

Reviewing Biopolymers: Pioneering Sustainable Alternatives to Traditional Plastics Using Algae

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Graphical abstract



Abstract

In the wake of escalating environmental concerns conventional associated with plastics, exploring garnered sustainable alternatives has significant attention. Biopolymers, particularly those derived from algae, have emerged as promising substitutes due to their renewable nature and biodegradability. This review synthesizes current research endeavours aimed at harnessing algae-based biopolymers as sustainable alternatives to traditional plastics. The utilization of algae in biopolymer production capitalizes on their inherent capacity for rapid growth and efficient carbon fixation through photosynthesis. Algae offer diverse species, each possessing unique biochemical compositions conducive to biopolymer synthesis. This diversity enables the customization of biopolymer properties to suit various applications, ranging from packaging materials to biomedical implants. The synthesis pathways for algaebiopolymers, notably polyhydroxyalkanoates, based polyhydroxybutyrate, including are elucidated, highlighting the role of genetic engineering and process optimization in enhancing production yields. Furthermore, advancements in downstream processing techniques for extracting and purifying biopolymers from algae biomass are discussed, addressing challenges related to scalability and cost-effectiveness. Environmental sustainability considerations, such as life cycle assessments and end-oflife disposal options, are integral to evaluating algaebased biopolymers. Comparative analyses with conventional plastics underscore the environmental benefits of biopolymer adoption, including reduced greenhouse gas emissions and decreased dependence on finite fossil resources. Moreover, this review examines algae-based biopolymers' economic viability and market potential, exploring factors influencing commercialization, such as production costs, regulatory frameworks, and consumer acceptance. Algae-based biopolymers represent a promising avenue for pioneering sustainable alternatives to traditional plastics.

Keywords: Polyhydroxybutyrate, polyhydroxyalkanoates, Biopolymers, Algae-based, Sustainable alternatives, Environmental sustainability

1. Introduction

The global demand for plastic has risen, driven by plasticbased goods, exerting strain on the existing waste treatment infrastructure. Plastics typically consist of synthetic and semi-synthetic materials, mainly composed of polymers. Plastics include lightweight, cost-effective, easily manufacturable, flexible, and long-lasting characteristics, making them widely utilized by several individuals [Abdo and Ali 2019]. Plastics consist of artificial polymers, including polyesters, polyurethanes, halogenated plastics, and acrylics. Plastics primarily impact the living environment of humans, wildlife, and numerous marine species. Plastic deterioration has a direct impact on human life via the drinking of tap water and an indirect effect through the consumption of animals [Abu 2020]. Plastic trash is categorized into main and secondary forms. Primary plastics were retrieved from their initial location, and the deterioration of these primary plastics resulted in the formation of secondary plastics. Plastics are often categorized as micro, macro, and mega trash. The plastic particles range from 2 mm to 5 mm and are classified as microtrash. Mega debris refers to plastic particles with a dimension of 20 mm and are primarily utilized in food stores [Abu 2020].

Annually, about 8 million metric tonnes of plastic trash are disposed of in the ocean, with the potential for repurposing through the creation of inventive packaging materials [Alaerts *et al.* 2018]. In general, plastics may be

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biodegradable. Annually, around 1% of the world's bioplastic is manufactured. The starch blend that provides 21% of the total bioplastic is anticipated to grow in the market. Bioplastics are primarily used in packaging, textile, construction, and car industries [Alaerts *et al.* 2018]. Natural polymers, such as proteins and carbohydrates, were the primary source of bioplastic synthesis.

The manufacturing of bioplastics has had a parallel growth, reaching around 2 million tonnes in 2014 and 6.5 million tonnes in 2018. These bioplastics are primarily composed of polylactic acid-based polymers and starch. Recently, bioplastics have been derived from crops such as maize and potatoes, creating competition with the food business. Bioplastic manufacturing from crops requires extensive land area, fertilizers, and water resources [Amaro *et al.* 2014].

Furthermore, historically, the marketing of bio-based products encountered significant challenges on a widespread level [Barros et al. 2015]. Microalgae are regarded as a promising biomass resource for the development of bioplastics due to their lack of association with food sources. This organism exhibits the ability to thrive in aquatic environments and can generate substantial amounts of lipids. Moreover, bioplastic production using microalgae is considered more environmentally friendly and may be readily introduced and distributed within the economy and the bio-economy [Beckstrom 2016]. Bioplastics are applicable in the medicines, cosmetics, and food packaging sectors. Research efforts encompass the processing of microalgae bioplastic within a bio-refinery framework, the formulation of microalgae-polymer composites, and the genetic modification of microalgae strains to produce biopolymers [Biron 2016].

The wastewater from the industrial sector was treated under specific conditions (stress, pH, temperature, aeration duration, and agitation speed) to optimize the synthesis of PHB (polyhydroxybutyrate), presumably by microorganisms present in the wastewater. Hydroxy butyryl CoA, a precursor that undergoes polymerization to generate polyhydroxybutyrate (PHB), was utilized in this process. PHB exhibits insolubility in water and solubility in hydrolytic compounds [Brányiková et al. 2011]. It exhibits superior durability against UV light but displays reduced resilience to acidic and alkaline environments. It is nontoxic and readily dissolves in chloroform and other chlorinated hydrocarbons. The melting point of PHB is 175°C, and its glass transition temperature is 2°C. The tensile strength of PHB measures 40 MPa, which is comparable to that of polypropylene [Brányiková et al. 2011].

Additionally, PHB exhibits reduced stickiness when heated compared to other plastics. This study involved the synthesis of polyhydroxybutyrate from algae, which was then combined with natural polymers in varying ratios. The aim was to survey the resulting mix's physical, chemical, and mechanical characteristics and degradation time. This technology has the potential for several commercial applications, including agriculture, medicine, and the food business. However, it is crucial to implement efficient technology to improve industrialization, commercialization, and scaling-up processes. This study exhaustively scrutinizes several microalgae species used in bioplastic production to uncover any research deficiencies in this emerging field. The text explains opportunities associated with the expansion of microalgae. This review study comprehensively examines all facets of the technique and production.

2. Characteristics and attributes of poly-βhydroxybutyrates in terms of their chemical and physical qualities

The synthesis of Poly-β-hydroxybutyrates begins with the precursor molecule acetyl coenzyme A (acetyl-CoA). This process involves three consecutive enzymatic reactions. The first step is the conversion of 2-acetyl CoA to 1-acetyl CoA, which is catalyzed by the enzyme 3-ketothiolase. The second step is the production of D-3-hydroxybuturylCoA, achieved through converting NADPH-dependent acetoacetyl-CoA reductase to acetoacetyl CoA [Brennan and Owende 2010]. The D-3-hydroxy butyryl moiety was connected to the existing PHB molecule by an ester bond facilitated by the last enzyme, PHB synthases. Therefore, substituents or unsaturation were detected in the fatty acids at positions 4, 5, and 6 of the pendant group, where a hydroxyl group is located. Simultaneously, PHB and poly three hydroxy valerates (PHV) were categorized as shortchain length PHAs and merged to constitute a PHA class [Bussa et al. 2010]. On the other hand, it has been reported that the typical chain length of PHAs ranges from carbon six to carbon 16, explicitly referring to 3-hydroxy fatty acids. Additionally, it has been proposed that the PHB homopolymer synthesized by bacteria consists of more than one molecule and contains 3% hydroxy valerate monomers [Carlozzi et al. 2003].

The combination of Valerate and glucose served as the substrate for the microorganisms. These microorganisms play a crucial role in synthesizing small-chain-linked PHAs [Chandra and Mohan 2014]. The formation of PHBHx copolymers, consisting of 3-hydroxyl hexanoate units in combination with PHAs copolymers of PHB, was described [Charlie et al. 2019]. Random copolymers were formed by alternatingly introducing different substrates into the current culture. This process led to the synthesis of a PHA block by bacteria. The molecular mass of PHBs generated by bacteria is typically 4.6×106 Da, with a poly disparity (Mw/Mn) of around 2.0 [Chen et al. 2011]. The biomaterial polypropylene has comparable material properties [Chisti 2008, 2007]. The PHB homopolymer [Ciapponi et al. 2019] exhibited a high degree of crystallinity. This material is rigid and brittle, yet it possesses elastic capabilities. When twisted into fibers, it was seen to have an elastic behavior [Costa et al. 2019]. The helical crystalline structure is a characteristic of homopolymers. This copolymer, characterized by its structural composition, exhibits superior durability against ultraviolet light but reduces resilience to acidic and

alkaline environments. It shares structural similarities with many other copolymers [Das *et al.* 2018]. A recent study analyzed PHB's melting behavior and crystallization [Di Caprio *et al.* 2016] and examined its physical properties.

Investigate trends in the generation of Poly-βhydroxybutyrates.

Microalgae gained attention for research following the discovery of Spirulina in Spain before 1519 [Dianursanti et al. 2019]. Scientists have extensively studied the nutritional properties of microalgae and made significant discoveries, such as the PHB homopolymer, which holds great potential for various applications benefiting humankind [Dianursanti et al. 2018]. Microalgae have been employed for bioremediation in wastewater treatment for an extended period. Using wastewater to culture microorganisms has become a leading practice in resource recovery-the authors of this publication aimed to highlight twenty years of trends in PHB research. A comprehensive evaluation was conducted on indexed publications published between 1999 and 2020, focusing on the search for microbial plastics and biosynthesis. The keywords used for the search were PHB, microbial bioplastics, and biosynthesis [DiGregorio 2009]. This assessment provides a comprehensive analysis of the research and development activities from a chronological standpoint. The literature research indicated that "PHBs" was the most often utilized keyword during twenty years, accounting for 50% of the occurrences. The process of synthesizing polyhydroxybutyrate (PHB) is seen in Figure 1.



Figure 1. Biosynthesis of PHB

While algae are well-known for their ability to produce polyhydroxybutyrate (PHB), other microorganisms, including certain types of bacteria, can also synthesize PHB. Activated sludge, a mixture of bacteria, fungi, protozoa, and other microorganisms used in wastewater treatment, can contain bacteria capable of producing PHB. Several types of bacteria, such as species of Bacillus, Pseudomonas, and Alcaligenes, are known to produce PHB as intracellular storage granules when they are under conditions of nutrient limitation, mainly when there is an excess of carbon source available. This ability makes them useful in various biotechnological applications, including producing biodegradable plastics like PHB [Dianursanti 2018; DiGregorio 2009].

4. Biological production of polyhydroxybutyrate

Multiple studies were conducted in biosynthesis to generate polyhydroxybutyrate the via microbial fermentative method [Dixon et al. 2018]. The production of polyhydroxybutyrate involves three fundamental steps catalyzed by three different enzymes. Firstly, the enzyme B-keto thiolase catalyzes the condensation of two acetyl-CoA molecules to form acetoacetyl-CoA. Secondly, the encoded with NADPH-dependent enzyme phaB, acetoacetyl CoA dehydrogenase, reduces acetoacetyl-CoA. Lastly, the enzyme PHA synthase, attached with phaC, catalyzes the polymerization of 3-hydroxy acid (3HA) units, specifically (R)-3-hydroxy butyryl-CoA. The result is the transformed acetyl-CoA used in the PHB biosynthetic pathway.

4.1. Microalgae and additives

The Algae species are classified as microalgae and macroalgae, with one million species identified [Draaisma]. Microalgae, microscopic organisms, utilize solar energy to make adenosine triphosphate (ATP), a compound that may be found in fresh waters and coastal locations [Eagan et al. 2017]. Microalgae are utilized as a substitute for several sources in biofuel manufacture. Pharmaceutical formulations extensively use it, while cosmetics might be a supplementary product in the food business [European Bioplastics]. The development of the bio-economy using microalgae has shown constant improvement [Berlin European Bioplastics 2018]. Microalgae consist primarily of proteins (6-52%), lipids (7-23%), and carbohydrates (5-23%) [Fabra et al. 2018]. In addition, microalgae have a composition that includes iron (1395-11,101 mg·kg⁻¹), phosphorous (0.7-1.5%), manganese (45–454 mg·kg⁻¹), copper (18–102 mg·kg⁻¹), magnesium (0.3–0.7%), selenium (0–0.5 mg \cdot kg⁻¹), calcium (0.1-3.0%), sulphur (0.4-1.4%), potassium (0.7-2.4%), sodium (0.8–2.7%), and zinc (28–64mg·kg⁻¹) [Dianursanti et al. 2019]. The subsequent sections will provide a detailed analysis of bioplastic manufacture, focusing on using additives, microalgae species, and chemicals [Fabra et al. 2018].

4.2. Chlorella

Chlorella is a genus of photosynthetic microorganisms that belong to the green algae family. It predominantly inhabits aquatic environments and primarily comprises around 58% protein. Its fracture resistance is highly effective because of its clustered cell walls and exceptional thermal stability compared to Spirulina [Fu et al. 2012]. These species are primarily used in mixes of biomass and polymers. The comparison between blends consisting mainly of polymers/additives and microalgae biomass (100%) for bioplastic manufacture demonstrated the necessity of efficient combining for commercial use [Gifuni et al. 2017]. The test findings showed that using chlorella vulgaris yielded superior bioplastics to Spirulina. Nevertheless, Spirulina has superior mixing characteristics compared to Chlorella [Gilbert 2016]. The study suggested that a high-quality product can be produced using a compatibilizer (maleic anhydride) at a concentration of 6% in the proposed mixture composition [Gozan 2018].

Starch, a notable biopolymer, is primarily used in the chemical, bioplastic, and food production industries. The gelatinization temperature of this substance is estimated to be around 110° C. This temperature is suitable for the production of bioplastics, mostly made from starch [Hai *et al.* 2000]. Combining *Chlorella* cells with hydrogen ions creates a homogeneous mixture without empty spaces [Harun *et al.* 2009]. Prior use of ultra-sonic homogenization as a pretreatment step before combining

significantly enhanced *Chlorella*-PVA blends' uniformity and surface characteristics. This technique was suggested as a potential substitute for food packaging [Hempel *et al.* 2011]. The researchers examined the disparity between *Chlorella*-PE-based composites with and without modifications of PE using maleic anhydride. The tensile strength of composites is enhanced by introducing PE Modification [Huo *et al.* 2011].

SL.NO	Algae name	% Of PHB & PHA production	Extraction method	Studies carried	References
1	Chlorella pyrenoidosa	27%	Fogg's media	Analysis of PHB content	[Murilo Moraes Mourão n.d.]
2	Chlorella fusca	17.4%	D-xylose	Characterization and quantification of polyhydroxybutyrate	[Bhati and Mallick 2014]
3	Spirulina platensis	6.20%	Zarrouk medium	PHB Production and comparision	[Clemens Trosch 2018]
4	Sargassum sp.	5.36%	Culture Mediaseaweed	Characterization of PHB (Fourier transform infrared (FTIR)), Differential scanning calorimetry, Nuclear magnetic resonance.	[Monshupanee 2016]
5	High-rate algal pond	17%	Different media	Characterization (SEM analysis, Mechanical properties)	[Monshupanee 2016]
6	Chroococcus	12%		Characterization (SEM analysis, Mechanical properties)	[Monshupanee 2016]
7	Haematococcus	35%		Characterization (SEM analysis, Mechanical properties)	[Monshupanee 2016]
8	Microcystis	45%		Characterization (SEM analysis, Mechanical properties)	[Monshupanee 2016]
9	Chlorella vulgaris		Sudan black B and Nile Blue Stain	Optimization	[Ranjana Bhati 2010]
10	Stigeoclonium sp.		Nile Red	Optimization	[Martins <i>et al.</i> 2014]
11	Nostoc muscorum	71%		Optimization	[Gopi <i>et al.</i> 2014]
12	cyanobacterium Synechocystis sp.	12.5%	Culture medium	Analysis and characterization of PHB	[Quines <i>et al.</i> 20015]
13	Chlorogloea fritschii	51%	heterotrophy	Material properties of PHB	[Riedel n.d.]
14	Phaeodactylum tricornutum	10.6%	Cell culture	Fluorescence and electron microscopy	[Riedel n.d.]
15	Anabaena cylindrica	2.8%	Yellore and Desia		[Knuckey <i>et al.</i> 2006]
16	Anabaena doliolum	3.5%	Yellore and Desia		[Knuckey <i>et al.</i> 2006]
17	Anabaena variabilis	3.1%	Yellore and Desia		[Knuckey <i>et al.</i> 2006]
18	Anacystis nidulans	4.4%	Yellore and Desia		[Knuckey <i>et al.</i> 2006]
19	Aphanocapsa sp.,	3.3%	Yellore and Desia		[Knuckey <i>et al.</i> 2006]
20	Aulosira fertilissima	6.5%	Yellore and Desia		[Knuckey <i>et al.</i> 2006]
21	Calothrix sp.	6.8%	Yellore and Desia		[Knuckey <i>et al.</i> 2006]

22	Fischerella muscicola	2 4%	Yellore and		[Knuckey et al.
		2.170	Desia		2006]
23	Microcystis aeruginosa	4 0%	Yellore and		[Knuckey <i>et al.</i>
		4.070	Desia		2006]
24	Microcystis sp.	3.6%	Yellore and		[Knuckey <i>et al.</i>
		5.0%	Desia		2006]
25	Nostoc linckia	3.6%	Yellore and		[Knuckey et al.
			Desia		2006]
26	Nostoc muscorumc	8.5%	Yellore and	Quantification DHR	[Knuckey et al.
			Desia	Qualitification PHB	2006]
27	Oscillatoria limosa	2.9%	Yellore and		[Knuckey et al.
			Desia		2006]
20	Pleurocapsa sp	2.9%	Yellore and		[Knuckey <i>et al.</i>
28			Desia		2006]
20	Disclassing	2 70/	Yellore and		[Knuckey et al.
29	Rivularia sp	2.7%	Desia		2006]
20	Scytonema sp.		Yellore and		[Knuckey et al.
30		7.4%	Desia		2006]
24	Spirulina platensis	2.5%	Yellore and		[Knuckey et al.
31		2.5%	Desia		2006]
	Synechocystis sp. PCC 6803		Yellore and		[Knuckey et al.
32		4.3%	Desia		2006]
	Tolypothrix sp	2.2%	Yellore and		[Knuckey et al.
33			Desia		2006]
	Westiellopsis	2.9%	Yellore and		[Knuckey et al.
34	prolifica		Desia		2006]
	Synechocystis			Analytical methods, Theoretical framework	[Knuckey et al.
35	PCC6803				2006]
36	Spirulina sp. LEB 18	44.2%		Sodium hypochlorite	[Kutz 2011]
37	Cyanobium sp	2.9%		10–12% active chlorine, Sodium hypochlorite	[Kutz 2011]
38	Nostoc ellipsosporum	19.2%		10–12% active chlorine, Sodium hypochlorite	[Kutz 2011]
	Synechococcus				
39	, nidulans	10.2%		10–12% active chlorine, Sodium hypochlorite	[Kutz 2011]
40	Phormidium sp	7.6%		Chloroform -Soxhlet method (hot extraction)	[Lee2001]
41	Svnechococcus sp.	4.5%		Chloroform -Soxhlet method (hot extraction)	[Lee2001]
42	Svnechocvstis sp	3.7%		Chloroform -Soxhlet method (hot extraction)	[Lee2001]
43	Anahaena sp	2.3%		Chloroform -Soxhlet method (hot extraction)	[lee2001]
-15	Ralstonia eutropha	25 – 99.0%		Propylene carbonate mainly with thermally	[] upatini et al
44				treated biomass	2017]
	Ralstonia eutropha	84%			
				Methyl isobutyl ketone mainly for cell	[MacArthur
45				disruption & ethyl acetate for recovery	2017]
					-

4.3. Spirulina

Spirulina can adapt to severe settings and has been used as a critical source of protein in the food industry for many years [Johnsson and Steuer 2018]. Spirulina platensis has a higher protein concentration [Kalia and Avérous 2016]. Several experiments were conducted to assess the potential of Spirulina for manufacturing bio-plastics. The comparable cell size of Spirulina and Chlorella makes them more desirable for creating bio-plastic blends [Kaparapu Although Spirulina and Chlorella 2018]. share commonalities, they exhibit different behaviours and bioplastic capabilities, mainly when mixed with PE, which can be attributed to their differing amino acid compositions. The use of compatibilizers [Kato 2019] can improve the characteristics of *Chlorella*-based bioplastic. The assimilation rate of the compatibilizer into *Spirulina* was approximately 6% by weight—a mixture of *platensis* with PVA.

Consequently, a bioplastic sheet with superior tensile strength was produced compared to the commercially available plastic bag. Using a compatibilizer enhances the plastic's potential, particularly in terms of elongation, and helps the formation of delicate layers [Khanra *et al.* 2018]. Nevertheless, the capacity for reinforcement was diminished when the particle size of microalgae biomass exceeded five μ m [Sruthy 2016]. The smaller particles were efficiently mixed with various materials [Cassuriaga 2018]. Glycerol, with percentages ranging from 15% to 30%, is primarily used to enhance the flexibility of the plastic. It is predominantly derived from *S. Platensis*, a

term. In bioplastic production, including glycerol at a concentration of 30% resulted in reduced elongation and increased tensile strength compared to conventional plastic bags [Uma Maheswari 2011]. This finding provides evidence that bioplastic may be extensively employed in the food sector for packaging, cosmetic, and medicinal purposes, particularly in cases where a significant amount of elongation is unnecessary [Nahid Azizi n.d.]. The plasticization of *Spirulina* yields two primary outcomes: enhanced inter-surface adhesion and uniform phase distribution. They are utilizing mixing processes that improve both the uniformity and tensile strength.

However, adding a compatibilizer [Sayeda M. Abdo 2019] does not enhance the mechanical characteristics of the bioplastic. Adding maleic anhydride-grafted PBS to the biomass as a primary compatibilizer improves the product's strength, particularly in tensile strength, and decreases the temperature, leading to deterioration [Rebecca Robert 2018]. The table provided, labeled Table 1, illustrates the process of characterizing and extracting PHB using several species of algae.

5. Alternative Microalgae Species Employed for Bioplastic Manufacturing

As mentioned earlier, most of the research in this emerging subject focused on studying bio-plastic production using *Spirulina* and *Chlorella*. Regarding these efforts, several research have examined the efficacy of several microalgae species [Martin Chaplin n.d.].

5.1 Hybrid Biological System

The heterotrophic culture of PHA has successfully met its growth objective, but the expense associated with acquiring feed remains a considerable concern [Mata et al. 2010]. Open pond culture often yields economic advantages, while ensuring a contamination-free monoculture requires constant monitoring and maintenance, which can be challenging. Likewise, the unpredictable variations in candlepower, pH, temperature, and carbonic acid gas levels caused by daily and cyclical swings lead to limited economic structures [McBride 2014]. Therefore, creating a fusion organic structure presents difficulties in utilizing event capability in both heterotrophic and photoautotrophic culture requirements [Medeiros et al. 2015]. Researchers have discovered that the claims on the potential of PHA production by photoautotrophic scientific agriculture are limited and can only be attained up to 55% in Synechococcus sp the number 68. Therefore, the scientific agricultural structures and procedures that were suggested to improve PHA production are as follows [Mohan 2019].

5.2 Mixotrophy

Mixotrophic or heterotrophic agronomy has consistently led to significant improvements in PHA earnings [Monshupanee *et al.* 2016]. However, there is still potential for a 2 to 9-fold increase in PHA output. This concept aimed to incorporate heterotrophic (tilling) and photoautotrophic elements inside the same structure for practice. The proposed design includes a live bacterium that functions as a PHA producer. This bacterium undergoes photoautotrophic cultivation during the day, using light, and then switches to heterotrophic cultivation at night. [Moreno-Garcia 2017] Bacteria may utilize carbon dioxide in sunlight as a carbon source. On the other hand, in the dark portion, an external supply of carbon (such as glucose, acetate, polymer precursors, etc.) is also introduced. An alternative improvement method involves enhancing biomass growth and reducing PHA production in the existing bacteria by cultivating photoautotrophic organisms. In this configuration, the bacteria are exposed to photoautotrophic conditions during the daytime without any restrictions on nitrogen/phosphorus (N/P) levels, resulting in significant biomass accumulation until the late-night exponential or dormant phase. During the dark phase, the bacteria can be in a heterotrophic state, but their growth is limited by N/P availability. It suggests the presence of a distinct segment within the structure, characterized by a notable difference in intensity, with one part appearing more prominent or darker and the other appearing more limited or brighter. This structural contrast is believed to promote the production of polyhydroxyalkanoates (PHA) in bacteria [Musa et al. 2019].

6. Association between organisms that are capable of photo autotrophy and heterotrophy.

The main goal of establishing an association was to create an additional economic framework that would provide greater access to resources, higher productivity, costeffective food production, social stability, non-competitive segregation, and the distribution of carbon or energy resources among members of a community-based metabolic system [Oh et al. 2018]. The reciprocal relationship refers to a mutual interaction or influence between different elements or factors. In biotechnology advancements, this reciprocal relationship has been observed as the primary driving force behind numerous developments in areas such as bioprocessing, biofuel production, and the creation of various other biotechnological products. Essentially, these advancements in one area often contribute to and benefit from progress in other related fields, leading to a symbiotic relationship that fuels innovation in biotechnology [Oh et al. 2018]. The photosynthetic association consists of an alga and a bacterium that consumes it. This association produces PHA, which makes up 20-30% of the cell dry weight without needing other elements. The method of metabolic network modelling is illustrated in Figure 2.

Impact of extraction techniques on the quantity and quality of Poly-β-hydroxybutyrate

Microorganisms that produce PHB can store as much as 90% of their mass in biopolymers [Osanai *et al.* 2013]. However, it is challenging to remove the polymer from inside the cell. According to reports, harvesting and extraction activities make up around 60-80% of the total production expenses [Otsuki *et al.* 2004]. Enhancements

in extraction techniques are required to achieve a significant recovery yield while ensuring the desired polymer properties [Pal et al. 2019]. Chemical, surfactantchelate, enzyme, and solvent extraction are commonly used techniques for extracting PHB. Over the years, several advanced methods for cell disruption and solvent procedures have been developed and employed to effectively release the desired product by lysing cells [Pandey et al. 2017]. The solvent extraction process usually entails the slow dissolution of microbial biomass in a combination of several solvents, which is then followed by the precipitation of polymers [Park and Seo 2011]. The robustness of the cell wall is crucial for disrupting microbial cells throughout the process of biopolymer recovery. Alterations in the structure of the microbial cell wall can be induced by modifying the content of the growth medium [Pragya et al. 2013]. Different circumstances, such as action duration, solvent type, concentration, and temperature range, impact a biopolymer's extraction rate, molecular weight, and purity [Proshad et al. 2018]. These parameters influence biopolymers' cost, characteristics, and monomeric composition, which affect their industrial applications [Pulz 2001].



Figure 2. Metabolic network modelling mechanism

Unlike the usage of plastic bags, which are intended for single use, the medical industry requires a polymer that is devoid of toxins and has a high level of purity [Quinn and Davis 2015]. The monomeric composition and molecular mass of biopolymers may be significantly affected by extraction methods, as demonstrated by research [Rahman et al. 2015]. This suggests that the choice of extraction technique is crucial for achieving the required properties and functionality of the polymers in industrial applications. PHB derived from the combination of heated chloroform, diethyl ether, and sodium hypochlorite was utilized for cancer detection [Ranade 2009]. Compared to normal epithelial cells, breast cancer cells (T47D) demonstrated much stronger adherence to the PHB sheets [Richmond et al. 1990]. PHA polymers, namely PHB, have been employed in treating wounds, bone regeneration, surgical instruments, and drug delivery owing to their compatibility with mammalian cells [Rudnik 2019].

Additionally, it has widespread popularity within the agriculture sector [Sabathini *et al.* 2018]. While the result

may be acceptable, it is crucial to consider the adverse ecological and financial repercussions of using highly concentrated chemicals in the extraction process [Salim et al. 2011]. Hydrothermal conversion and biological extraction, which are eco-friendly technologies, are being used because of the limitations associated with the mentioned approaches, such as high cost, time consumption, and toxicity. Predation systems, mealworm digesting systems, and Bacteriophage-mediated lysis systems are all instances of bio-extraction technologies [Sato et al. 2006]. These strategies provide economic benefits and minimize negative impacts on environmental and human well-being [Scott et al. 2010]. The PHB was obtained from Enterobacter aerogenes cultivated in sewage water using hypochlorite, yielding 96.25%. The biomass derived from PHB, employed in wastewater treatment (WWT), was efficiently transformed into propylene using hydrothermal conversion. This conversion process examined solvents, including phenetole, anisole, and cyclohexanone, often used in industrial settings for extended periods [Sereni 2016]. Additionally, the extraction of Spirulina LEB 18 was successfully carried out in wastewater treatment. To decrease the expenses associated with collecting microalgae, it is imperative to consider using environmentally benign and reusable harvesting techniques, such as crystalline nanocellulose [Lopez-Exposito et al. 2019; Shen et al. 2009].

8. Utilization of Microalgae Biomass in Material Blending

Conventional polymers are commonly employed to create composite bioplastics derived from microalgae sources to improve plastic properties. The most often employed polymers in the combining process are polyethylene (PE) and polypropylene (PP). They account for more than twothirds of the worldwide demand for plastic. PE is utilized in several industries, such as medical items, food packaging, cosmetics, and prostheses. UHMW-PE is a high molecular weight polyethylene with a molecular weight ranging from 2 to 6 million. Bioplastic manufacturers favour this material due to its absence of toxins, odour, and low moisture absorption capacity. Wang conducted a trial where he mixed UHMW-PE with Spirulina at different ratios ranging from 20% to 80% with intervals of 15%. Using a PE-Spirulina-EG (Ethylene Glycol) ratio of 80:13:7 led to an increase in tensile strength, as reported in reference [Singh et al. 2017].

Polypropylene (PP) is particularly suitable for packaging yogurt, medicinal products, and beverages because of its somewhat translucent appearance and its resistance to the impact of heat and mechanical responses. A study was conducted to produce films formed of bioplastic containing *Chlorella* at various temperatures, using PVA as the primary material [Singh and Sharma 2012]. The adhesion between composite materials was less robust at elevated temperatures. Ultrasonication improved the quality of the blended material for blends by reacting to the uniformity of the whole mixture. Extensive studies have been conducted on the use of wheat gluten for the production of durable bioplastics. Although the material is

inherently fragile, its structure can be strengthened by using additives and fillers. Wheat gluten has a substantial protein concentration, rendering it appropriate for various uses. PBS is a newly developed biopolymer that has the potential to fulfill the market's need for bioplastics. PBS is often favoured over low-density polyethylene and polypropylene (PP). Due to its efficient processing capabilities, PBS is especially favoured in the textile industry. Examples of materials derived from it are meltblown and split yarn.

Additionally, it is employed to produce molded items in the plastics sector [Singh and Sharma 2012]. When PBS is combined with other polymers, its mechanical properties are enhanced, enabling its use in a broader range of applications. The amalgamation of PBS and *Spirulina* can improve the cost-effectiveness of *Spirulina*-based bioplastics. Multiple chemicals were employed in tests to enhance the combining process's efficiency and improve the result's quality [Subramanian 2017]. PVA is renowned for enhancing a product's strength, durability, and flexibility.

However, it is necessary to modify PVA with MA to enhance the dimensional stability and surface properties of packaging materials and their mechanical characteristics. The biomass was rinsed with acetone to obtain tiny spherical particles suitable for mixing. Sodium sulphide was employed in bioplastic synthesis to cleanse biomass before amalgamating it with other constituents. To enhance the combiningproperties of the benzoyl peroxide (BPO) and *Chlorella*-PP composite, it was mixed with MA and acetone and then put onto powdered PE [Thielen 2014].

9. Plasticizers and compatibilizers

Plasticizers are expansive chemical molecules used in substances to enhance their flexibility and ease of processing. The efficacy of the plasticizer is judged by its ability to render the target material more pliable [Tibbetts 2015]. Glycerol, with the chemical formula C3H8O3, is a widely used plasticizer in the production of bioplastics derived from microalgae. Glycerol enhanced the accessibility of macromolecules for the process of breakdown [Torres et al. 2015], resulting in higher flexibility and primary extensibility. This led to the production of high-quality products and increased elongation in the study. The plasticization capabilities of octanoic acid and glycerol were also observed. A series of tests showed that 1, 4-butanediol, and glycerol are suitable for plasticization due to their high water permeability [Tredici 2004]. In addition, carboxymethylcellulose (CMC) was employed to augment the mechanical characteristics of the plastic. CMC is a compound formed by alkali, cellulose, and chloroacetic acid reactions. It dissolves rapidly in cold water, has low primary viscosity, and is resistant to changes in temperature [Klunklin et al. 2021]. Compatibilizers are employed to facilitate the bonding of two polymers. The process of compatibilization improved the mechanical

strength of the heterogeneous biopolymers. Different types of blends can utilize various compatibilizers, including poly(ethylene-co-glycidyl) meth acryloyl carbamate, grafted ethylene/propylene rubber, maleic anhydride, and diethyl succinate [Troschl 2018]. The use of maleic anhydride enhanced the flexibility and uniformity of the products. PE-g-MA was incorporated into UHMW-PE and Spirulina composites at eight different concentrations, with each concentration rising by 3%. The addition did not have a specific impact on the mechanical characteristics of the composite. In addition to the plasticizers and compatibilizers mentioned above, compatibilizer initiators such as potassium peroxide sulphate (KPS) were used. The process involved liquefying DMSO (15 mL), MA, and PVA, followed by the addition of KPS (1% PVA) [Uduman *et al.* 2010].

10. Life Cycle Assessment Studies on Bioplastic Production from Microalgae

Limited studies on Life Cycle Assessment (LCA) exist, particularly for bioplastics generated from microalgae. The researchers surveyed the production of PLA using microalgae and plant sources. They found that using microalgae significantly impacts land utilization and terrestrial ecotoxicity [Bussa *et al.* 2019]. The investigation of the effects of greenhouse gases on several microalgae cultivation methods for bioplastic production revealed that various cyclic flow photobioreactors exhibited little variations compared to mixed systems and open raceway ponds. However, the study's findings do not reveal the comparative performance of microalgae-based bioplastics concerning conventional alternatives [Ugwu et al. 2008].

Nevertheless, extensive studies on microalgae farming using Life Cycle Assessment (LCA) might provide insights into specific patterns and trends [Valerie 2007]. According to the analysis of the LCA trials, fossil fuels outperformed microalgae-derived biofuels [Ali et al. 2023]. According to their study, microalgal manufacturing systems show significantly more efficiency increases in greenhouse gas emissions than fossil fuels [Vonshak 1997]. Data uncertainty significantly affects life cycle assessments (LCAs) on microalgae biofuel production, leading to divergent outcomes [Wang 2014]. Microalgae-based food production has high efficiency in land utilization, but it falls short in other critical areas, such as meeting the demand for freshwater [Wang et al. 2016]. The natural performance of microalgae-based products is primarily unclear. However, studies often highlight the possibility of improvement in microalgal production systems [Wijffels et al. 2010]. One way to establish synergies is by implementing bio-refineries that generate many byproducts and enhance farming techniques. Utilizing microalgal waste to produce bioplastics might enhance overall life cycle assessment (LCA) ratings. Microalgae production systems generally focus on minimizing land usage [Wretfors 2009]. Table 2 demonstrates the diverse applications of PHB in different sectors.

Table 2. W	ay of using	g PHB
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SL.NO	Area	Application
1	medical field	tissue cultures for implants, part of bones, Surgical implants and engineering of heart valves, pins, replanted veins, Sutures and dressings
2	Package	food package
3	Pharmacological	Encapsulation of different medicines for susceptible release
4	Environmental	Bottles, items of personal hygiene, involvement, remediation of areas affected by oil spills, bags, and disposal items.
5	Agricultural	Encapsulation of fertilizers
6	Industrial	Recovery of monomers and oligomers for new application in the synthesis of polymers
7	Agricultural	Encapsulation of fertilizers

11. Tissue engineering

Typically, the PHAs that were accessible were not used for medical implants. Consequently, the PHA quality evaluation has been postponed to obtain permission from Drug Administrators. It is necessary to generate PHAs with a high purity level and thoroughly examine their biodegradation in laboratory settings and their potential for use in scaffold creation and surface modification [Xu et al. 2010]. PHAs that undergo crucial modifications can substantially contribute to tissue engineering and the development of medicinal and therapeutic goods, particularly for applications such as vascular grafts, heart valves, and neural tissue engineering [Yan 2016]. PHAs with enhanced mechanical strength can be utilized to fabricate scaffolds for medicinal purposes. Scaffolds fabricated using PHAs facilitate cellular development by providing nourishment [Zeller et al. 2013]. The primary medical goods include screws, pins, sutures, films, and other similar items [Zhang et al. 2019]. Poly(3hydroxybutyrate-4-hydroxybutyrate-3-hydroxyvalerate) promotes the proliferation and adherence of stem cells [Zhang, Endo et al. 2000]. The polymer (3HB-3HV-3HHx) can serve as a scaffold to build liver tissue [Zhang, Kabeya *et al.* 2000].

Additionally, the use of PHA nanofibers has led to the development of three-dimensional scaffolds [Zhou *et al.* 2011]. The study discovered that P(3HB-3HO) effectively repaired the cartilage using a scaffold made from PHAs [Zhu *et al.* 2017]. The recombinant organism produces the novel P(3-HB-3HV-2,3-diHB). To improve the capacity, inorganic bio ceramics have been integrated with PHAs to boost the mechanical strength and flexibility of PHAs, resulting in the development of innovative composites for engineering tissues. Blends consisting of PHA and ceramic composites are utilized to create various mixtures. Hydroxyapatite and PHA are employed in tissue engineering as well.

12. Conclusions

This study examined the current state of bio-plastic production using resources derived from microalgae. *Chlorella* and *Spirulina* were the dominant algae species that produced plastic blends and biopolymers. To improve

the overall quality of the final product, several additives such as compatibilizers, plasticizers, and other chemicals were employed to combine the ingredients. Based on the literature assessment of this study, further advancement of techniques for producing bioplastics from microalgae is necessary to address economic feasibility concerns in large-scale industrial applications, which hinder the general adoption of microalgae-based bioplastic products in the market. Bioplastics were derived from the byproduct of efficient chemicals produced from microalgae as part of a biorefinery concept. Therefore, it may be seen as a very efficient product. In addition, different chemicals might impose restrictions on the potential applications of microalgae products, particularly in areas like healthcare and food packaging. Further research is required to enhance the efficiency of industrial and manufacturing operations while minimizing the need for additives through more inventive design.

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References

- Abdo, S.M.; Ali, G.H. (2019). Analysis of polyhydroxybutrate and bioplastic production from microalgae. *Bull. Natl. Res. Cent.* 43, 207
- Abu, Y. (Ed.) (2020). Microalgae Cultivation for Biofuels Production. Fundamentals of Microalgae Cultivation; Acedemic Press: Kumargaon, Bangladesh.
- Alaerts, L.; Augustinus, M.; van Acker, K. (2018). Impact of biobased plastics on current recycling of plastics. *Sustainability*. 10, 1487.
- Amaro, H.M.; Malcata, F.X.; Sousa-Pinto, I. (2014). Production and Supply Logistics of Microalgae as an Energy Feedstock. In Sustainable Bioenergy Production; CRC Press: Boca Raton, FL, USA, pp. 148–167.
- Barros, A.I.; Gonçalves, A.L.; Simões, M.; Pires, J.C.M. (2015). Harvesting techniques applied to microalgae: A review. *Renew. Sustain. Energy Rev.* 41, 1489–1500. (Accessed on 11 April 2020).

- Beckstrom, B.D. (2019). Bioplastic Production from Microalgae with Fuel Co-Products: A Techno-Economic and Life-Cycle Assessment. Master's Thesis, Colorado State University, Fort Collins, CO, USA.
- Biron, M. (2016). Recycling: The First Source of Renewable Plastics. In Industrial Applications of Renewable Plastics; Elsevier: Amsterdam, The Netherlands, pp. 67–114. ISBN 9780323480666.
- Brányiková, I.; Maršálková, B.; Doucha, J.; Brányik, T.; Bišová, K.; Zachleder, V.; Vítová, M. (2011). Microalgae—Novel highly efficient starch producers. *Biotechnol. Bioeng.* 108, 766–776.
- Brennan, L.; Owende, P. (2010). Biofuels from microalgae—A review of technologies for production, processing, and extractions of biofuels and co-products. *Renew. Sustain. Energy Rev.* 14, 557–577.
- Bussa, M.; Eisen, A.; Zollfrank, C.; Röder, H. (2019). Life cycle assessment of microalgae products: State of the art and their potential for the production of polylactid acid. J. Clean. Prod. 213, 1299–1312.
- Carlozzi, P. (2003). Dilution of solar radiation through "culture" lamination in photobioreactor rows facing south-north: A way to improve the efficiency of light utilization by cyanobacteria (Arthrospira platensis). *Biotechnol. Bioeng.* 81, 305–315.
- Chandra, R.; Mohan, S.V. (2014). Enhanced bio-hydrogenesis by co-culturing photosynthetic bacteria with acidogenic process: Augmented dark-photo fermentative hybrid system to regulate volatile fatty acid inhibition. *Int. J. Hydrogen Energy* 39, 7604–7615.
- Charlie, M.; Pauline, P.; Benjamin, G.; Jean-François, S.; Florian, D.; Nicolas, L.M. (2019). Microalgae starch-based bioplastics: Screening of ten strains and plasticization of unfractionated microalgae by extrusion. *Carbohydr. Polym.*, 208, 142–151.
- Chen, C.-Y.; Yeh, K.-L.; Aisyah, R.; Lee, D.-J.; Chang, J.-S. (2011). Cultivation, photobioreactor design and harvesting of microalgae for biodiesel production: A critical review. *Bioresour. Technol.* 102, 71–81.
- Chisti, Y. (2008). Biodiesel from microalgae beats bioethanol. *Trends Biotechnol.*, 26, 126–131.
- Chisti, Y. (2007). Biodiesel from microalgae. *Biotechnol. Adv.* 25, 294–306.
- Ciapponi, R.; Turri, S.; Levi, M. (2019). Mechanical Reinforcement by Microalgal Biofiller in Novel Thermoplastic Biocompounds from Plasticized Gluten. *Materials*, 12, 1476.
- Costa, S.S.; Miranda, A.L.; de Morais, M.G.; Costa, J.A.V.; Druzian, J.I. (2019). Microalgae as source of polyhydroxyalkanoates (PHAs)—A review. *Int. J. Biol. Macromol.* 131, 536–547.
- Das, S.K.; Sathish, A.; Stanley, J. (2018). Production of Biofuel and Bioplastic from Chlorella Pyrenoidosa. Mater. *Today Proc.* 5, 16774–16781.
- Di Caprio, F.; Visca, A.; Altimari, P.; Toro, L.; Masciocchi, B.; laquaniello, G.; Pagnanelli, F. (2016). Two stage process of microalgae cultivation for starch and carotenoid production. *Chem. Eng. Trans.* 49, 415–420.
- Dianursanti, C.N.; Windiani, L.; Gozan, M. (2019). Effect of Compatibilizer Addition in Spirulina Platensis Based Bioplastic Production; American Institute of Physics Conference Series: Erbil, Iraq.
- Dianursanti, S.A.K.; Khalis, S.A. (2018). The Effect of Compatibilizer Addition on Chlorella Vulgaris Microalgae

Utilization as a Mixture for Bioplastic; E3S Web of Conferences; EDP Sciences: Bali, Indonesia.

- DiGregorio, B.E. (2009). Biobased performance bioplastic: Mirel. *Chem. Biol.* 16, 1–2.
- Dixon, C.; Wilken, L.R. (2018). Green microalgae biomolecule separations and recovery. *Bioresour. Bioprocess.* 5, 1–24.
- Doucha, J.; Straka, F.; Lívanský, K. (2005). Utilization of flue gas for cultivation of microalgae Chlorella sp.) in an outdoor open thin-layer photobioreactor. J. Appl. Phycol. 17, 403–412.
- Draaisma, R.B.; Wijffels, R.H.; Slegers, P.M.E.; Brentner, L.B.; Roy, A.; Barbosa, M.J. (2013). Food commodities from microalgae. *Curr. Opin. Biotechnol.* 24,169–177.
- Eagan, J.M.; Xu, J.; Di Girolamo, R.; Thurber, C.M.; Macosko, C.W.; LaPointe, A.M.; Bates, F.S.; Coates, G.W. (2017). Combining polyethylene and polypropylene: Enhanced performance with PE/iPP multiblock polymers. *Science* 355, 814–816.
- European Bioplastics. (n.d.). Bioplastics Market Data. Available online: https://www.european-bioplastics.org/market/ (accessed on 1 April 2020).
- Berlin European Bioplastics. (2018). Fact Sheet, European Bioplastics: What are the bioplastics? European Bioplastics e.V. Germany.
- Fabra, M.J.; Martínez-Sanz, M.; Gómez-Mascaraque, L.G.; Gavara, R.; López-Rubio, (2018). A. Structural and physicochemical characterization of thermoplastic corn starch films containing microalgae. *Carbohydr. Polym.* 186, 184–191.
- Fu, W.; Gudmundsson, O.; Feist, A.M.; Herjolfsson, G.; Brynjolfsson, S.; Palsson, B.Ø. (2012). Maximizing biomass productivity and cell density of Chlorella vulgaris by using light-emitting diode-based photobioreactor. *J. Biotechnol.* 161, 242–249.
- Gifuni, I.; Olivieri, G.; Krauss, I.R.; D'Errico, G.; Pollio, A.; Marzocchella, A. (2017). Microalgae as new sources of starch: Isolation and characterization of microalgal starch granules. *Chem. Eng. Trans.* 57, 1423–1428.
- Gilbert, M. (2016). Brydson's Plastics Materials. Chapter 23— Bioplastics: New Routes, New Products; William Andrew: Amsterdam, The Netherlands, ISBN 0323370225.
- Gozan, M.; Noviasari, C. (2018) The Effect of Glycerol Addition as Plasticizer in Spirulina Platensis Based Bioplastic; Dianursanti, M., Ed.; E3S Web of Conferences; EDP Sciences: Bali, Indonesia.
- Hai, T.; Ahlers, H.; Gorenflo, V.; Steinbüchel, A. (2000). Axenic cultivation of anoxygenic phototrophic bacteria, cyanobacteria, and microalgae in a new closed tubular glass photobioreactor. *Appl. Microbiol. Biotechnol.* 53, 383–389.
- Harun, R.; Danquah, M.K.; Forde, G.M. (2009). Microalgal biomass as a fermentation feedstock for bioethanol production. J. Chem. Technol. Biotechnol. 99.
- Hempel, F.; Bozarth, A.S.; Lindenkamp, N.; Klingl, A.; Zauner, S.; Linne, U.; Steinbüchel, A.; Maier, U.G. (2011). Microalgae as bioreactors for bioplastic production. *Microb. Cell Factories* 10, 81.
- Huo, Y.-X.; Cho, K.M.; Rivera, J.G.L.; Monte, E.; Shen, C.R.; Yan, Y.; Liao, J.C. , (2011). Conversion of proteins into biofuels by engineering nitrogen flux. *Nat. Biotechnol.* 29, 346–351.
- Johnsson, N.; Steuer, F. (2018). Bioplastic Material from Microalgae: Extraction of Starch and PHA from Microalgae to

Create a Bioplastic Material; KTH Royal Institute of Technology: Stockholm, Sweden.

- Kalia, S.; Avérous, L. (2016). Biodegradable and Biobased Polymers for Environmental and Biomedical Applications; John Wiley & Sons: Beverly, MA, USA, ISBN 1119117348.
- Kaparapu, J. (2018). Polyhydroxyalkanoate (PHA) Production by Genetically Engineered Microalgae: A Review. J. New Biol. Rep. 7, 68–73.
- Kato, N. (2019) Production of crude bioplastic-beads with microalgae: Proof-of-concept. *Bioresour. Technol. Rep.* 6, 81–84.
- Khanra, S.; Mondal, M.; Halder, G.; Tiwari, O.N.; Gayen, K.; Bhowmick, T.K. (2018). Downstream processing of microalgae for pigments, protein and carbohydrate in industrial application: A review. *Food Bioprod. Process.* 110, 60–84.
- Sruthy K Das., (2016). Production of Biofuel and Bioplastic From Chlorella Pyrenoidosa.
- A.P.A. (2018). Cassuriaga, Innovative polyhydroxybutyrate production by Chlorella fusca grown with pentoses.
- N. Uma Maheswari, (2011). Production of bioplastic using Spirulina platensis and comparison with commercial plastic.
- Nahid Azizi, (n.d.). Acid pretreatment and enzymatic saccharification of brown seaweed for cyanobacterium Synechocystis sp.
- Sayeda M. Abdo, (2019). Analysis of polyhydroxybutrate and bioplastic production from microalgae.
- Rebecca Robert, (2018). Isolation and Optimization of PHB (Polyβ-hydroxybutyrate) Based Biodegradable Plastics from Chlorella vulgaris.
- Murilo Moraes Mourão, Optimization of Polyhydroxybutyrate Production by Amazonian Microalga Stigeoclonium sp. B23.
- Bhati R, Mallick N. (2014). Poly (3- hydroxybutyrate-co-3hydroxyvalerate) copolymer production by the diazotrophic cyanobacterium Nostoc muscorum Agardh: Process optimization and polymer characterization.
- Clemens Trosch, (2018). Pilot-scale production of poly-βhydroxybutyrate with the cyanobacterium Synechocytis sp. CCALA192 in a non-sterile tubular photobioreactor, Algal Research. 34:116-125.
- Monshupanee, T.; Nimdach, P.; Incharoensakdi, A. (2016). Twostage (photoautotrophy and heterotrophy) cultivation enable efficient production of bioplastic poly-3hydroxybutyrate in auto-sedimenting cyanobacterium.
- Ranjana Bhati (2010). Poly-β-hydroxybutyrate accumulation in cyanobacteria under photoautotrophy.
- Martins RG, Gonc, alves IS, Morais MG, Costa JAV. (2014). Bioprocess engineering aspects of biopolymer production by the cyanobacterium Spirulina strain LEB 18. Int J Polym Sci 2014:1–6.
- Gopi K, Balaji S, Muthuvelan B. (2014). Isolation purification and screening of biodegradable polymer PHB producing cyanobacteria from marine and fresh water resources. *Iranica J Energ Environ.* 5(1):94–100.
- Quines LKM, lenczak JL, Schmidt M, *et al.* (2015). Extrac,a[~]o de poli(3-hidroxibutirato), produzido por Cupriavidus necator, com carbonato de propileno. Quim Nova, a; 38:214–220.
- Riedel SL, Jahns S, Koenig S, *et al.* Polyhydroxyalkanoates production with Ralstonia eutropha from low quality waste animal fats. *J Biotechnol* 214: 119–127; 20 Chlorogloea fritschii

- Knuckey, R.M.; Brown, M.R.; Robert, R.; Frampton, D.M.F. (2006). Production of microalgal concentrates by flocculation and their assessment as aquaculture feeds. *Aquac. Eng.* 35, 300–313.
- Kutz, M. (2011). Applied Plastics Engineering Handbook: Processing and Materials; William Andrew: Amsterdam, The Netherlands, ISBN 1437735142.
- Lee, Y.-K. (2001). Microalgal mass culture systems and methods: Their limitation and potential. J. Appl. Phycol. 13, 307–315.
- Lupatini, A.L.; Colla, L.M.; Canan, C.; Colla, E. (2017). Potential application of microalga Spirulina platensis as a protein source. J. Sci. Food Agric, 97, 724–732, 2017.
- MacArthur, E. Beyond plastic waste. Science, 358, 843.
- Martin Chaplin. Carboxymethylcellulose (CMC). Available online: http://www1.lsbu.ac.uk/water/carboxymethylcellulose.html (accessed on 18 February 2020).
- McBride, R.C.; Lopez, S.; Meenach, C.; Burnett, M.; Lee, P.A.; Nohilly, F.; Behnke, C. (2014). Contamination Management in Low Cost Open Algae Ponds for Biofuels Production. *Ind. Biotechnol.* 10, 221–227
- Medeiros, D.L.; Sales, E.A.; Kiperstok, A. (2015). Energy production from microalgae biomass: Carbon footprint and energy balance. *J. Clean. Prod.* 96, 493–500.
- Milledge, J.J.; Heaven, S. (2011). Disc Stack Centrifugation Separation and Cell Disruption of Microalgae: A Technical Note. *Environ. Nat. Resour. Res.* 1, 17–24
- Mohan, S.V.; Hemalatha, M.; Chakraborty, D.; Chatterjee, S.; Ranadheer, P.; Kona, R. (2019). Algal biorefinery models with self-sustainable closed loop approach: Trends and prospective for blue-bioeconomy. *Bioresour. Technol.* 295, 122128.
- Monshupanee, T.; Nimdach, P.; Incharoensakdi, A. (2016). Twostage (photoautotrophy and heterotrophy) cultivation enable efficient production of bioplastic poly-3hydroxybutyrate in auto-sedimenting cyanobacterium. *Sci. Rep*, 6, 37121.
- Moreno-Garcia, L.; Adjallé, K.; Barnabé, S.; Raghavan, G.S.V. (2017). Microalgae biomass production for a biorefinery system: Recent advances and the way towards sustainability. Renew. Sustain. Energy Rev. 76, 493–506.
- Morgott, D.A. Acetone. In Patty's Toxicology, 5th ed.; Patty, F.A., Ed.; compl. rev. and updated.; Wiley: New York, NY, USA, p. 336. ISBN 9780471125471, 2001. [Google Scholar]
- Musa, M.; Ayoko, G.A.; Ward, A.; Rösch, C.; Brown, R.J.; Rainey, T.J. (2019). Factors Affecting Microalgae Production for Biofuels and the Potentials of Chemometric Methods in Assessing and Optimizing Productivity. *Cells*, 8, 851.
- Oh, Y.-K.; Hwang, K.-R.; Kim, C.; Kim, J.R.; Lee, J.-S. (2018). Recent developments and key barriers to advanced biofuels: A short review. *Bioresour. Technol.* 257, 320–333
- Olaizola, M. (2000). Commercial production of astaxanthin from Haematococcus pluvialis using 25,000-liter outdoor photobioreactors. J. Appl. Phycol. 12, 499–506
- Osanai, T.; Numata, K.; Oikawa, A.; Kuwahara, A.; Iijima, H.; Doi, Y.; Tanaka, K.; Saito, K.; Hirai, M.Y. (2013). Increased bioplastic production with an RNA polymerase sigma factor SigE during nitrogen starvation in Synechocystis sp. PCC 6803. DNA Res. 20, 525–535.
- Otsuki, T.; Zhang, F.; Kabeya, H.; Hirotsu, T. (2004). Synthesis and tensile properties of a novel composite of Chlorella and polyethylene. *J. Appl. Polym. Sci.* 92, 812–816.

- Pal, P.; Chew, K.W.; Yen, H.-W.; Lim, J.W.; Lam, M.K.; Show, P.L. (2019). Cultivation of Oily Microalgae for the Production of Third-Generation Biofuels. Sustainability, 11, 5424.
- Pandey, A.; Madamwar, D.; Rastogi, R.P. (2017). Algal Green Chemistry. Recent Progress in Biotechnology; Elsevier: Amsterdam, The Netherlands, ISBN 9780444640413.
- Park, S.-J.; Seo, M.-K. (2011). Interface Science and Composites; Academic Press: Burlington, NJ, USA, ISBN 0123750490.
- Pragya, N.; Pandey, K.K.; Sahoo, P.K. (2013). A review on harvesting, oil extraction and biofuels production technologies from microalgae. *Renew. Sustain. Energy Rev.* 24, 159–171.
- Proshad, R.; Kormoker, T.; Islam, M.S.; Haque, M.A.; Rahman, M.M.; Mithu, M.M.R. (2018). Toxic effects of plastic on human health and environment: A consequences of health risk assessment in Bangladesh. *Int. J. Health*, 6, 1–5.
- Pulz, O. (2001). Photobioreactors: Production systems for phototrophic microorganisms. Appl. Microbiol. *Biotechnol*. 57, 287–293.
- Quinn, J.C, Davis, R. (2015). The potentials and challenges of algae based biofuels: A review of the techno-economic, life cycle, and resource assessment modeling. *Bioresour. Technol.* 184, 444–452.
- Rahman, A.; Putman, R.J.; Inan, K.; Sal, F.A.; Sathish, A.; Smith, T.; Nielsen, C.; Sims, R.C.; Miller, C.D. (2015).
 Polyhydroxybutyrate production using a wastewater microalgae based media. *Algal Res.* 8, 95–98.
- Ranade, V.V. (2009). The Future of Glycerol-New Usages for a Versatile Raw Material; LWW: Cambridge, MA, USA
- Rawat, I.; Ranjith Kumar, R.; Mutanda, T.; Bux, F. (2011). Dual role of microalgae: Phycoremediation of domestic wastewater and biomass production for sustainable biofuels production. *Appl. Energy* 88, 3411–3424.
- Richmond, A.; Lichtenberg, E.; Stahl, B.; Vonshak, A. (1990). Quantitative assessment of the major limitations on productivity ofSpirulina platensis in open raceways. J. Appl. Phycol. 2, 195–206.
- Rudnik, E. Compostable Polymer Materials; Newnes: Amsterdam, The Netherlands, ISBN 0080994423, 2019.
- Sabathini, H.A.; Windiani, L.; Gozan, M. (2018). Mechanical Physicial Properties of Chlorella-PVA based Bioplastic with Ultrasonic Homogenizer. E3S Web Conf. 67, 3046.
- Salim, S.; Bosma, R.; Vermuë, M.H.; Wijffels, R.H. (2011). Harvesting of microalgae by bio-flocculation. J. Appl. Phycol. 23, 849–855.
- Sato, T.; Usui, S.; Tsuchiya, Y.; Kondo, Y. (2006). Invention of outdoor closed type photobioreactor for microalgae. *Energy Convers. Manag.* 47, 791–799.
- Scott, S.A.; Davey, M.P.; Dennis, J.S.; Horst, I.; Howe, C.J.; Lea-Smith, D.J.; Smith, A.G. (2010). Biodiesel from algae: Challenges and prospects. *Curr. Opin. Biotechnol.* 21, 277–286.
- Sereni, J.G.R. (2016). Reference module in materials science and materials engineering. *Mater. Sci. Eng. A*.
- Lopez-Exposito, P. *et al.* (2019) 'Microalgae harvesting with the novel flocculant hairy cationic nanocrystalline cellulose', Colloids and Surfaces B: Biointerfaces, 178, pp. 329–336.
- Shen, Y.; Yuan, W.; Pei, Z.J.; Wu, Q.; Mao, E. (2009). Microalgae Mass Production Methods. Trans. ASABE 52, 1275–1287.

- Singh, A.K.; Sharma, L.; Mallick, N.; Mala, J. (2017). Progress and challenges in producing polyhydroxyalkanoate biopolymers from cyanobacteria. J. Appl. Phycol. 29, 1213–1232
- Singh, R.N.; Sharma, S. (2012). Development of suitable photobioreactor for algae production—A review. Renew. Sustain. Energy Rev. 16, 2347–2353, 2012.
- Subramanian, V.D.V. (Ed.) (2017). Biopolymer Composites in Electronics. Thermoelectric Properties of Biopolymer Composites; Elsevier: Amsterdam, the Netherlands.
- Thielen, M. (2014). Bioplastics: Plants and Crops Raw Materials Products; Fachagentur Nachwachsende Rohstoffe eV (FNR) Agency for Renewable Resources: Gülzow, Germany
- Tibbetts, S.M.; Milley, J.E.; Lall, S.P. (2015). Chemical composition and nutritional properties of freshwater and marine microalgal biomass cultured in photobioreactors. J. Appl. Phycol. 27, 1109–1119.
- Torres, S.; Navia, R.; Campbell Murdy, R.; Cooke, P.; Misra, M.; Mohanty, A.K. (2015). Green Composites from Residual Microalgae Biomass and Poly (butylene adipate- coterephthalate): Processing and Plasticization. ACS Sustain. Chem. Eng. 3, 614–624
- Tredici, M.R. (2004). Mass production of microalgae: Photobioreactors. Handb. Microalgal Cult. Biotechnol. Appl. Phycol. 1, 178–214, .
- Klunklin, W.; Jantanasakulwong, K.; Phimolsiripol, Y.; Leksawasdi,
 N.; Seesuriyachan, P.; Chaiyaso, T.; Insomphun, C.;
 Phongthai, S.; Jantrawut, P.; Sommano, S.R.; *et al.* (2021).
 Synthesis, Characterization, and Application of
 Carboxymethyl Cellulose from Asparagus Stalk End.
 Polymers, 13, 81.
- Troschl, C. (2008). Bioplastic production with cyanobacteria. Ph.D. Thesis, Universität für Bodenkultur Wien, Wien, Austria, 2018.
- Uduman, N.; Qi, Y.; Danquah, M.K.; Forde, G.M.; Hoadley, A. (2010). Dewatering of microalgal cultures: A major bottleneck to algae-based fuels. J. Renew. Sustain. Energy 2, 12701.
- Bussa, M. *et al.* (2019) 'Life cycle assessment of microalgae products: State of the art and their potential for the production of polylactid acid', *Journal of Cleaner Production*, 213, pp. 1299–1312.
- Ugwu, C.U.; Aoyagi, H.; Uchiyama, H. (2008). Photobioreactors for mass cultivation of algae. Bio resource. Technol. 99, 4021–4028.
- Valerie, C. (2007). Final report on the safety assessment of Polyethylene. *Int. J. Toxicol.* 26, 115–127
- Ali, S.S. et al. (2023) 'Bioplastic production in terms of life cycle assessment: A state-of-the-art review', Environmental Science and Ecotechnology, 15, p. 100254.
- Vonshak, A. (1997). Outdoor mass production of Spirulina: The basic concept. In Spirulina Platensis Arthrospira; CRC Press: Boca Raton, FL, USA, pp. 97–118
- Wang, K. (2014). Bio-Plastic Potential of Spirulina Microalgae. Master's Thesis, The University of Georgia, Athens, GA, USA, 2014.
- Wang, K.; Mandal, A.; Ayton, E.; Hunt, R.; Zeller, M.A.; Sharma, S. (2016). Chapter 6—Modification of Protein Rich Algal-Biomass to Form Bioplastics and Odor Removal. In Protein Byproducts; Academic Press: Cambridge, MA, USA, pp. 107–117. ISBN 978-0-12-802391-4.

- Wijffels, R.H.; Barbosa, M.J.; (2010). Eppink, M.H.M. Microalgae for the production of bulk chemicals and biofuels. *Biofuels Bioprod. Biorefin.* 4, 287–295.
- Wretfors, C.; Cho, S.-W.; Hedenqvist, M.S.; Marttila, S.; Nimmermark, S.; Johansson, E. (2009). Use of industrial hemp fibers to reinforce wheat gluten plastics. *J. Polym. Environ.* 17, 259
- Xu, J.; Guo, B.-H. (2010). Poly (butylene succinate) and its copolymers: Research, development and industrialization. *Biotechnol. J.* 5, 1149–1163.
- Yan, Y. (2016). Developments in fibers for technical nonwovens. In Advances in Technical Nonwovens; Elsevier: Sawston, UK, pp. 19–96
- Zeller, M.A.; Hunt, R.; Jones, A.; (2013). Sharma, S. Bioplastics and their thermoplastic blends from Spirulina and Chlorella microalgae. *J. Appl. Polym. Sci.* 130, 3263–3275.
- Zhang, C.; Wang, C.; Cao, G.; Wang, D.; Ho, S.-H. (2019). A sustainable solution to plastics pollution: An eco-friendly bioplastic film production from high-salt contained Spirulina sp. residues. J. Hazard. Mater. 121773.
- Zhang, F.; Endo, T.; Kitagawa, R.; Kabeya, H.; Hirotsu, T. (2000). Synthesis and characterization of a novel blend of polypropylene with Chlorella. *J. Mater. Chem.* 10, 2666–2672, 2000.
- Zhang, F.; Kabeya, H.; Kitagawa, R.; Hirotsu, T.; Yamashita, M.; Otsuki, T. (2000). An exploratory research of PVC-Chlorella composite material (PCCM) as effective utilization of Chlorella biologically fixing CO₂. J. Mater. Sci. 35, 2603–2609, 2000.
- Zhou, N.; Zhang, Y.; Wu, X.; Gong, X.; Wang, Q. (2011). Hydrolysis of Chlorella biomass for fermentable sugars in the presence of HCl and MgCl2. *Bioresour. Technol.* 102, 10158–10161, 2011.
- Zhu, N.; Ye, M.; Shi, D.; Chen, M. (2017). Reactive compatibilization of biodegradable poly (butylene succinate)/Spirulina microalgae composites. *Macromol. Res.* 25, 165–171, 2017.