
1 **Study on the removal performance of modified *Enteromorpha*-**

2 **immobilized bacterial agent in oil contaminated seawater**

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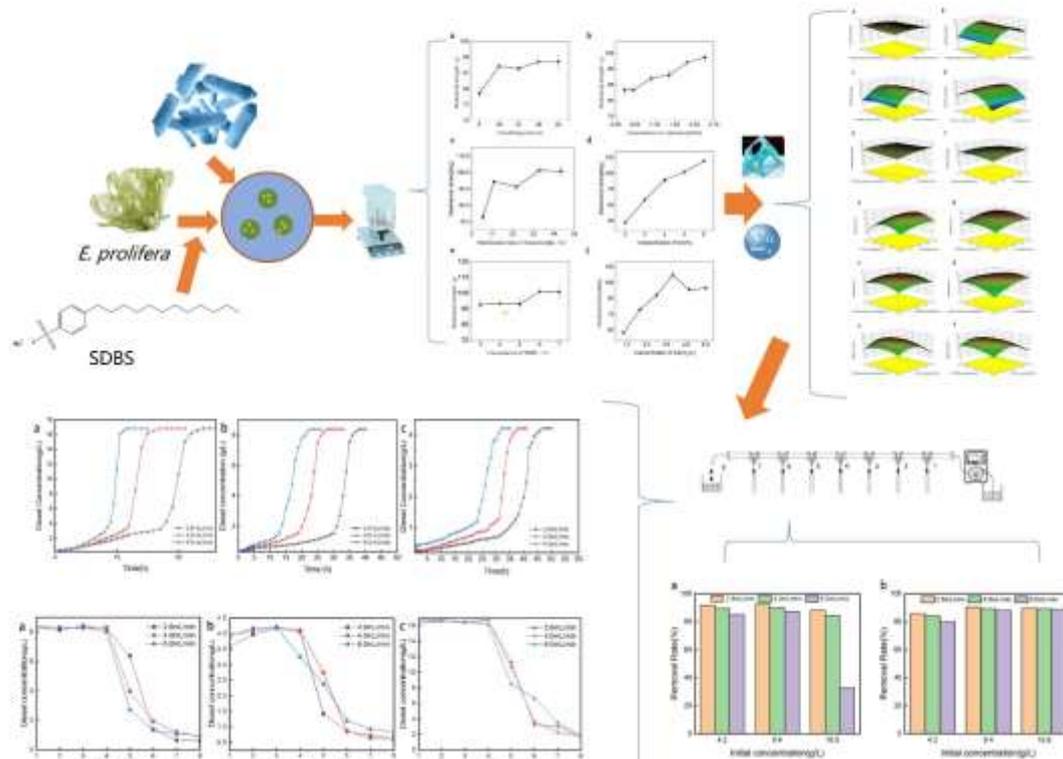
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14 **GRAPHICAL ABSTRACT**

The modified *Enteromorpha*-immobilized bacterial agent was optimized to reach excellent removal efficiency of diesel (90.39%) and recyclability.



15

16 **Abstract**

17 Compared with free bacteria, immobilized bacteria have a higher oil degradation rate,
 18 making them highly promising for the bioremediation of offshore oil pollution. In our
 19 study, preparation conditions of modified *Enteromorpha*-immobilized bacterial agent
 20 were optimized by response surface method. A reactor was constructed and the effect
 21 of diesel concentration and flow rate on the removal rate were investigate. The
 22 recyclability of modified *Enteromorpha*-immobilized bacterial agent was further
 23 studied. The result showed that, under the optimal preparation conditions, the removal
 24 rate of diesel in 5 days reached 90.39% and the mechanical strength reached a
 25 maximum value of 173.67g. When the inlet flow rate was not greater than 6.0mL/min
 26 and the concentration of diesel was less than 16.8g/L, the removal rate of diesel was
 27 guaranteed within 10.5h. After three cycles of diesel breakthrough, the removal rate can
 28 still attain 82.35%. This study provides a valuable insight into the practical application
 29 of immobilization technology to remove petroleum from seawater in the future.

30 **Keywords:** Oil pollution; *Enteromorpha prolifera*; Modification; Immobilized
31 bacterial agent

32

33 **1. Introduction**

34 Marine oil pollution was caused by the leakage of oil during the process of
35 exploitation, transportation, refining and use, which has brought huge disasters to
36 marine life and environment(Chen et al., 2020a; Chen et al., 2020b; Li et al., 2020;
37 Naeem and Qazi, 2020). As oil pollutants are persistent and bioaccumulative(Yang et
38 al., 2021), the establishment of practical and effective restoration and control measures
39 is imminent.

40 To remediate areas contaminated with oil, various traditional technologies have been
41 employed, including thermal extraction, steam stripping, adsorption, and
42 bioremediation(Xue et al., 2019). Among them, bioremediation has been proved to be
43 a low-cost, easy-to-operate, and pollution-free repair method, which has developed
44 rapidly in recent years(Tang et al., 2019; Wang et al., 2019). Microbial agents has been
45 widely employed to remediate oil-contaminated soil. However, their effectiveness may
46 be compromised when added to the ocean, as the agents can be inactivated and their
47 density reduced. Simultaneously, the low solubility, non-polarity and hydrophobicity of
48 oil also limited the oil degradation(Fu et al., 2019; Pereira et al., 2019; Varjani and
49 Upasani, 2017; Xu et al., 2018b). The immobilized microorganism technology confines
50 microbial agents within an enclosed space, making it an effective solution for the
51 aforementioned issues(Kumar et al., 2019). Compared with free cells, immobilized
52 system provides a more stable living environment for microorganisms(Abu Talha et al.,
53 2018; Qin et al., 2013; Xue et al., 2022).

54 Although considerable attention has been focused on developing immobilization
55 carriers with several specific properties (including strength, chemical and thermal
56 stability, durability and surface properties), the activity of immobilized bacteria is still
57 limited due to the lack of nutrients. Chen (2012) applied polyurethane-polyurea
58 copolymer as an immobilization carrier and determined that the degradation efficiency

59 is only 47.25% without providing sufficient nutrients(Dehnavi and Ebrahimipour,
60 2022). Alginates are a heterogeneous group of polymers, with a wide range of
61 functional properties, their success as immobilization matrices will rely on an
62 appropriate choice of materials and methodology for each application(Mardiana et al.,
63 2019; Smidsrod and Skjakbraek, 1990). The alginate could improve the degradation
64 rate of diesel. Common methods for microbial immobilizing include adsorption,
65 embedding, cross-linking, and covalent bonding(Zhang et al., 2010). The embedding
66 immobilization technique is widely used due to its simple operation, mild reaction
67 conditions, low risk of microorganism leakage, excellent stability and reusability, high
68 microbial activity and cell capacity, and high immobilization efficiency (>70%)
69 compared to other methods like adsorption, cross-linking, and covalent
70 bonding(Kuyukina et al., 2009; Wu et al., 2009).

71 The *E. proliferans* formed large-scale blooms (the so-called “green tide”) from 2007
72 in the Yellow Sea, China. Currently, the majority of *E. proliferans* collected from the sea
73 is considered as solid waste. Therefore, it is urgent to develop *E. proliferans* utilization
74 technique and reduce the pressure of marine environmental pollution. At present, the
75 most common utilization of *E. proliferans* is fertilizer and feed. *E. proliferans* was used to
76 prepare a tunable amphiphilic phyllox modified graphene gascoagulant for oil-water
77 separation(Ji et al., 2021). Hydrogel calcium alginate microparticles derived from *E.*
78 *prolifera* were utilized as a biomaterial to remove heavy metals from water(Duc et al.,
79 2021). *E. proliferans* is composed of a single layer of cells, surrounded by a tubular or
80 adhesive ribbon, this unique physical structure provides considerable surface for
81 adsorption. In addition, it would be more porous after modification by sodium
82 dodecylbenzene sulfonate (SDBS), and its adsorption capacity was further
83 improved(Xu et al., 2018a; Yang et al., 2021). Meanwhile, it can provide nutrients for
84 growth of bacteria such as N and P(Shi et al., 2020). Therefore modified *E. proliferans* is
85 a suitable carrier for microbial immobilization, and this provides a new idea for the
86 resource utilization of *E. proliferans*.

87 In this paper, the *E. proliferans* was modified and combined with oil-degrading
88 bacteria to remove oil. The single factor experiment, response surface and the

89 breakthrough curve were used for this study. Therefore, to the aim of this study is: (1)
 90 determine optimal preparation conditions of the modified *Enteromorpha*-immobilized
 91 bacterial agent; (2) study the removal performance of the agent under the optimal
 92 conditions; (3) evaluate the recyclability of the agent.

93 2. Material and methods

94 2.1 Materials and strain

95 All the chemicals used in this study are of analytical grade. *E. prolifera* was collected
 96 from 35°57'33.22"N, 120°14'55.36"E in Golden Beach, Qingdao, July 2020. The
 97 highly efficient oil-degrading strain Sp8 (*Shewanella algae*) was isolated from seawater
 98 samples from 35°56'27.92"N, 120°12'52.59"E in Tangdao Bay, Qingdao. Diesel (0#)
 99 was purchased from Petro China (Qingdao Center, Huangdao District), and filtered
 100 with 0.22 μm membrane before using.

101 2.2 Single factor experiment

102
 103 Table 1. Single factor experimental design scheme

Single factor	Fixed condition
SDBS concentration (3%, 4%, 5%, 6%, 7%, w/v)	Digging time 24 h
	SA concentration 4%
	Cross-linking time 27 h
	CaCl ₂ concentration 3%
	<i>E. prolifera</i> concentration 1.5%
	SDBS concentration 5%
	SA concentration 4%
	Cross-linking time 27 h
	CaCl ₂ concentration 3%
	<i>E. prolifera</i> concentration 1.5%
Digging time (10 h, 20 h, 30 h, 40 h, 50 h, w/v)	SDBS concentration 5%
	SA concentration 4%
	Cross-linking time 27 h
	CaCl ₂ concentration 3%
	<i>E. prolifera</i> concentration 1.5%
	SDBS concentration 5%
	Digging time 24 h
	Cross-linking time 27 h
	CaCl ₂ concentration 3%
	<i>E. prolifera</i> concentration 1.5%
SA concentration (2%, 3%, 4%, 5%, 6%, w/v)	SDBS concentration 5%
	Digging time 24 h
	Cross-linking time 27 h
	CaCl ₂ concentration 3%
	<i>E. prolifera</i> concentration 1.5%
	SDBS concentration 5%
	Digging time 24 h
	SA concentration 4%
	Cross-linking time 27 h
	CaCl ₂ concentration 3%
<i>E. prolifera</i> concentration (0.25%, 0.5%, 1%, 1.5%, 2%, 2.5%, w/v)	Digging time 24 h
	SA concentration 4%
	Cross-linking time 27 h
	CaCl ₂ concentration 3%
	SDBS concentration 5%

CaCl ₂ concentration (1%, 2%, 3%, 4%, 5%, w/v)	SDBS concentration 5% Digging time 24 h SA concentration 4% Cross-linking time 27 h <i>E. prolifer</i> a concentration 1.5%
Cross-linking time (9 h, 18 h, 27 h, 36 h, 45 h)	SDBS concentration 5% Digging time 24 h SA concentration 4% CaCl ₂ concentration 3% <i>E. prolifer</i> a concentration 1.5%

104

105 The basic process of modified *Enteromorpha*-immobilized bacterial agent
 106 preparation was described by Yang et al (Yang et al., 2021). *E. prolifer*a was dipped in
 107 a SDBS solution, then dried and ground for later usage. After enrichment, bacteria and
 108 modified *E. prolifer*a were mixed into SA solution and transferred into CaCl₂ with a
 109 dropper to form beads. At last, the beads were stored at 4 °C containing CaCl₂ to cross-
 110 linking. The modified *Enteromorpha*-immobilized bacterial agent was stored in
 111 distilled water before use. Concentration of SA, SDBS, *E. prolifer*a, and time of dipping
 112 were used as influencing factors. A single factor experiment was designed to optimize
 113 the conditions and parameters of the immobilization process. Meanwhile the
 114 mechanical strength of modified *Enteromorpha*-immobilized bacterial agent was target.
 115 The design of single factor experimental was shown in Table 1. The mechanical
 116 strength was performed as described by Xue et al (Xue et al., 2019).

117 2.3 Optimization of preparation conditions

118 Table 2. Experimental design and experimental results of condition optimization

Run	A: Digging time(h)	B:SA concentration(%)	C:SDBS concentration(%)	D: Enteromorpha concentration (g)	Mechanical strength (g)	Removal rate(%)
1	24	5	7	2.0	135.333	60.459
2	48	4	6	2.0	88.333	67.488
3	36	5	6	2.0	164.750	85.214
4	36	5	5	1.5	137.000	49.016
5	24	5	5	2.0	148.000	57.216
6	36	4	6	1.5	94.333	52.530
7	36	6	6	1.5	118.667	47.394
8	48	5	6	1.5	187.333	95.241

9	48	6	6	2.0	107.000	49.106
10	48	5	5	2.0	173.667	79.472
11	36	5	6	2.0	174.000	78.543
12	48	5	6	2.5	167.000	73.255
13	24	5	6	2.5	149.667	63.343
14	36	5	6	2.0	154.000	80.421
15	36	5	5	2.5	155.667	66.406
16	24	5	6	1.5	148.333	55.233
17	36	4	6	2.5	175.000	76.776
18	24	6	6	2.0	115.667	36.221
19	36	5	7	2.5	165.000	86.590
20	36	6	5	2.0	124.333	54.422
21	24	4	6	2.0	84.000	60.550
22	36	5	6	2.0	164.750	87.672
23	36	6	7	2.0	119.000	49.286
24	36	4	7	2.0	81.000	65.596
25	36	6	6	2.5	135.667	66.406
26	36	5	7	1.5	157.000	64.514
27	48	5	7	2.0	137.000	88.753
28	36	4	5	2.0	89.000	62.262
29	36	5	6	2.0	156.000	87.554

119

120 Statistical Product Service Solutions (SPSS 15.0) was used for data analysis,
 121 according to the mechanical strength, the range of the four factors were: the dipping
 122 time of *E. prolifer*a of 24-48(h), SA concentration of 4-6%(w/v), SDBS concentration
 123 of 5-7%(w/v), *E. prolifer*a concentration of 1.5-2.5%(w/v) (Table 2).

124 Using the central combination design principle of Box-Benhnken, the factors of
 125 SA, SDBS, *E. prolifer*a concentrations and the dipping time were analyzed by response
 126 surface(Calera et al., 2020; Maleki and Karimi-Jashni, 2020), and the influences of
 127 agent prepared under different combinations of four factors on removal rate of diesel
 128 (%) in 5 days and mechanical strength (g) were discussed. The data of response surface
 129 method was analyzed using analysis of variance. The regression results were fitted to
 130 the experimental results using Design-Expert 8.0 software to gain quadratic multiple
 131 regression models, 3D surface plots with contour plots were established to obtain the
 132 optimal preparation conditions of modified *Enteromorpha*-immobilized microbial
 133 agent.

134 The mechanical strength of the modified *Enteromorpha*-immobilized bacterial
135 agent and its removal rate of diesel were taken as response values. The optimal
136 preparation conditions of the agent were obtained. The removal rate of diesel was
137 obtained according to the following formula:

$$138 \quad D = (1 - C_1/C_0) \times 100\% \quad (1)$$

139 Where C_0 (mL/L) is the initial concentration of diesel, C_1 (mL/L) is the residual
140 concentration of diesel, D (%) is the removal rate of diesel.

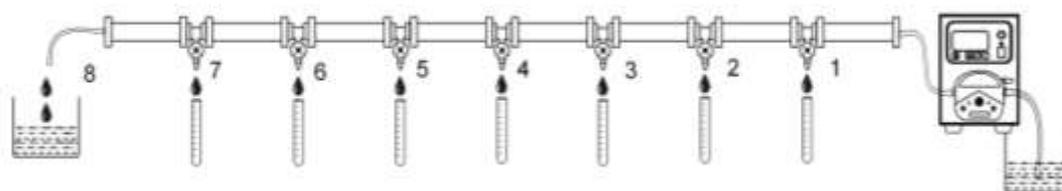
141 The predicted value was compared with the experimental value, and the reason was
142 analyzed.

143 **2.4 Research of removal rate**

144 The diesel (control group) and remaining diesel by modified *Enteromorpha*-
145 immobilized bacterial agent at 7 days were detected via gas chromatography-mass
146 spectrometry (GC-MS) (Agilent 19091S-433, USA). The method of the pre-treatment
147 GC-MS was performed as previously described by Shi et al. (Shi et al., 2019).

148 **2.5 Research on removal performance**

149 A reactor was designed to study removal performance of modified *Enteromorpha*-
150 immobilized bacterial agent. The reactor was composed of four parts: peristaltic pump,
151 sample tube, reaction column and outlet pipe (Figure 1). The reaction column is divided
152 into 8 sections, each length of 10 cm, and a water outlet and a final water outlet are set
153 between each two sections, numbered from outlets 1 to 8, to detect the concentration of
154 water (Figure 1). The modified *Enteromorpha*-immobilized bacterial agent was filled
155 into the reaction column.



158 To evaluate the impact of flow rates and diesel concentrations on the degradation
159 rate and removal effectiveness, the inlet flow rates were set as 2.6 mL/min, 4.0 mL/min,
160 6.0 mL/min, and inlet diesel concentrations were set as 4.2 g/L, 8.4 g/L, 16.8 g/L,
161 respectively. Degradation rates of outlets before breakthrough point at different inlet
162 flow rates and diesel concentration was measured, the breakthrough time (T), $1/2T$ and
163 10h for different diesel concentrations and flow rates was determined.

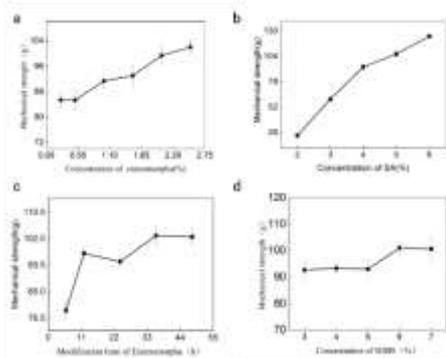
164 **2.6 Recyclability of modified *Enteromorpha*-immobilized bacterial agent**

165 Continuous removal and elution were used as reusability experiments. The removal
166 rate of diesel, surface adsorption and internal intake of each cycle was measured by
167 ultraviolet spectrophotometry. After the removal rate of diesel, surface adsorption and
168 internal intake was measured in each cycle, the modified *Enteromorpha*-immobilized
169 bacterial agent was filtered out and washed with physiological saline for 2–3 times, and
170 then the agent was added to the new degradation medium to start a new cycle of
171 experiments. Take five days as an experimental cycle, a total of three cycles of cyclic
172 experiments were carried out. In order to reduce experimental errors, each group of
173 experiments had three parallel samples.

174 **3. Results and discussion**

175 **3.1 Influence of different factors on mechanical strength**

176 The influence of four factors on the mechanical strength of the modified
177 *Enteromorpha*-immobilized bacterial agent was illustrated in Figure 2. The mechanical
178 strength increased as the concentration of SA and *E. prolifera* increased, with a
179 tendency to further increase as the SDBS concentration rose. The most significant
180 change was observed when the SDBS concentration increased from 5% to 6%. In
181 addition, With the extension of the modification time of the *E. prolifera*, the mechanical
182 strength tended to increase significantly, and the change was not obvious after 36 h.



183

184 Figure 2 Mechanical strength of immobilized petroleum degrading bacteria prepared under different (a)
 185 Concentration of *E. proliferata*; (b) Concentration of SA; (c) Modification time of *E. proliferata*; (d) Concentration of
 186 SDBS.

187 The concentrations of SA, SDBS, and *E. proliferata*, as well as the modification time,
 188 were included in the multi-factor experimental design (Table 3), which is consistent
 189 with the findings of Fu et al. (2022) (Fu et al., 2022). The multivariate ANOVA results
 190 of mechanical strength and removal rate of the modified *Enteromorpha*-immobilized
 191 bacterial agent was significant (Table 3, $P < 0.01$).

192

Table 3 Experimental settings of four factors and three levels

Factor	Level		
	-1	0	1
A: Dipping time	24	36	48
B:SA concentration	4	5	6
C: SDBS concentration	5	6	7
D: <i>E. proliferata</i> concentration	1.5	2	2.5

193 **3.2 Optimization of preparation conditions of modified *Enteromorpha*-**
 194 **immobilized microbial agent**

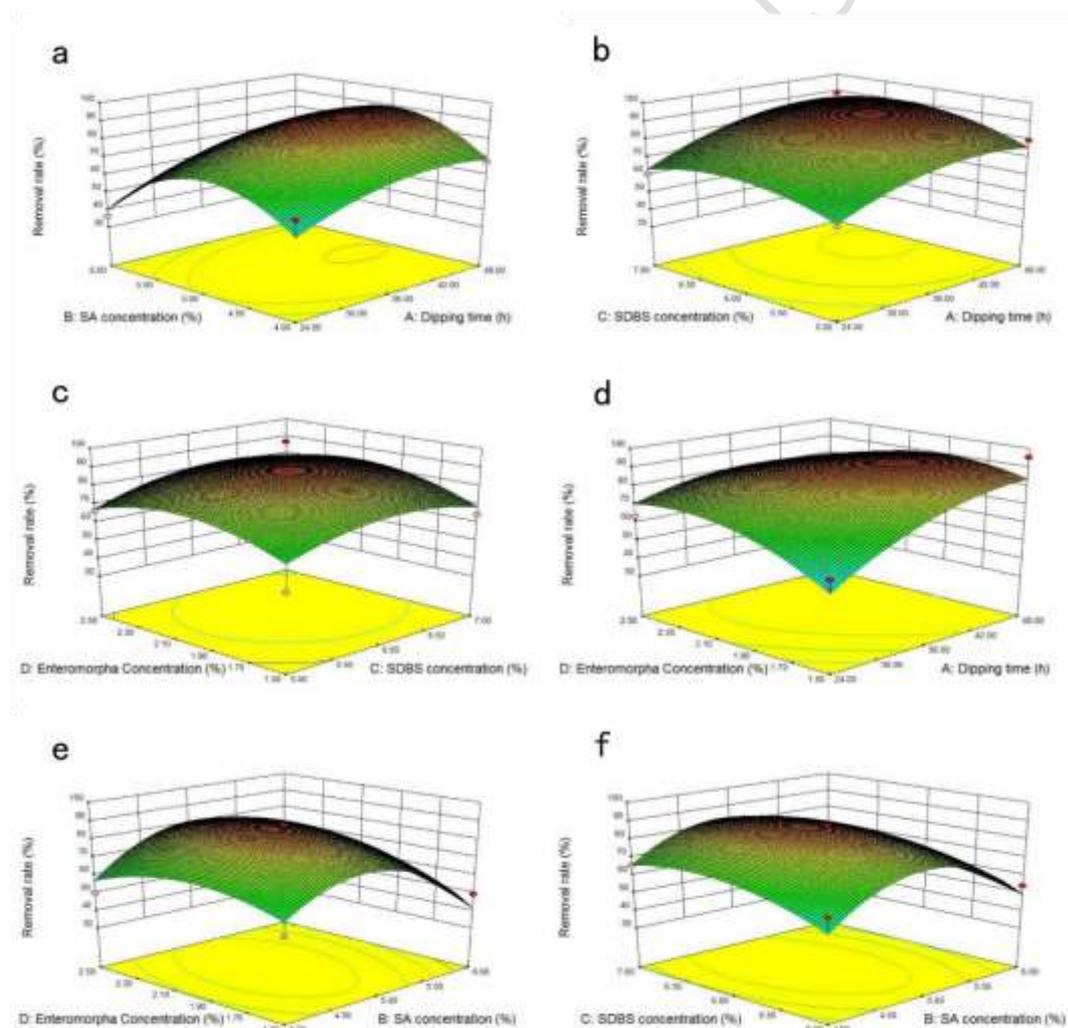
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Table 4. ANOVA response surface quadratic model

Source	Sum of Squares		df	Mean Square		F-Value		P-Value		
	Mechanical strength	Removal rate		Mechanical strength	Removal rate	Mechanical strength	Removal rate	Mechanical strength	Removal rate	
Model	24975	6129.06	14	1783.93	437.79	13.88	13.88	< 0.0001	0.0018	significant
A	524.48	1205.87	1	524.48	1205.87	4.08	14.56	0.0629	0.0019	
B	2914.11	259.27	1	2914.11	259.27	22.68	3.13	0.0003	0.0986	
C	92.6	179.44	1	92.60	179.44	0.72	2.17	0.4103	0.1631	
D	60.76	148.82	1	60.76	148.82	0.47	1.80	0.5029	0.2014	

AB	42.25	8.84	1	42.25	8.84	0.33	0.11	0.5755	0.7487
AC	144	9.11	1	144.00	9.11	1.12	0.11	0.3077	0.7450
AD	117.36	226.44	1	117.36	226.44	0.91	2.73	0.3555	0.1204
BC	1.78	17.94	1	1.78	17.94	0.014	0.22	0.9080	0.6488
BD	53.77	114.01	1	53.77	114.01	0.42	1.38	0.5282	0.2602
CD	28.45	5.49	1	28.45	5.49	0.22	0.066	0.6453	0.8005
A ²	316.58	423.82	1	316.58	423.82	2.46	5.12	0.1388	0.0401
B ²	19940.00	3641.68	1	19940.00	3641.68	155.16	43.98	< 0.0001	< 0.0001
C ²	573.49	455.12	1	573.49	455.12	4.46	5.50	0.0531	0.0343
D ²	23.14	663.13	1	23.14	663.13	0.18	8.01	0.6777	0.0134
Residual	1799.20	1159.22	14	128.51	82.80				
Lack of Fit	1799.20	1159.22	10	179.92	115.92				not significant

196



197

198

Figure 3 Response surface graph (response value: diesel removal rate. (A: Dipping time; B: SA concentration; C:

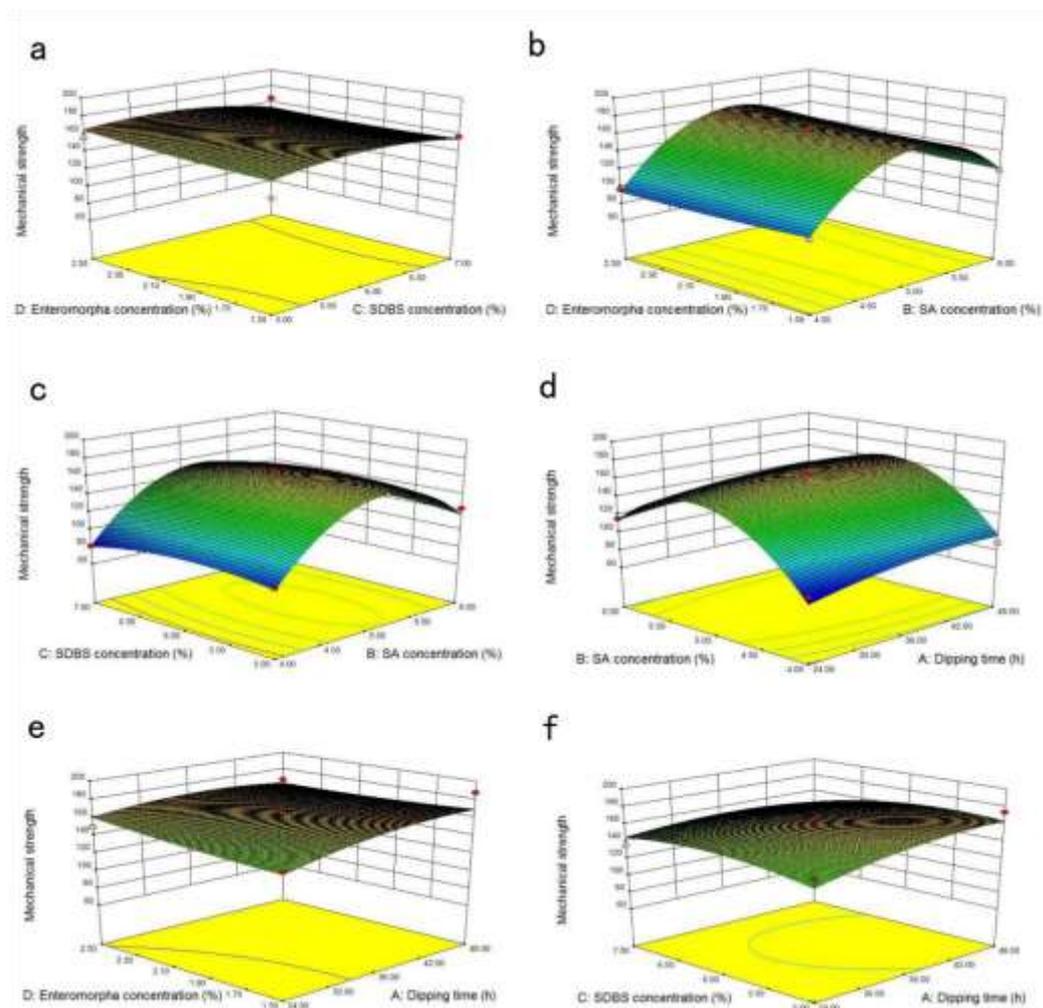
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SDBS concentration; D: *E. prolifer*a Concentration)

200 The coded independent variables (SA, SDBS, *E. prolifer*a concentrations, and
201 dipping time) and experimental values (removal rate of diesel and mechanical strength)
202 used in the RSM design are summarized in Table 4. The removal rate of diesel ranged
203 from 36.22% to 95.24%, while the mechanical strength varied between 81.00 and
204 187.33 g throughout the experiments. And the sum of squares (SS) should also be
205 regarded (Maleki and Karimi-Jashni, 2020). As the SS value increases, the importance
206 of the variable increases as well. The influences of SA, SDBS, *E. prolifer*a
207 concentrations and the dipping time on removal rate of diesel and mechanical strength
208 are revealed in Table 2. The results distinctly revealed that the model fit was statistically
209 significant ($P < 0.002$). The SA concentration presented a significant influence on
210 mechanical strength ($P < 0.0005$, $F=22.68$, $SS=2914.11$). For removal rate of diesel,
211 the influence of the dipping time was significant ($P < 0.002$, $F=14.56$, $SS=1205.87$).

212 From Figure 3 and 4, the maximum of removal rate of diesel and mechanical strength
213 under optimum condition were obtained. The maximum predicted values of the removal
214 rate of diesel and mechanical strength of the modified *Enteromorpha*-immobilized
215 bacterial agent was 90.71% and 166.56 g, respectively. The observed removal rates of
216 diesel using the modified *Enteromorpha*-immobilized immobilized bacterial agent are
217 higher than those reported for bacteria immobilized on cinnamon shell (Fu et al., 2019),
218 maize straw (Xue et al., 2019), and wood chips (Xue et al., 2017). Under optimal
219 conditions, the modified *Enteromorpha*-immobilized bacterial agent was prepared and
220 tested for removal rate of diesel and mechanical strength. Results showed a removal
221 rate of 90.39% and mechanical strength of 173.67 g.

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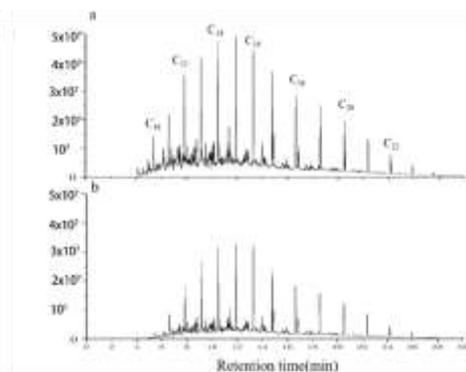


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Figure 4 Response surface graph (response value: mechanical strength. A: Dipping time; B: SA concentration; C: SDBS concentration; D: *Enteromorpha* Concentration.)

226

3.3 Removal rate of diesel by modified *Enteromorpha*-immobilized bacterial agent



227
228

Figure 5 GC-MS analysis of inlet diesel (a) and outlet diesel (b)

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230

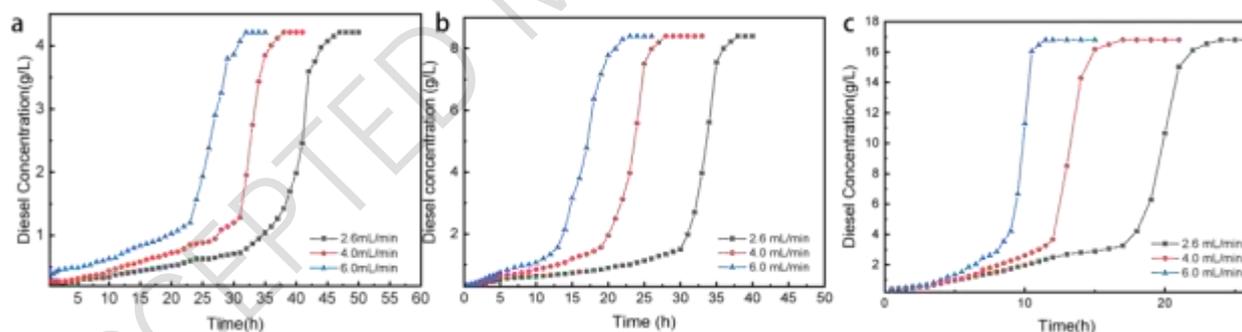
The chromatographic peak in Figure 5 indicated a decrease in all organic content, with C₁₇ demonstrating the most significant reduction and a removal rate of 73%. The

231 removal rate for C10 was also high at 64%, while removal rates for other hydrocarbons
232 were approximately 60%. Therefore, immobilized cells are particularly effective when
233 pollutants are predominantly composed of C₁₇.

234

235 3.4 Study of immobilizing agents for the removal performance of diesel

236 The breakthrough time ranged from 10.5 h to 47 h, with the highest value observed
237 at a diesel concentration of 4.2 g/L and an inlet flow rate of 2.6 mL/min, and the lowest
238 value observed at a diesel concentration of 16.8 g/L and an inlet flow rate of 6.0 mL/min
239 (Figure 6). The breakthrough time increased as the inlet concentration decreased, which
240 may be attributed to the increased viscosity of oil at higher initial concentrations,
241 making it easier for the filler to become saturated and adsorbed³². In addition,
242 contaminated water diffuses rapidly through the adsorption saturation section and
243 partial saturation section, reaching the non-adsorption section faster and reducing the
244 overall adsorption time. The results of our study demonstrate that the breakthrough
245 point for diesel spills occurs earlier at higher concentrations.



246

247 Figure 6 Breakthrough curve at different inlet flow rates and different inlet diesel concentration. (a:4.2g/L;

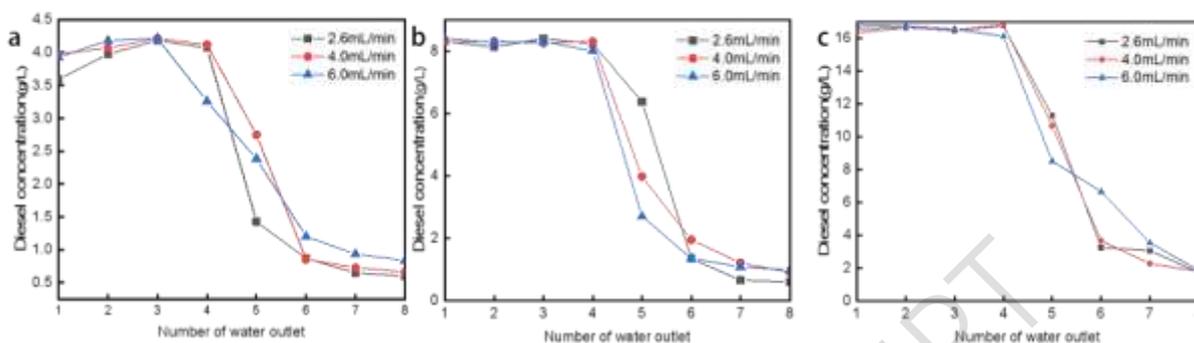
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b:8.4g/L; c:16.8g/L.)

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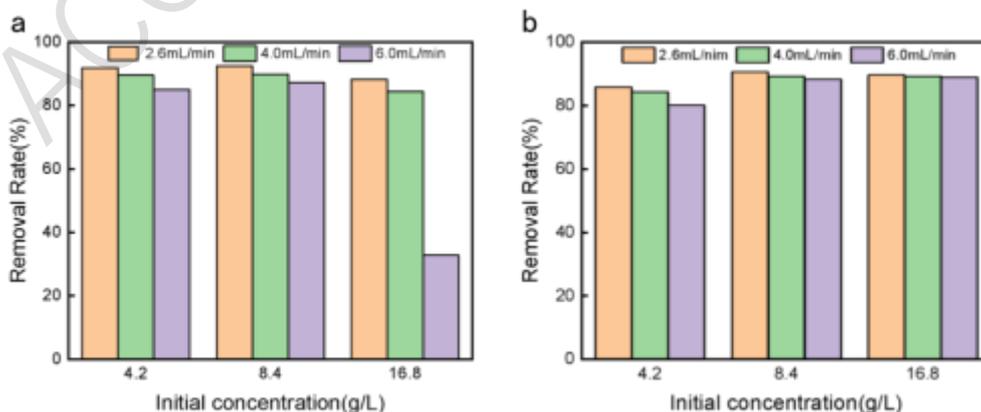
250 At 1 / 2 T, the diesel concentrations of outlet 1 ~ 4 were closed to that of inlet
251 diesel, and as the distance from the outlet decreased, the concentration of diesel oil at
252 the outlet approached 0. This is consistent with the premise that contaminated water is
253 adsorbed and saturated in the reactor progressively (Figure 7). When the initial
254 concentration of diesel was 4.2 g/L, the effluent concentration of outlets 1 ~ 4 was lower
255 than the inlet concentration of diesel, indicating that when the concentration is low, the

256 modified *E. prolifer*a fixed fossil oil degrading bacteria filled in the reactor have begun
257 internal absorption or biodegradation after adsorption saturation.

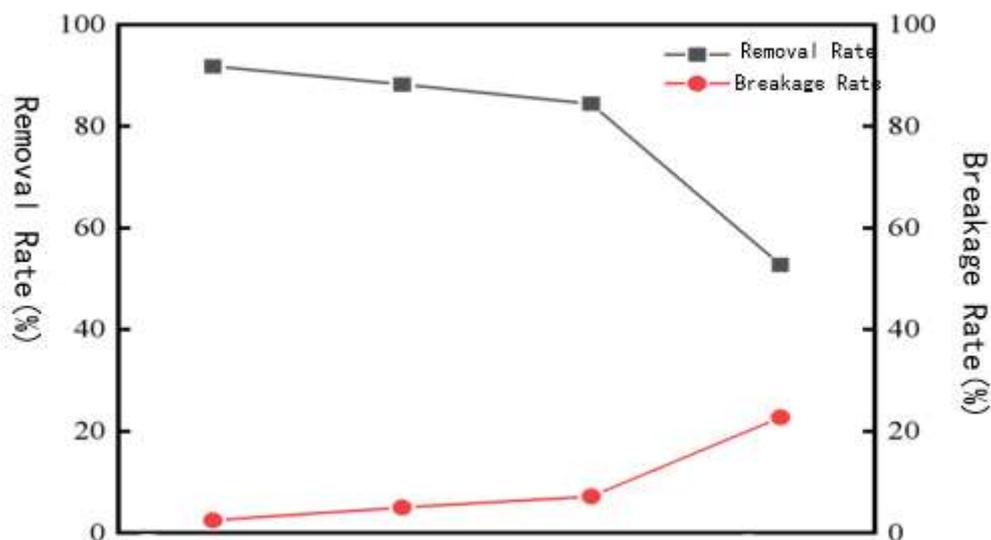


258
259 Figure 7 Diesel concentration at each outlet at 1/2 T under different initial concentrations and flow rates. (a:4.2g/L;
260 b:8.4g/L; c:16.8g/L.)

261 When the inlet flow rate was 6.0mL/min and the concentration of diesel was 16.8g/L,
262 the removal rate was decreased obviously and approached the breakthrough point
263 before 10.5h (Figure 8a). However, when the initial concentration was kept constant,
264 higher diesel removal rates were achieved at slower flow rates. There was little
265 difference ($P > 0.05$) in diesel removal rate under different initial concentration of rate
266 and flow rate (Figure 8). This showed that the flow rate and concentration of diesel oil
267 will not have much influence on the removal rate when the diesel oil flow is not
268 completely penetrated. When the inlet flow rate was not greater than 6.0mL/min and
269 the concentration of diesel was less than 16.8g/L, the removal rate of diesel was
270 guaranteed within 10.5h.



271
272 Figure 8 Diesel removal rate at 10 h(a) and 1/2 T(b) under different initial concentrations and flow rates



274

275 Figure 9. Changes in the diesel removal rate and fragmentation rate of modified *Enteromorpha*-immobilized
 276 bacterial agent under different utilization times

277

278 During multiple recycling processes, the removal rate of diesel oil decreased
 279 gradually with the increase of reuse times of immobilized bacteria, and the
 280 fragmentation rate of immobilized fossil oil degrading bacteria of modified *E. prolifera*
 281 increased gradually (Figure 9, Table S4). For the first three utilization cycles, no
 282 significant changes in diesel removal and fragmentation rates were observed ($P > 0.05$),
 283 while after the fourth cycle, the diesel removal rate decreased significantly and the
 284 crushing rate increased significantly ($P < 0.05$). Therefore, the optimum recycling times
 285 of filler in the reactor with modified *Enteromorpha*-immobilized bacterial agent as
 286 filling material was 3 times.

287 In the recyclability experiments, the main reasons for the increase of fragmentation
 288 rate of immobilized bacteria and the decrease of diesel removal rate might be: after a
 289 long time of immersion and scouring, the embedded materials fall off and the
 290 mechanical strength decreases continuously. In addition, washing of the normal saline
 291 may cause the falling off of embedded materials and the loss of internal oil degrading

292 bacteria(Ye et al., 2021).

293 **4. Conclusion**

294 In this study, a new type of immobilized cells, free cells combined with sodium
295 alginate and modified *E. prolifera* that could degrade diesel, was studied. The optimal
296 conditions for the preparation of modified *Enteromorpha*-immobilized bacterial agent
297 were obtained by designing response surface experiment and the highest removal
298 degraded rate reached 90.39% in 5 days When the inlet flow rate was not greater than
299 6.0mL/min and the concentration of diesel was less than 16.8g/L, the removal rate of
300 diesel was guaranteed within 10.5h. The modified *Enteromorpha*-immobilized bacterial
301 agent exhibited optimal recycling potential, with a recommended reuse limit of three
302 times in the reactor. These findings demonstrate the feasibility of using immobilized
303 cells for the treatment of oil pollution.

304 **Declarations**

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313 **Competing Interest**

314 The authors declare that they have no known competing financial interests or personal
315 relationships that could have appeared to influence the work reported in this paper.

316 **Author contribution**

317 Jinxiao Wei: Investigation, formal analysis, writing-original draft, visualization.
318 Yuping Yang: Funding acquisition, supervision, conceptualization, methodology,
319 resources, writing—review and editing. Yanlu Qiao: Formal analysis, writing-review.
320 Wenhui Xu: Analysis, visualization. All the authors contributed to critically reading of
321 the paper and approved the final manuscript.

322 **Data Availability**

323 All data and code generated or used during the study appear in the submitted article.

324 **References**

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