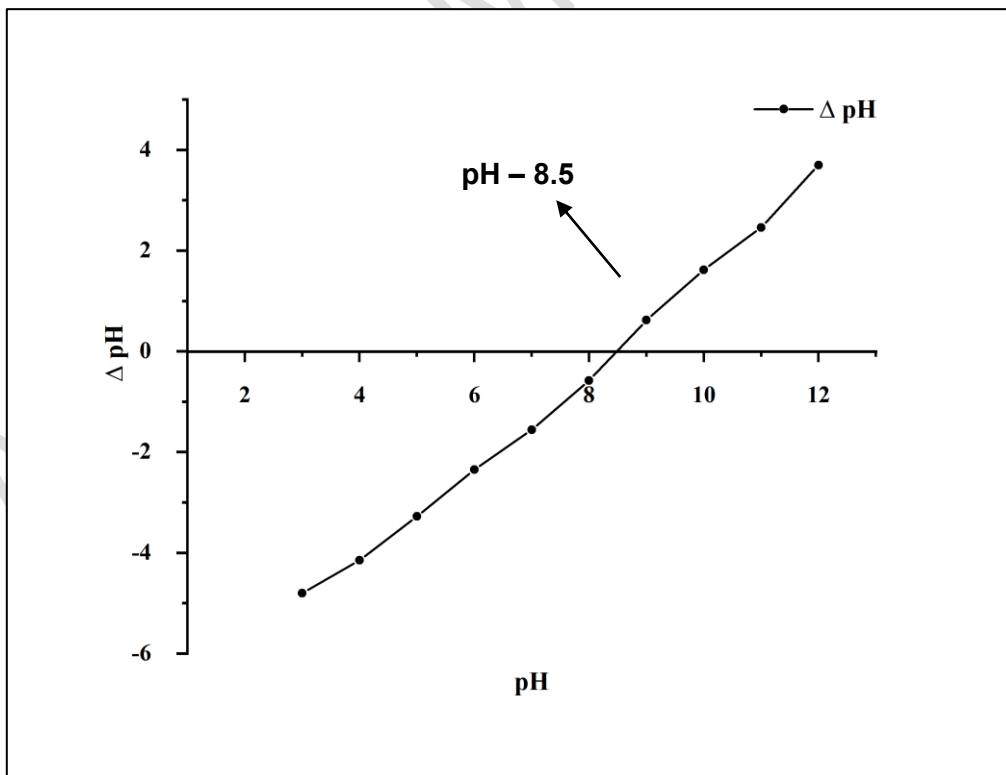
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235 **Figure 3.** SEM image (a) before adsorption, (b) after adsorption of CPX, Elemental analysis

236 (c) before and (d) after adsorption of CPX

237 **3.2. Point of Zero Charge**

238 The quality of the relationship between pH and antibiotic adsorption can be assessed using  
239 the Point of Zero Charge (pHpzc). Zero charge theory states that because there are less OH-  
240 ions on the surface of biochar with pH values below pzc, the surface is positively charged,  
241 and when pH values exceed pzc, the surface is negatively charged because there are more  
242 OH-ions. The surface charge of the biochar can be determined by performing pHpzc and it  
243 was reported as 8.5 which was shown in figure 4. Generally, the antibiotic Cephalexin (CPX)  
244 is in the zwitterionic form, which implies it has both positive and negative charges. In this  
245 regard, if pH less than pHpzc, the surface of biochar carries positive charge which may  
246 interact with negative charge of the antibiotic due to electrostatic interactions. Furthermore,  
247 increase in pH values higher than pHpzc the surface of biochar will become negatively charged  
248 does not encourage the electrostatic interactions due to higher concentration of OH<sup>-</sup> ions  
249 (Dehghan *et al.* 2019). Similar results were reported the electrostatic interactions of CPX and  
250 *Anthriscus sylvestris* derived activated biochar by performing the pHpzc (Shirani *et al.* 2020).



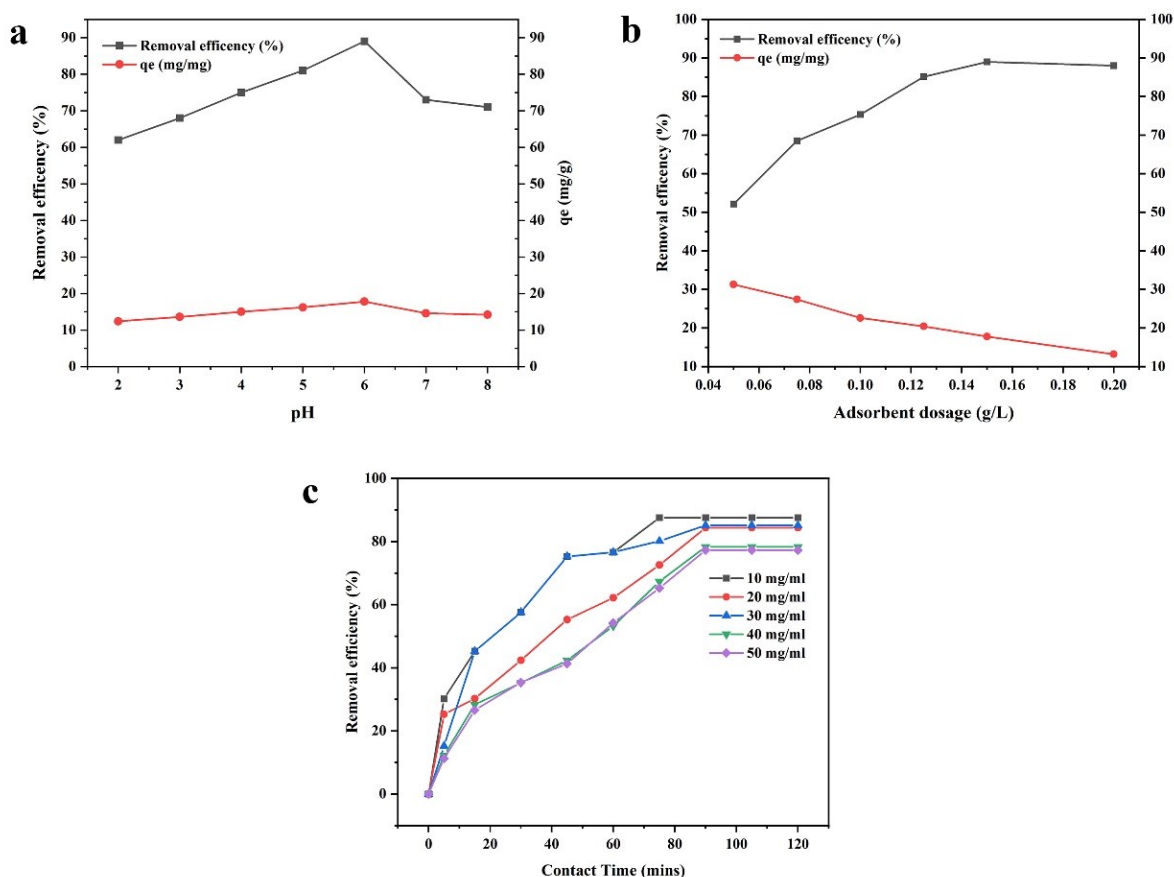
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252 **Figure 4.** Determination of pH pzc for *Nephelium lappaceum* seeds BC.

253 **3.3. Parameter Optimization**

254 **3.3.1. Influence of pH**

255 The mechanism by which the adsorbate and adsorbent interact is significantly influenced by  
256 pH. Figure 5a illustrates the removal of CPX with respect to pH and adsorption capacity. To  
257 investigate the elimination of CPX, the pH was altered from 2 to 8. CPX can be found in the  
258 form of zwitterion which implies the presence of positive and negative charges encourages  
259 the adsorption through electrostatic interactions. From the result of pH<sub>pzc</sub>, as the pH  
260 increases from 2-6, the surface charge of biochar interacts with the negatively charged  
261 antibiotic by electrostatic interactions which results in increased removal efficiency causes an  
262 increased adsorption efficiency. The maximum removal efficiency was reported in pH 6 with  
263 89% removal efficiency  $r_f$  and adsorption capacity  $q_e$  17.80 respectively. Further rise in pH,  
264 removal efficiency reduced. This may be due to higher concentration of OH<sup>-</sup> ions which  
265 results in lower adsorption efficiency. Similar findings were stated from the study of  
266 cephalexin removal by green synthesis of nanocomposites using pomegranate peel which  
267 relates the impact of pH and the adsorption process are due to the electrostatic interactions  
268 (Rashtbari *et al.* 2020, Priyadharsini *et al.* 2023).



269

270 **Figure 5.** Parameter optimization a) Influence of pH b) Influence of biochar dose with  
 271 respect to adsorption capacity and c) Influence of Contact time and Initial CPX Concentration  
 272 with respect to removal efficiency

### 273 3.3.2. Influence of biochar dose

274 The influence of adsorbent dose on removal efficiency and adsorption capacity was shown in  
 275 figure 5b. From the figure, it was noted that as adsorbent dosage increases with increase in  
 276 removal efficiency from 52.79% to 87.23% respectively. This increasing trend of removal  
 277 efficiency is due to the existence of more adsorption site as well as higher surface area  
 278 present in the biochar. Conversely, when the adsorbent dose was increased from 0.5 to 2 g/L,  
 279 the adsorption capacity declined. The adsorption capacity range was reduced from 31.6 to  
 280 17.37 mg/g. At higher dose of biochar accumulation reduces the availability of active surface  
 281 sites results in lower adsorption capacity. The optimal adsorbent dose was determined to be

282 1.25g/L, and when the adsorbent dosage increased, the removal efficiency and adsorption  
283 capacity remained constant. Similar trend was already reported in Miao *et al.* 2016.

### 284 **3.3.3. Effect of Contact time and Initial CPX Concentration**

285 In order to examine the impact of contact time and initial CPX concentration, the experiments  
286 were carried out by adding 1.25g/L of the biochar into flasks containing CPX initial  
287 concentration varying from 10 to 50 mg/L. Figure 5c showed the effect of contact time  
288 ranged from 10 to 120 mins. Initially the CPX adsorption was fast in the first 60 mins and  
289 decline slowly then finally reaches equilibrium. The equilibrium was reached at 90 mins.  
290 Further increase in contact time does not result in significant amount of rise in removal  
291 efficiency. Initially the adsorption was fast because there is the availability of more  
292 adsorption sites present in the adsorbent at the initial stage of adsorption (Miao *et al.*  
293 2016). Similar results were recorded by Rashtbari *et al.* 2020.

### 295 **3.4. Adsorption Kinetics**

296 The effect of contact time (0-120min) was investigated for CPX adsorption onto BC with  
297 varying initial CPX concentration (10 to 50 mg/L), adsorbate dosage -1.25g/L and pH -6.  
298 From figure 8 it was clear that initially increases rapidly in the adsorption rate and reaches  
299 equilibrium at 90 mins this is due to the higher concentration of CPX in solution and high  
300 availability of adsorption site in the BC surface. After it reaches equilibrium the adsorption  
301 rate decreased due to the lesser concentration of CPX as well as low availability of vacant  
302 sites (Rashtbari *et al.* 2020). The rate of CPX adsorption on BC was analyzed through kinetic  
303 studies: Pseudo-first order kinetics, Pseudo-second order kinetics, Elovich kinetics,  
304 Intraparticle diffusion kinetic model.

#### 305 **3.4.1. Pseudo-first order kinetics**



306 Pseudo –first order kinetics occurs when the concentration of one reactive molecule is kept  
307 constant or in excess. The equations used to determine the Pseudo-first order kinetics studies  
308 are mentioned in eq (6)

$$309 \ln(q_e - q_t) = \ln q_e - K_1 t \quad (6)$$

310 The equilibrium rate constants for pseudo-first (PFO) kinetics is  $K_1$  ( $\text{min}^{-1}$ ). Figure 6a  
311 represents the linear plot  $\ln(q_e - q_t)$  versus  $t$  at various concentration of CPX antibiotics  
312 indicating the intercept equal to  $K_1$  and  $\ln(q_e)$  as slope. The amounts of adsorbed CPX at  
313 equilibrium and time ‘ $t$ ’ are represented as  $q_e$  (mg/g) and  $q_t$  (mg/g). Table 2 represents the  
314 linear regression coefficient values and constants for pseudo first order kinetics.  $R^2$  values  
315 ranges from 0.802 – 0.969 which infers relatively low  $R^2$  values compared to other kinetics  
316 and the  $K_1$  values for various concentrations ranges from (0.019- 0.065). The adsorption  
317 kinetics data shows a deviation from straight line and implies that pseudo first order kinetics  
318 does not fit well for CPX adsorption with the biochar. The obtained findings were same with  
319 the  $\text{Cu}^{2+}$  adsorption with *Manikara zapota* sees biochar (Karthik *et al.* 2023).

#### 320 **3.4.2. Pseudo-second order kinetics**

321 The pseudo-second order model describes the chemical adsorption process which influence  
322 electrostatic forces and it is considered as the rate limiting step in adsorption process. The  
323 equations used to determine the Pseudo-second order kinetics studies are mentioned in eq (7)

$$324 \frac{1}{q_t} = \frac{1}{K_2 q_e^2} + \left(\frac{1}{q_e}\right) t \quad (7)$$

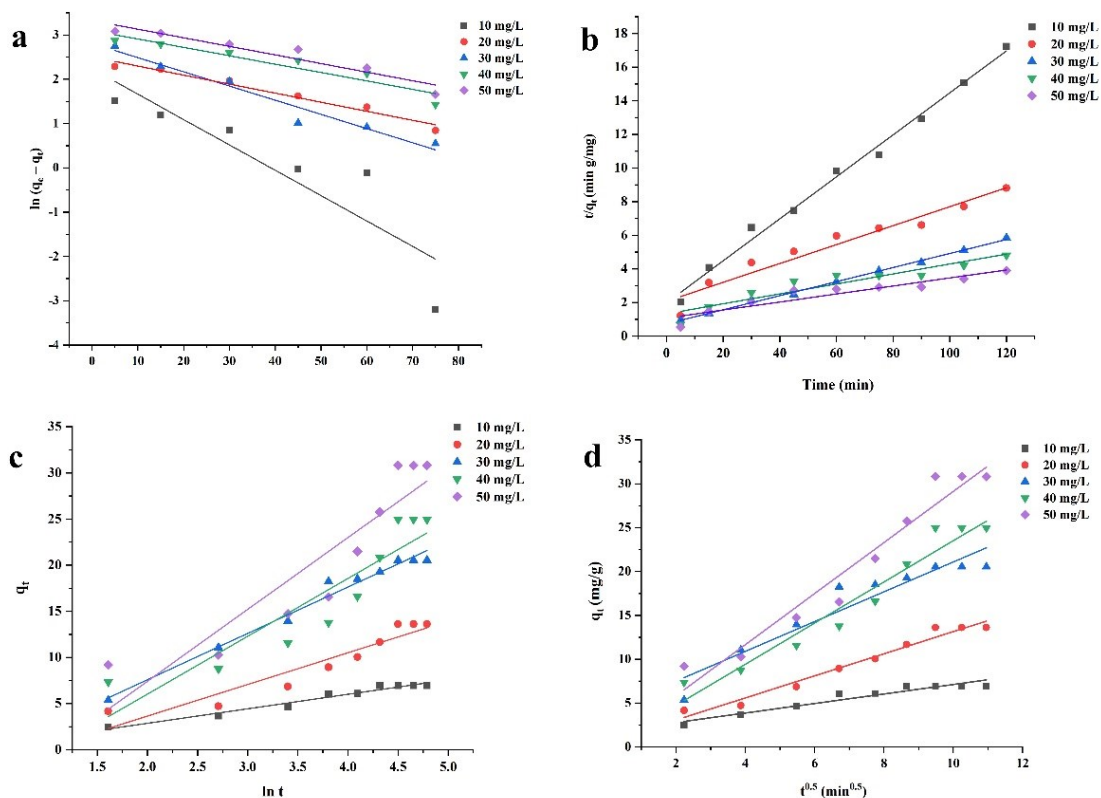
325 Equation (7) obeys Pseudo second order kinetics where  $K_2$  is the rate constant. Figure 6b  
326 plotted between  $(t/q_t)$  versus  $t$  at different concentration. Table 2 and figure 6b represents  
327 pseudo second order kinetics shows higher  $R^2$  value indicates better fit and the rate of  
328 adsorption was controlled by chemisorption. The findings presented here are supported by a  
329 chemisorption process that is compatible with the kinetic adsorption behavior reported in  
330 reference (Ma *et al.* 2020).

331 **3.4.3. Elovich Kinetics**

332 Elovich model suggests the surface is heterogeneous and supports the second order kinetics  
 333 (Zhang *et al.* 2021). The equations used to determine the Elovich Kinetics studies are  
 334 mentioned in eq (8)

$$335 \quad q_t = \frac{1}{\beta} \ln(\alpha\beta) + \frac{1}{\beta} \ln t \quad (8)$$

336 The adsorption capacity was calculated using equation (8), where  $\alpha$  represents the initial  
 337 adsorption rate and  $\beta$  denotes the desorption constant. From table 2 and figure 6c the values  
 338 of  $R^2$  ranges from (0.839-0.966) and the initial adsorption rate value was increased from  
 339 1.27-3.07 as the concentration increases from 10mg/L to 30 mg/L, indicates the second best  
 340 fit and it also confirms the process is chemisorption. Similar responses were reported in  
 341 cephalosporin antibiotic adsorption using soap nut biochar (Velusamy *et al.* 2021).



342 **Figure 6.** (a) Pseudo first order(b) Pseudo second order(c) Elovichand (d) Intraparticle  
 343 diffusion at varying contact time (min), pH 6.0, 1.25 g/L adsorbent dosage, Initial  
 344 concentration 10 – 50 mg/L.

345 **3.4.4. Intraparticle Diffusion**

346 Intraparticle diffusion is considered as rate limiting step in the adsorption process. The  
 347 adsorption capacity was calculated using the following eq (9)

348 
$$q_t = K_p t^{0.5} + C \tag{9}$$

349 The linear plot was plotted between  $q_t$  versus  $t^{0.5}$  and it is represented in the figure 6d where C  
 350 is the constant related to thickness of the boundary layer and  $K_p$  is the rate constant for  
 351 intraparticle diffusion. C and  $K_p$  values were computed from the graph using the intercept  
 352 and slope as parameters. If the C value increases, then the boundary layer effect will be more  
 353 whereas from the table 2 it was clear the intercept value is small indicating lower boundary  
 354 layer effect.

355 If adsorption is regulated by intra particle diffusion, the plot of  $q_t$  vs  $t^{0.5}$  should pass through  
 356 origin. From the plot it was concluded that the adsorption process is not dominated by intra  
 357 particle diffusion. Elimination of tetracycline by hazelnut shell biochar yielded similar results  
 358 (Fan *et al.* 2016).

359 **Table 2.** Kinetic parameter of PFO, PSO, Elovich and Intraparticle diffusion for the BC.

Initial Concen tration (mg/L)	PFO kinetics			PSO kinetics			Elovich kinetics			Intraparticle diffusion		
	$q_e$ mg/g	$k_1$ $\text{min}^{-1}$	$R^2$	$q_e$ mg/ g	$k_2$ $\text{min}^{-1}$	$R^2$	$\alpha$ mg/g min	$\beta$ g/m g	$R^2$	$k_p$ mg/g $\text{min}^0$	C mg/ g	$R^2$

10	9.35	0.057	0.801	8.00	0.007	0.99	1.27	0.65	0.966	0.54	1.69	0.927
20	12.20	0.020	0.969	17.76	0.002	0.94	1.34	0.29	0.894	1.24	2.54	0.970
30	17.11	0.033	0.951	23.92	0.0023	0.99	3.07	0.19	0.979	1.69	4.10	0.901
40	22.14	0.065	0.805	33.78	0.0006	0.875	2.21	0.15	0.839	2.35	0.03	0.947
50	27.72	0.019	0.914	41.84	0.0005	0.878	2.73	0.12	0.842	2.92	0.02	0.95

360

### 361 3.5 Adsorption Thermodynamics

362 The thermodynamic studies were conducted for CPX at varying temperatures (303.15,  
 363 313.15, 323.15 and 333.15 K) were analyzed by eq (10). The linear plot was plotted  $\ln K_L$   
 364 against  $1/T$  (Figure 7) and the thermodynamic parameters such as  $\Delta S^\circ$  and  $\Delta H^\circ$  were  
 365 determined from the slope and intercept values of the graph and it was summarized in table 3.

366 **Table 3.** Thermodynamic factors for CPX adsorption under various temperatures.

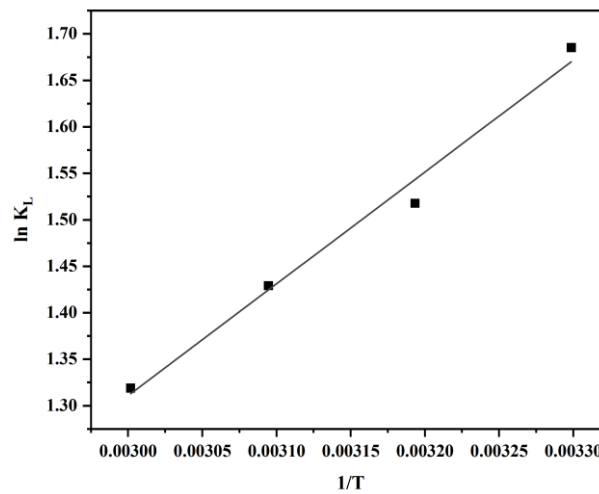
Temp (K)	$\Delta G^\circ$ (J/mol)	$-\Delta H^\circ$ (J/mol)	$-\Delta S^\circ =$ (J/mol)
303.15	-4247.551		
313.15	-3951.587		
323.15	-3839.413	9.996	19.091
333.15	-3653.311		

367

368  $\Delta G^\circ = -RT \ln K_L$  (10)

369 The values of Gibbs free energy changes from -4247.551, -3951.587, -3839.413 and -  
 370 3653.311 J/mol for CFX adsorption on BC at 303.15, 313.15, 323.15 and 333.15 K. The G  
 371 values were negative, revealing that the CPX adsorption mechanism was spontaneous in  
 372 nature. Increase in temperature results in increase in Gibbs free energy value implies the

373 possibility of adsorption at higher temperature. The  $\Delta H$  value for CPX adsorption was found  
374 to be  $-9.996$  J/mole. The result confirmed the nature of adsorption to be exothermic.  
375 Similarly,  $\Delta S$  value found to be  $-19.091$  J/mol which denotes the decrease in the degree of  
376 randomness at adsorbate and adsorbent interface in the process of adsorption (Ahmed et al.  
377 2012). Similar results were reported on cephalexin removal using *Albizia lebbek* seeds pods  
378 by microwave induced KOH and  $K_2CO_3$  activations.



379  
380 **Figure 7.** Thermodynamics graph between  $1/T$  versus  $\ln K_L$ .

### 381 **3.6. Adsorption Isotherm Models**

382 The technique by which CPX adsorbs on BC was investigated using adsorption isotherms.  
383 Dynamic phenomena of the adsorbate equilibrium distribution occur between an aqueous  
384 solution and an adsorbent. The isotherm models of Freundlich, Langmuir and Temkin  
385 described the process. To identify the better fit the parameters obtained from linear regression  
386 should be close to unity.

#### 387 **3.6.1 Freundlich Isotherm**

388 The non-equal binding sites are explained by the Freundlich model, which illustrates a  
389 multiple-layer physical adsorption process on a heterogeneous surface. The linear graph was  
390 plotted between  $\ln q_e$  versus  $\ln C_e$ . From the graph  $8b$   $K_f$ ,  $R^2$  and  $n$  values are calculated and it

391 was displayed in the table 4. From the table  $R^2$  value was found to be 0.993. The adsorption  
 392 of CPX by using *Nephelium lappaceum seeds* BC is well fitted with Freundlich isotherm. The  
 393 parameter  $n$  is also identified as the heterogeneity factor to assess whether the adsorption  
 394 process is physical ( $n > 1$ ), chemical ( $n < 1$ ), or linear ( $n = 1$ ). The  $n$  value demonstrates that  
 395 the process is physical and is found to be 1.35. The ratio  $1/n$  provides information on the  
 396 surface heterogeneity. Therefore, the closer  $1/n$  ratio approaches zero, the more heterogeneous  
 397 the surface material is in nature. The obtained  $1/n$  score 0.7396 indicates that the BC exhibits  
 398 substantial heterogeneity (Tang et al. 2018, Pezoti et al. 2014). Comparable outcomes were  
 399 noted when using coconut fiber biochar to remove Co (II) (Karthik et al. 2023).

### 400 **3.6.2 Langmuir Isotherm**

401 The theoretical maximum capacity for the adsorption of biochar, where there is no interaction  
 402 among adsorbates and when an adsorbate is adsorbed onto an adsorbent's homogeneous  
 403 surface, is identified using the Langmuir model, a single layer chemical absorption model  
 404 (Karthik et al. 2023). The linear graph was plotted against  $C_e/q_e$  versus  $C_e$ . From the graph 8a  
 405  $K_L$  and  $q_m$  values were calculated and tabulated in table 4. As indicated by the table, the  
 406 determination coefficient ( $R^2$ ) was discovered to be 0.9574. The separation factor, or  $R_L$ , is a  
 407 dimensionless metric that is utilized to identify whether the adsorption process is irreversible,  
 408 unversible, or favorable. The equation used to calculate  $R_L$  is represented in Equation  
 409 11. The Langmuir model fit into the adsorption process depends on adsorption capacity value  
 410 and the implications are  $R^2$  and  $K_L$  values.

411 **Table 4.** Langmuir, Freundlich and Temkin isotherm parameters for the adsorption of CPX  
 412 on BC.

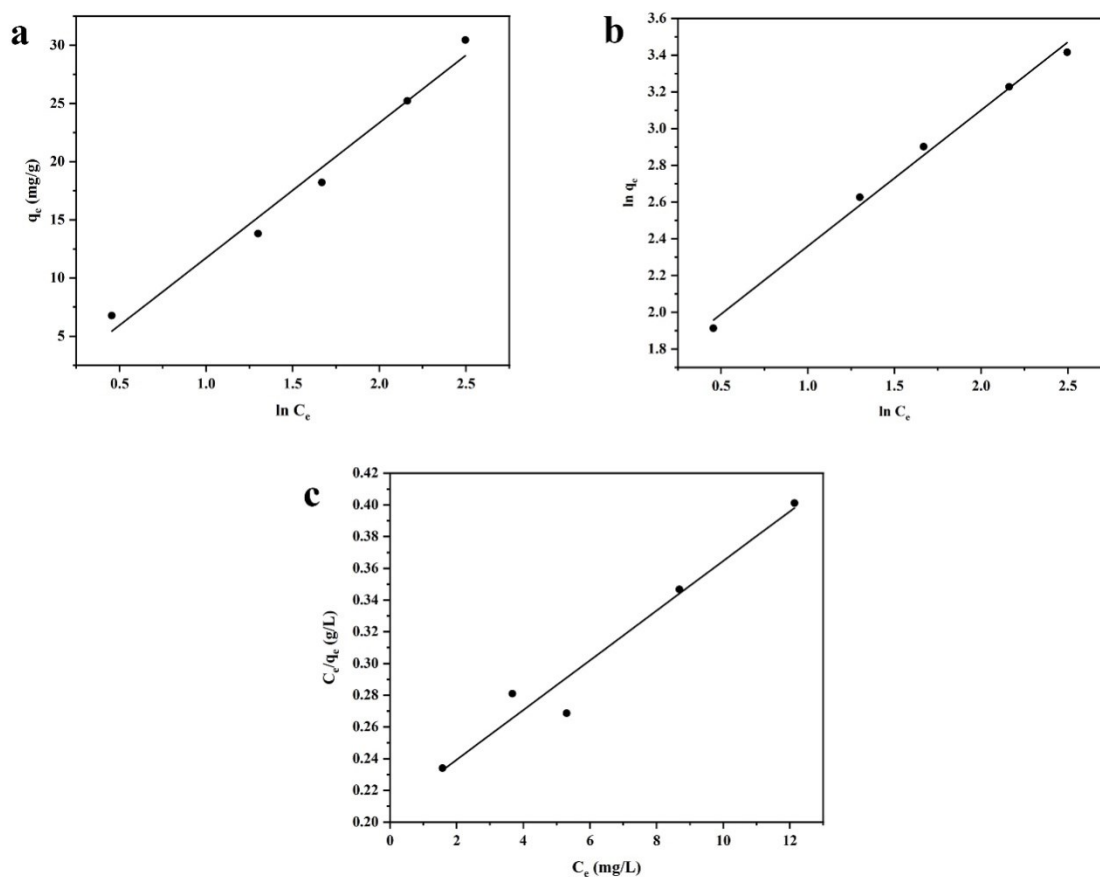
Isotherm models	Factors	$R^2$
Langmuir	$q_m = 63.694(\text{mg g}^{-1})$	0.957
	$K_L = 0.075(\text{mg}^{-1})$	

Freundlich	$K_f=5.060$ $((\text{mg} \cdot \text{g}^{-1} \cdot (1/\text{mg})^{1/n})$	0.993
	$n= 1.352$	
Temkin	$b_T=11.601$ $K_T=1.012(\text{mg}^{-1})$	0.979

413

414  $R_L = \frac{1}{K_L C_0}$  (11)

415  $R_L$  indicates whether biosorption is irreversible when ( $R_L = 0$ ), linear ( $R_L = 1$ ), feasible ( $0 <$   
416  $R_L < 1$ ), or uncertain ( $R_L > 1$ ). The separation factor value ( $R_L = 0.325$ ) suggests that the  
417 physical adsorption approach is preferable (Acelas *et al.* 2021). The Langmuir model fit into  
418 the adsorption process depends on adsorption capacity value and the implications are  $R^2$  and  
419  $K_L$  values. From the  $R^2$  values it was concluded that it does not fit well with Langmuir  
420 compared to Freundlich isotherm.



421 **Figure 8.** (a) Langmuir, (b) Freundlich and (c) Temkin isotherms at pH 6.0, 1.25g/L  
 422 adsorbent, initial concentration 10–50 mg/L.

423

424

### 425 3.6.3 Temkin Isotherm

426 The Temkin model was used to demonstrate the chemisorption process. The dispersion of  
 427 interaction energies during the adsorption process and the linear decline in adsorption energy  
 428 with surface coverage is used to illustrate the chemical adsorption process (Liu *et al.* 2017,  
 429 Sun *et al.* 2016). The linear plot was plotted between  $\ln C_e$  versus  $q_e$ . From the graph 8c  $R^2$   
 430 value,  $b_T$  and  $K_T$  values were evaluated and it was tabulated in table 4. As indicated by the  
 431 table, the determination coefficient ( $R^2$ ) was discovered to be 0.979. The adsorption is well



432 fitted with Temkin based on the regression coefficient. The adsorption energy variation is  
433 known as the Temkin constant, or  $b_T$ , and it determines whether an adsorption is exothermic  
434 when ( $b_T > 1$ ) or endothermic ( $b_T < 1$ ). The CPX adsorption using BC that arises  
435 exothermically in various concentrations was examined according to the  $b_T$  value of 11.601.  
436 Comparable outcomes were noted when using coconut fiber biochar to remove Co (II)  
437 (Karthik *et al.* 2023).

### 438 3.7 Potential Adsorption Capacity of Adsorbents

439 Table 5 illustrated the cephalexin adsorption capacities of the organic adsorbents. These  
440 methods have two drawbacks: low reactivity and a complicated preparatory process.  
441 Biochar has a distinct surface area and pore structure because of the existence of particular  
442 functional compounds like hydroxyl or carboxyl groups. The economic feasibility of  
443 employing BC in water treatment is also influenced by its chemical makeup and potential for  
444 regeneration and reuse. In contrast to these adsorbents, the overall carbon footprint of the BC  
445 production procedure may be calculated by considering the energy consumed during  
446 pyrolysis, the sustainability of the biomass supply, and the entire carbon footprint of the  
447 pyrolysis process. Because of its relatively easy production method and reasonably good  
448 adsorption capacity, *Nephelium lappaceum* seed BC could be an efficient adsorbent.

449

450

451 **Table 5.** Adsorption capacity of cephalexin using biochar

Adsorbent	Adsorption capacity mg/g	References
Oil palm fiber biochar	7.9	[Grisales-Cifuentes <i>et al.</i> 2021]

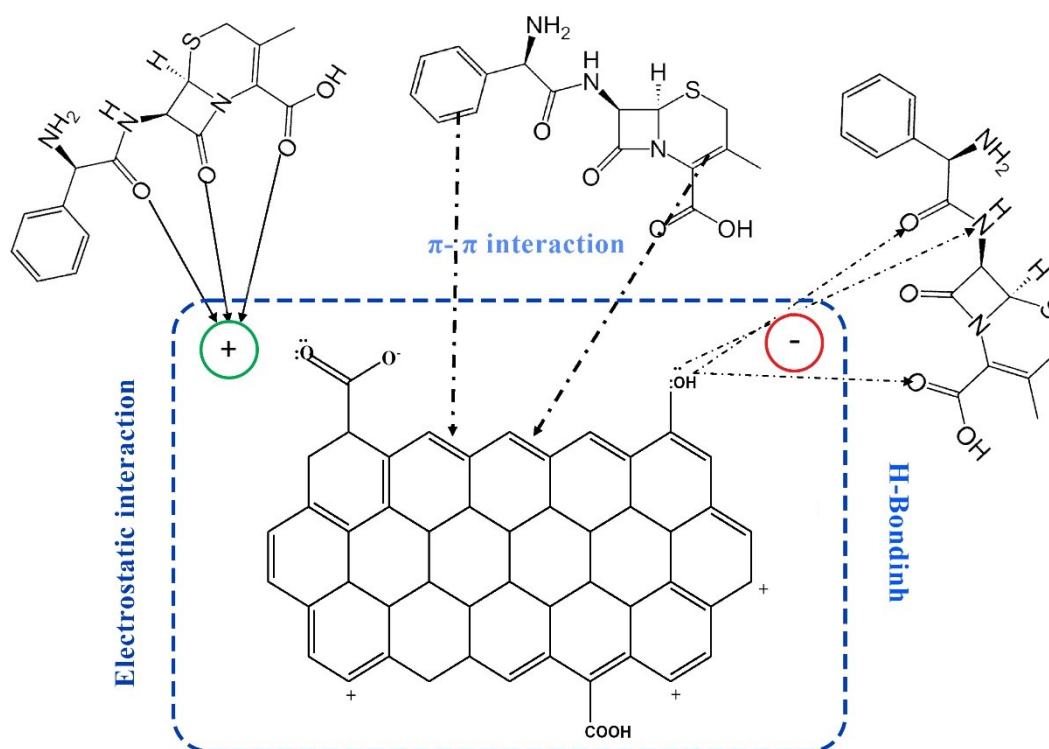
Natural zeolite	16.1	[Samarghandi <i>et al.</i> 2015]
Activated carbon	17.361	[Al-Khalisy <i>et al.</i> 2010]
Activated carbon from lotus	38.80	[Liu <i>et al.</i> 2011]
Alligator weed biochar	45.00	[Miao <i>et al.</i> 2016]
Activated carbon from pomegranate peel	48.78	[Rashtbari <i>et al.</i> 2020]
Palm oil fiber biochar	57.47	[Sun <i>et al.</i> 2016]
<i>Nephelium lappaceum</i> seed biochar	63.69	This study

452

### 453 **3.8. Adsorption Mechanism**

454 The primary mechanisms enabling CPX adsorption on BC are the hydrogen bond,  
455 hydrophobic, and electrostatic interactions depicted in Figure 9. The biochar surface's  
456 functional groups of the BC surface changed both before and after CPX adsorption,  
457 suggesting the participation of OH stretching, C-H stretching, and carbonyl C=O stretching.  
458 CPX's N-H functional groups can form hydrogen bonds with BC containing abundant  
459 oxygen, including C=O, -OH, and -COOH. The presence of these oxygen rich compounds in  
460 the BC was demonstrated by FTIR study. XRD analysis suggested changes in the crystal  
461 structure after adsorption. SEM-EDAX images confirmed the adsorption of cephalixin on the  
462 BC surface, with elemental analysis showing the presence of alumina and copper after  
463 adsorption. The point of zero charge (pHpzc) analysis indicated that the BC surface was  
464 positively charged at pH < 8.5 and negatively charged at pH > 8.5, making it effective for the  
465 electrostatic adsorption of CPX. Because of the electrostatic interaction, the charge on the  
466 surface of BC was induced by the pH of the sample interacting with the charged group in  
467 cephalixin. The aromatic ring portion of the CPX molecule can react with electrons in BC,

468 promoting  $\pi - \pi$  interaction and boosting adsorption. BC's surface has become more  
469 hydrophobic as oxygen-containing functional groups significantly influence the adsorption by  
470 providing specific sites for interactions, which results in the creation of hydrophobic  
471 interactions of BC that can attract the hydrophobic moieties in cephalixin to facilitate  
472 adsorption (Al Gheethi *et al.* 2021, Rashtbari *et al.* 2020).



473

474 **Figure 9.** Mechanism of possible interaction between biochar and cephalixin.

#### 475 4. CONCLUSIONS

476 The research evaluated using biochar produced from *Nephelium lappaceum* seeds to remove  
477 the widely used antibiotic cephalixin from an aqueous medium. FTIR, XRD, and SEM-  
478 EDAX were employed to evaluate the biochar before and after antibiotic adsorption. The  
479 impacts of different operating factors, namely pH, contact time, and cephalixin  
480 concentration, were examined with adsorption kinetics, isotherms, and thermodynamics. The  
481 maximum removal effectiveness was found at pH 6, with a temperature of 303.15 K, an  
482 adsorbent dose of 1.25 g/L, and an initial cephalixin concentration of 10-50 mg/L, with an

483 adsorption capacity of (63.69) mg/g. Based on kinetic investigations, the PSO model was  
484 more reliable with respect to the adsorption process, confirming a chemisorption mechanism.  
485 Thermodynamic analysis suggested that the adsorption process was feasible, spontaneous,  
486 and endothermic. Adsorption isotherm studies using Langmuir, Freundlich, and Temkin  
487 models exhibited good fits, with the Freundlich model having the highest  $R^2$  value. This  
488 study provides insightful information regarding the prospective use of fruit seed-derived  
489 biochar for antibiotic removal, which will improve the development of effective and  
490 sustainable water treatment methods. Further research could focus on optimizing the  
491 conditions of BC production and exploring its uses in real-world water treatment scenarios.  
492 After further suitable improvement, cephalexin and perhaps other antibiotics could be  
493 removed from wastewater using this adsorbent.

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